

Sequencing alignment

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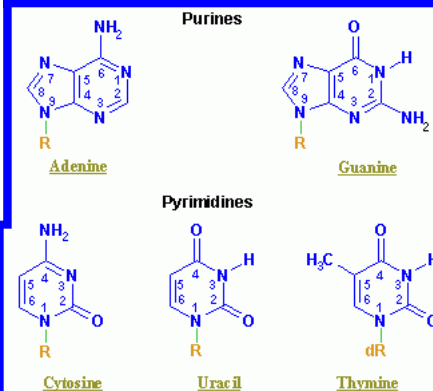
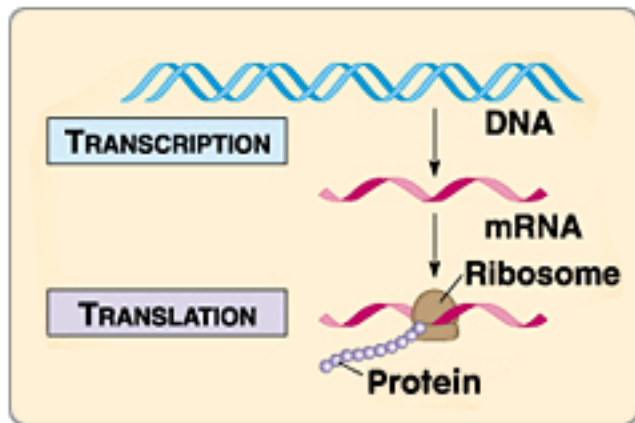
Why perform a multiple sequence alignment?

MSAs are at the heart of comparative genomics studies which seek to study evolutionary histories, functional and structural aspects of sequences, and to understand phenotypic differences between species

The Building Blocks...

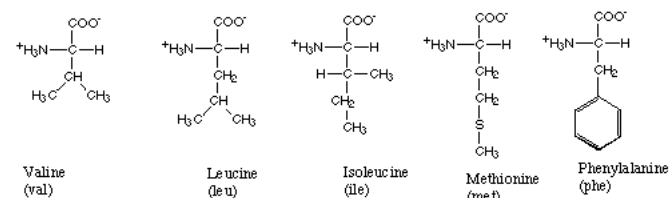
Schematic of protein synthesis

(© 1999, Addison Wesley Longman Inc.)

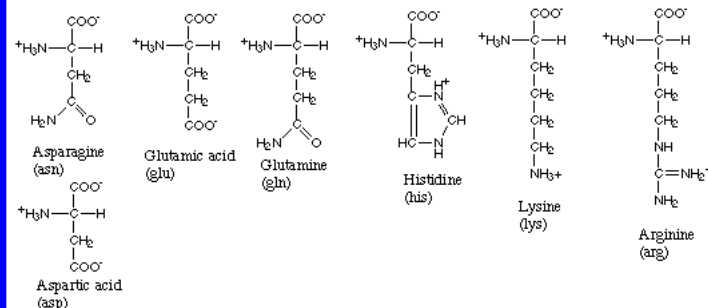


ATGC

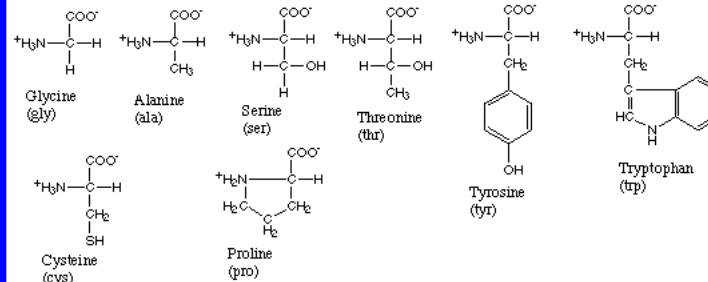
Amino acids with hydrophobic side groups



Amino acids with hydrophilic side groups



Amino acids that are in between



VLMFNQEDHKRCSTPYW

```

MVNLT SDEKTAVLALWNKVDVEDCGGEALGRLLVVYPWTQRFFE...
||  ||  ||||| ||| ||  ||  ||||| ||||| ||||| ||||| |||||
MVHLTPEEKTAVNALWGKVNVDVGGEALGRLLVVYPWTQRFFE...
  
```

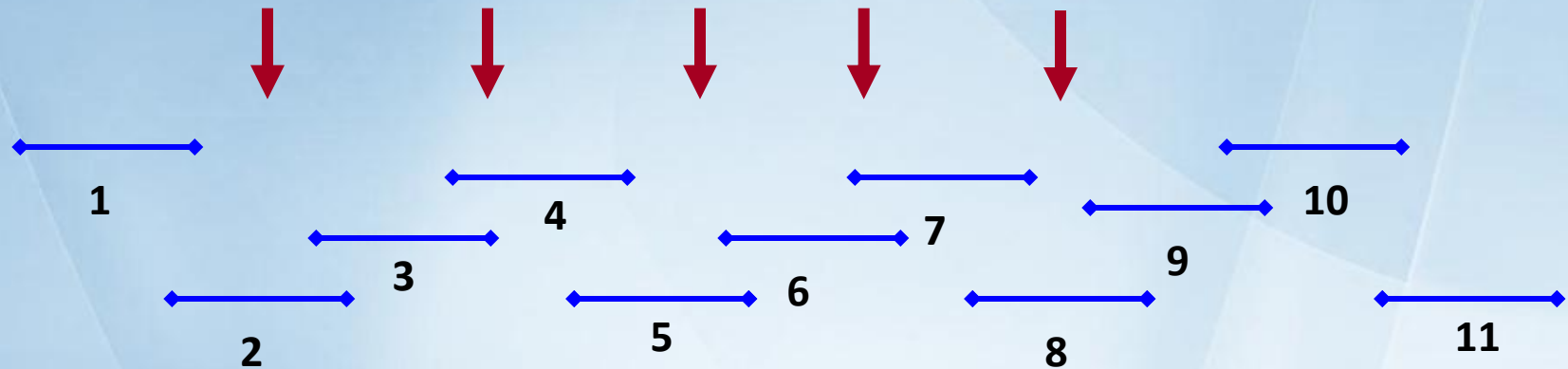
Alignment: Comparing two (pairwise) or more (multiple) sequences. Searching for a series of identical or similar characters in the sequences.

Why Align Sequences?

- Discover functional, structural, and evolutionary information
- Similar Sequences may have similar function
 - Gene Regulation
 - Biochemical Function
 - Similar Structure
- Identify primers and probes to search for homologous sequences in other organisms
- Crucial for genome sequencing

Genome Sequencing Strategy

genome



1. Obtain a large collection of BAC clones
2. Map them onto the genome (Physical Mapping)
3. Select a minimum tiling path
4. Sequence each clone in the path with shotgun
5. Assemble
6. Put everything together

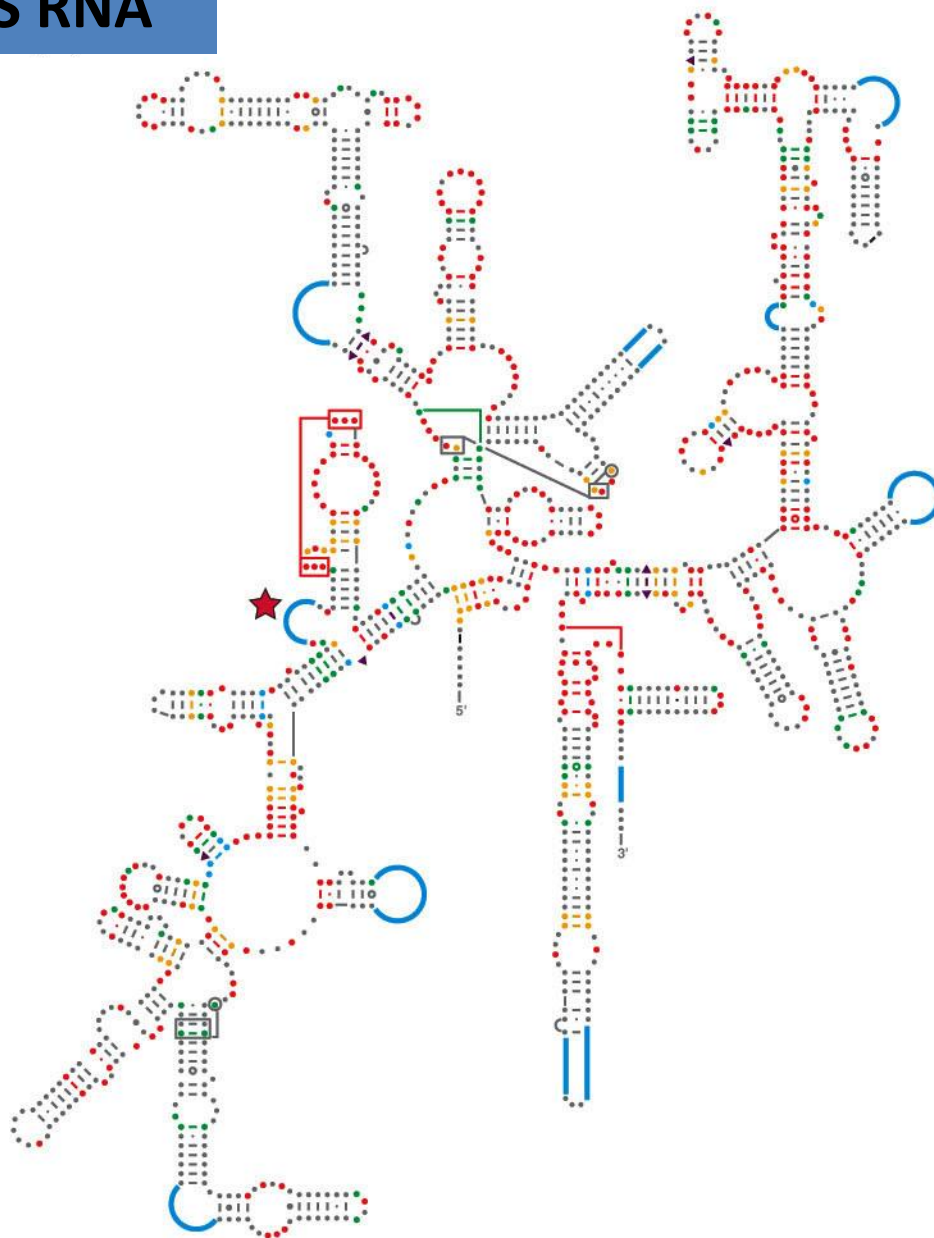
Why Align Sequences?



- Homology
 - Similar sequences may have a common ancestor
- In 1990 Carl Woese and colleagues proposed the Tree of Life, using a molecular phylogenetic approach.
- It is based on sequencing rRNA (16S and 18S), which all organisms share.
- All organisms were separated into three domains: *Bacteria*, *Archaea*, *Eukarya*.



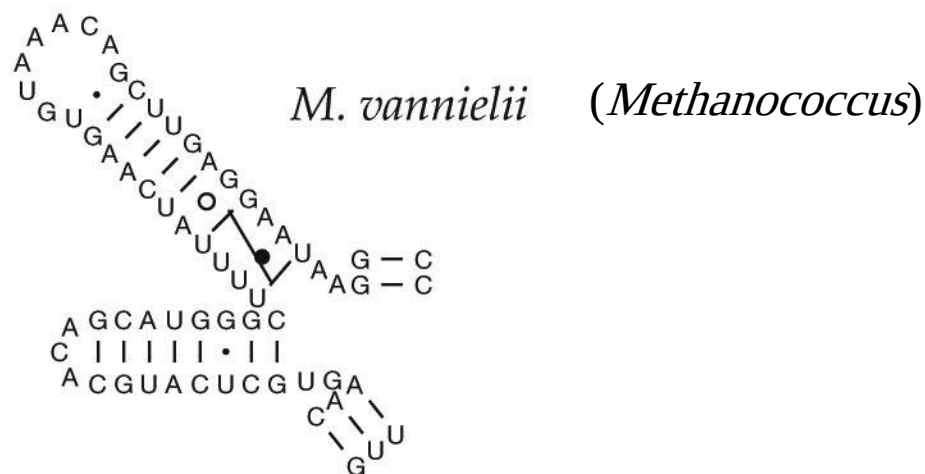
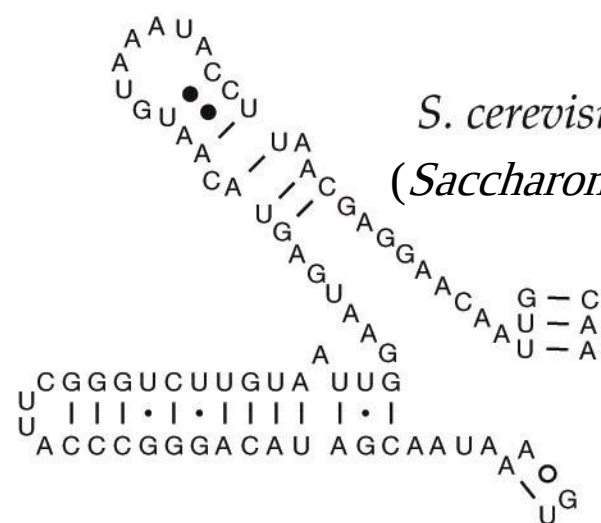
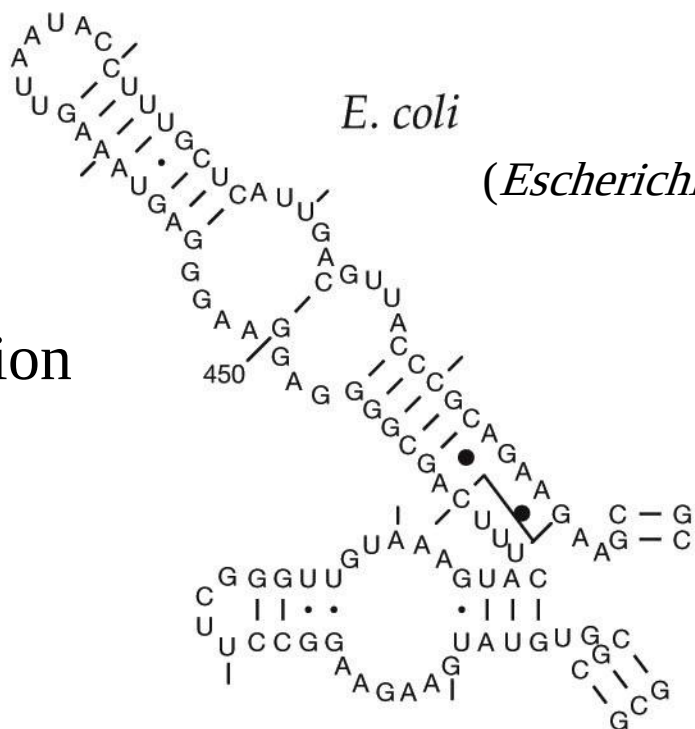
16S RNA

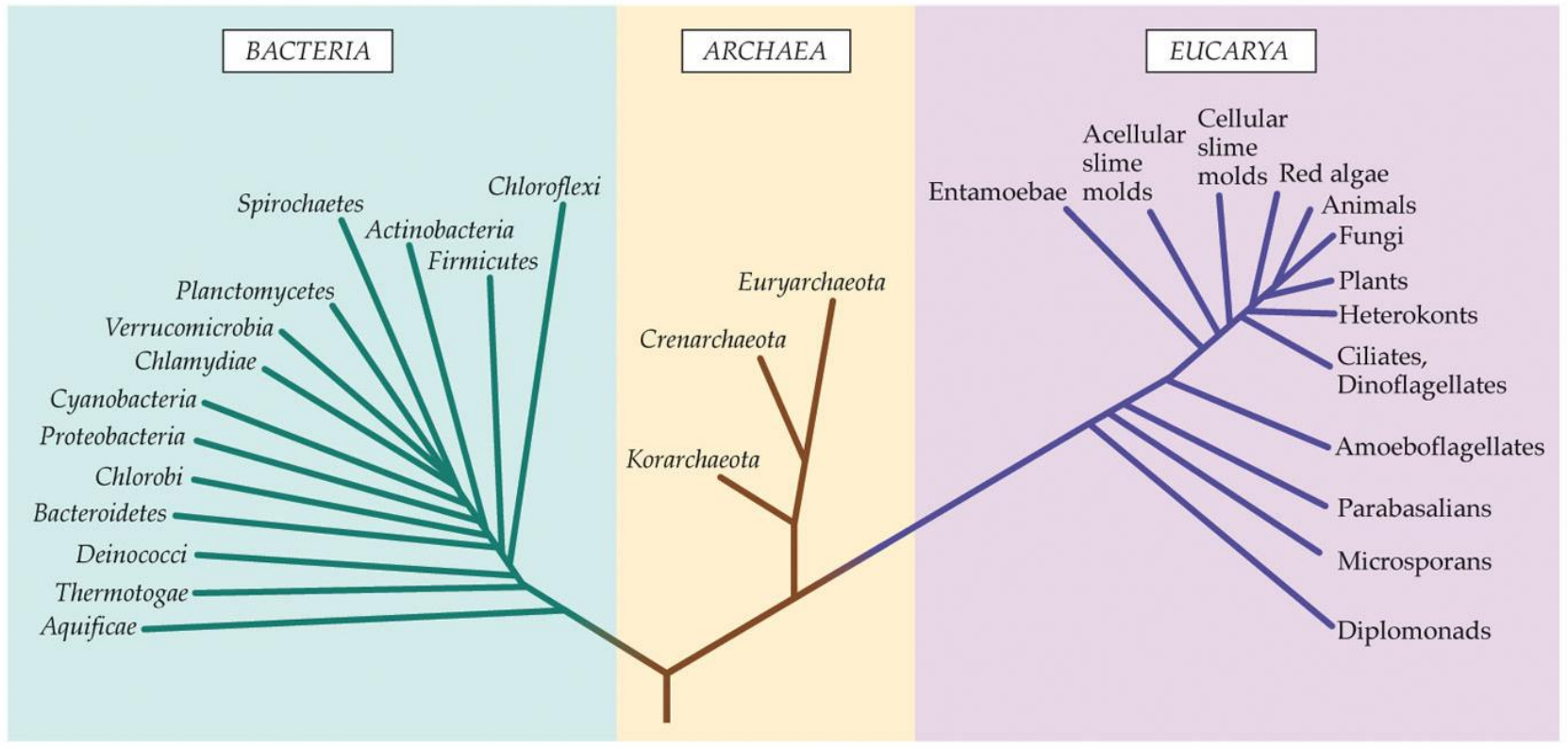


- Identical in 98% or more of all organisms
- Conserved only in the *Bacteria*
- Conserved only in the *Archaea*
- Conserved only in the *Eukarya*
- ▲ Conserved within each domain, variable among domains
- Regions that vary structurally among domains

S. cerevisiae
(*Saccharomyces*)

 Region





Evolutionary changes in sequences

Three types of nucleotide changes:

1. **Substitution** – a replacement of one (or more) sequence characters by another: **AAGA** → **AACA**
2. **Insertion** - an insertion of one (or more) sequence characters: **AAG ~~A~~**
3. **Deletion** – a deletion of one (or more) sequence characters: **~~A~~AGA**

Insertion + Deletion → Indel

How DNA Sequence Data is Compared

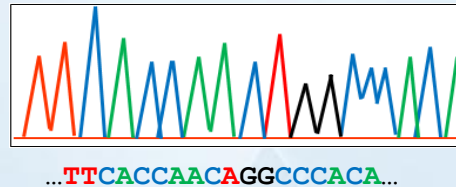
Obtain Samples:
Blood, Saliva, Hair
Follicles, Feathers, Scales



Genetic Data

Extract DNA from Cells

Sequence DNA



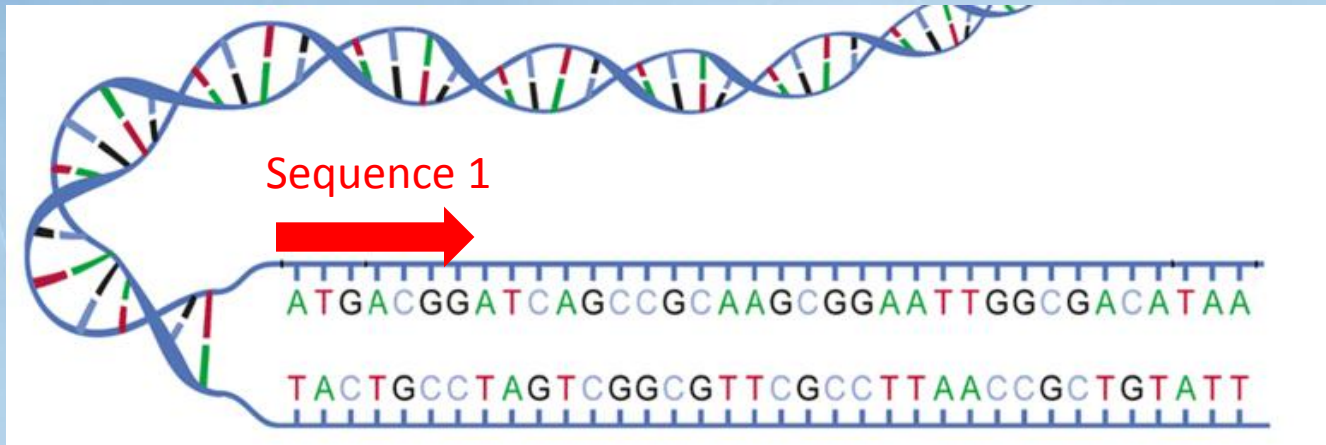
**Compare
DNA
Sequences to
One Another**

TTCAACAACAGGCCAC
TTCACCAACAGGCCAC
TTCATCAACAGGCCAC

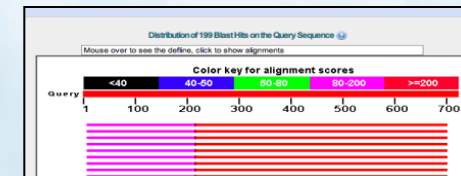
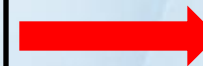
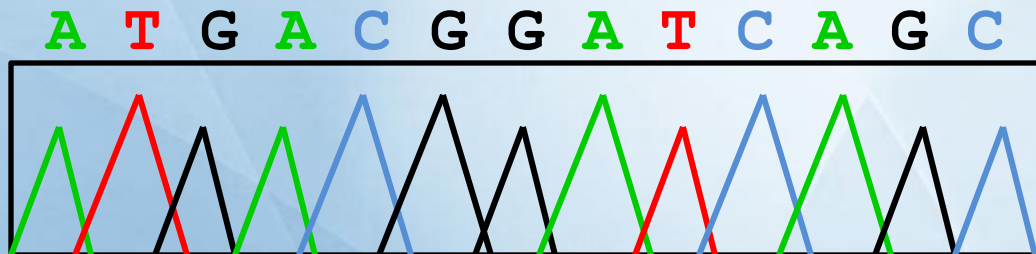
GOALS:

- Identify the organism from which the DNA was obtained.
- Compare DNA sequences to each other.

Sequence Both Strands of DNA



Sequence :



Bioinformatics tools like **BLAST** can be used to compare the sequences from Different individuals.

Comparing two sequences

MV**N**LT**S**DEKTAV**L**ALWNK**V**D**V**ED**C**G**G**E
 || || || || || || || || || || || ||
 MV**H**LT**P**EEKTAV**N**ALWGKV**N**VD**A**V**G**G**E**



Multiple sequence alignment

```
Seq1 VTISCTGSSSNIGAG-NHVKWYQQLPG
Seq2 VTISCTGTSSNIGS--ITVNWYQQLPG
Seq3 LRLSCSSSGFIFSS--YAMYWVRQAPG
Seq4 LSLTCTVSGTSFDD--YYSTWVRQPPG
Seq5 PEVTCVVVDVSHEDPQVKFNWYVDG--
Seq6 ATLVCLISDFYPGA--VTVAWKADS--
Seq7 AALGCLVKDYFPEP--VTVSWNSG---
Seq8 VSLTCLVKGFYPSD--IAVEWWSNG--
```

Similar to pairwise alignment BUT ***n*** sequences are aligned instead of just **2**

Each row represents an individual sequence
Each column represents the 'same' position

Multiple sequence alignment

Seq1 VTIS**C**TGSSSNI**G**AG-NHV**K**WY**Q**L**P**G
Seq2 VTIS**C**GTSSNI**G**S--ITV**N**WY**Q**L**P**G
Seq3 LRLS**C**SSSGFIF**S**--YAM**Y**WVR**Q**A**P**G
Seq4 LSLT**C**TVSGT**S**F**D**D--YYS**T**WVR**Q**P**P**G
Seq5 PEVT**C**VVVVDVSH**E**D**P**QVKF**N**WYV**D**G--
Seq6 ATLV**C**LISDFYP**G**A--VTV**A**W**K**A**D**S--
Seq7 AALG**C**LVKDYF**P**E**P**--VTV**S**W**N**S**G**--
Seq8 VSLT**C**LVKGFYP**S**D--IAV**E**WWS**N**G--



variable conserved

Multiple Sequence Alignment: Approaches

- **Local alignment** – finds regions of high similarity in **parts** of the sequences

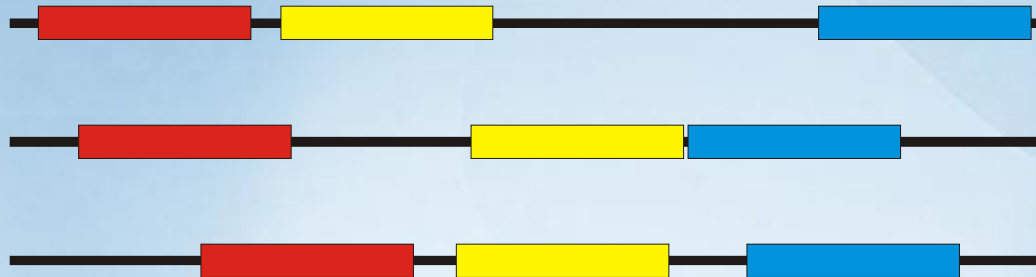
ADLGAVFAL	CDRYFQ
ADLGRTQN	CDRYYQ

- **Global alignment** – finds the best alignment across the **entire** two sequences

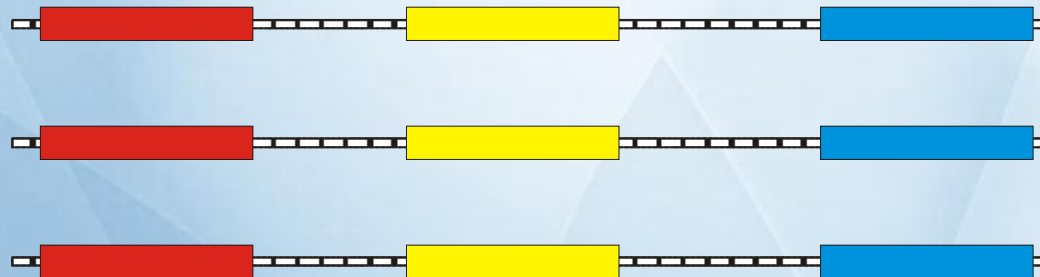
ADLGAVFAL	CDRYFQ
ADLGR	TQN-CDRYYQ

How to optimize alignment algorithms?

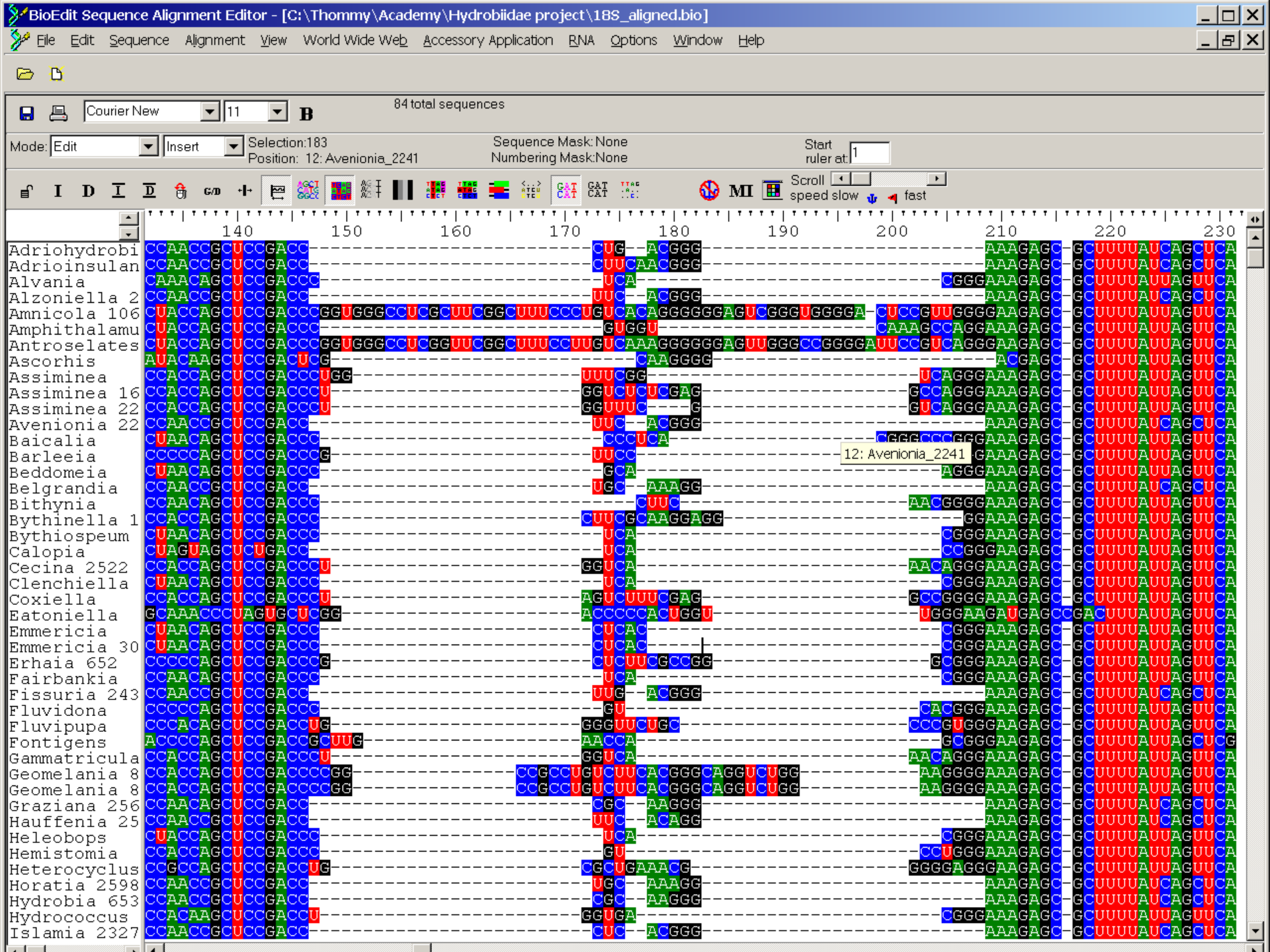
- Sequences often contain highly conserved regions



These regions can be used for an initial alignment



By analyzing a number of small, independent fragments, the algorithmic complexity can be drastically reduced!



Choosing an alignment:

- Many **different** alignments between two sequences are possible:

AAGCTGAATTCGAA
AGGCTCATTCTGA

AAGCGAAATTCGAAC
A-G-GAA-CTCGAAC

AAGCGAAATTCGAAC
AGG---AACTCGAAC

How do we determine which is the best alignment?

Assessing the significance of an alignment score

True

AAGCTGAATTCGAA
AGGCTCATTCTGA

AAGCTGAATTC-GAA
AGGCTCATTCTGA-

28.0

Random

AGATCAGTAGACTA
GAGTAGCTATCTCT

AGATCAGTAGACTA-----
-----GAGTAG-CTATCTCT

26.0

•
•

CGATAGATAGCATA
GCATGTCATGATTC

CGATAGATAGCATA-----
-----GCATGTCATGATTC

16.0

“Optimal” vs. “correct” alignment

For a given group of sequences, there is no single “correct” alignment, only an alignment that is “optimal” according to some set of calculations

This is partly due to:

- the complexity of the problem,
- limitations of the scoring systems used,
- our limited understanding of life and evolution

Determining what alignment is best for a given set of sequences is really up to the judgment of the investigator

Success of the alignment will depend on the similarity of the sequences. If sequence variation is great it will be very difficult to find an optimal alignment

Web servers for pairwise alignment

NCBI Home

Resource List (A-Z)

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NCBI Announcements

Coffee Break tutorial: Brown fat and obesity

Apr 1, 2014

The latest Coffee Break tutorial discusses EUMT1 as a...

New NCBI YouTube video: Create custom databases for BLAST

Mar 28, 2014

In the newest NCBI video on YouTube, we show you how to create custom...

Basic Local Alignment Search Tool



BLAST Basic Local Alignment Search Tool

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NCBI/ BLAST/ blastn suite

blastn blastp blastx tblastn tblastx

Enter Query Sequence

Enter accession number, gi, or FASTA sequence [Clear](#) Query subrange

From

To

Or, upload file [Browse...](#)

Job Title

Enter a descriptive title for your BLAST search

☒ Align two or more sequences

Enter Subject Sequence

Enter accession number, gi, or FASTA sequence [Clear](#) Subject subrange

From

To

Or, upload file [Browse...](#)

Program Selection

Optimize for

- ☒ Highly similar sequences (megablast)
- ☐ More dissimilar sequences (discontiguous megablast)
- ☐ Somewhat similar sequences (blastn)

Choose a BLAST algorithm

BLAST Search nucleotide sequence using Megablast (Optimize for highly similar sequences)

☐ Show results in a new window

Algorithm parameters

blastn –
nucleotide
blastp – protein

BLAST – programs



BLAST

Basic Local Alignment Search Tool

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BLAST finds regions of similarity between biological sequences. [more...](#)

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BLAST Assembled Genomes

Choose a species genome to search, or [list all genomic BLAST databases](#).

- ☐ [Human](#)
- ☐ [Mouse](#)
- ☐ [Rat](#)
- ☐ [Arabidopsis thaliana](#)

- ☐ [Oryza sativa](#)
- ☐ [Bos taurus](#)
- ☐ [Danio rerio](#)
- ☐ [Drosophila melanogaster](#)

- ☐ [Gallus gallus](#)
- ☐ [Pan troglodytes](#)
- ☐ [Microbes](#)
- ☐ [Apis mellifera](#)

Basic BLAST

Choose a BLAST program to run.

[nucleotide blast](#)

Search a **nucleotide** database using a **nucleotide** query
Algorithms: blastn, megablast, discontinuous megablast

[protein blast](#)

Search **protein** database using a **protein** query
Algorithms: blastp, psi-blast, phi-blast

[blastx](#)

Search **protein** database using a **translated nucleotide** query

[tblastn](#)

Search **translated nucleotide** database using a **protein** query

[tblastx](#)

Search **translated nucleotide** database using a **translated nucleotide** query

Specialized BLAST

Choose a type of specialized search (or database name in parentheses.)

- ☐ Make specific primers with [Primer-BLAST](#)
- ☐ Search [trace archives](#)
- ☐ Find [conserved domains](#) in your sequence (cds)
- ☐ Find sequences with similar [conserved domain architecture](#) (cdart)
- ☐ Search sequences that have [gene expression profiles](#) (GEO)
- ☐ Search [immunoglobulins](#) (IgBLAST)
- ☐ Search for [SNPs](#) (snp)
- ☐ Screen sequence for [vector contamination](#) (vecscreen)
- ☐ [Align](#) two (or more) sequences using BLAST (bl2seq)
- ☐ Search [protein](#) or [nucleotide](#) targets in PubChem BioAssay
- ☐ Search SRA [transcript libraries](#)
- ☐ Constraint Based Protein [Multiple Alignment Tool](#)

BLAST – bl2seq



BLAST
 Basic Local Alignment Search Tool

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☐ [Gallus gallus](#)
☐ [Pan troglodytes](#)
☐ [Microbes](#)
☐ [Apis mellifera](#)

Basic BLAST

Choose a BLAST program to run.

nucleotide blast	Search a nucleotide database using a nucleotide query <i>Algorithms:</i> blastn, megablast, discontinuous megablast
protein blast	Search protein database using a protein query <i>Algorithms:</i> blastp, psi-blast, phi-blast
blastx	Search protein database using a translated nucleotide query
tblastn	Search translated nucleotide database using a protein query
tblastx	Search translated nucleotide database using a translated nucleotide query

Specialized BLAST

Choose a type of specialized search (or database name in parentheses.)

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- ☐ Search SRA [transcript libraries](#)
- ☐ Constraint Based Protein [Multiple Alignment Tool](#)

Sequence alignment - Wik x Multiple Sequence Alignm x COBALT:Multiple Alignm x

www.ncbi.nlm.nih.gov/tools/cobalt/cobalt.cgi?link_loc=BlastHomeLink

COBALT *Constraint-based Multiple Alignment Tool*

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Cobalt Constraint-based Multiple Protein Alignment Tool

COBALT computes a multiple protein sequence alignment using conserved domain and local sequence similarity information. [Reset page](#)

Enter Query Sequences

Enter at least 2 protein accessions, gis, or FASTA sequences [Clear](#)

Or, upload FASTA file No file chosen

Job Title

☐ Show results in a new window

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Bl2seq results

▼ **Alignments** ☐ Select All [Get selected sequences](#) **NEW**

```
>lc1143385 gi150902697|gb|AAT86412.1| ABC transporter-associated protein
[Streptococcus pyogenes MGAS10394]
Length=472

Score = 394 bits (1013), Expect = 5e-114, Method: Compositional matrix adjust
Identities = 190/304 (62%), Positives = 232/304 (76%), Gaps = 4/304 (1%)
```

Query	566	PTYDIQVV	GLENFVANGIVAHNSFIYVPPGVHVDIPLQAYFRINTENMGQFERTLIIADT	625
		P D ++	L + V +G +FIYVP GV VDIPLQ YFRIN EN GQFERTLII D	
Sbjct	173	PPTDNKLA	ALNSAVWSG---GTFIYVPGKGVKVDIPLQTYFRINNENTGQFERTLIIIVDE	228
Query	626	GSYVHVE	ECTAPIYKSDSLASAVVEIIVKPHARVRYTTIQNWSNNVYNLVTKRARVETG	685
		G+ V FVEG	TAP Y S+SLH+A+VEI A +RYTTIQNWS+NVYNLVTKRAR T	
Sbjct	229	GASVHYVE	CTAPTYSSNSLHAATVEIFALDGAYMRYTTIQNWSDNVYNLVTKRARALTD	288
Query	686	ATNEWIDG	NIGSKVTMKYPAVWMTGAKGEVLSVAFAGEGQHQDTGAKMLHLASNTSSN	745
		ATNEWIDGN	+G+K TMKYP+V++ G R+G +LS+AFA GQHQDTGAKM+H A +TSS+	
Sbjct	289	ATNEWIDGN	LGAKTTMKYPSVYLDGPGGRTMLSIAFANAGQHQDTGAKMIHNAPHTSSS	348
Query	746	IVSKSVARG	SGRYSYRGLVQVNGKAHGSRSYKCDALLVDTISRSDTYPYVDIREDDVTM	805
		IVSKS+A+	GG+ YRG V NK + S S +CD +L+D IS+SDT P+ +I V +	
Sbjct	349	IVSKSIAKS	GGKVQYRGQVTFNKQSKSVSHESDITILMDDISKSDTIPFNEIHNSQVAL	408
Query	806	GHEATVSKV	SENQLFYLMRSGLAEDEAMAMVVRGFEPIAKELPMEYALELNRLIELQME	865
		HEA VSK+SE	QL+YLMRGL+E EA M+V GVEP KELPMEYA+ELNRLI +ME	
Sbjct	409	EHEAKVSKI	SEEQLYYLMRGLSESEATEMIVMGFEPEPKELPMEYAVELNRLISYEME	468
Query	866	QAVG	869	
		Q+VG		
Sbjct	469	ESVG	472	

Match

Similarity

Dissimilarity

Gaps

MSA input: multiple sequence Fasta file

>gi|4504351|ref|NP_000510.1| delta globin [Homo sapiens]
MVHLTPEEKTAVNALWGKVNVDVGGGEALGRLLVVYPWTQRFFESFGDLSSPDVAMGNPKVKAHGKKVLG
AFSDGLAHLNLTGTFSQLSELHCDKLHVDPENFRLLGNVLCVLARNFGKEFTPQMQAAYQKVAVAGVAN
ALAHKYH

>gi|4504349|ref|NP_000509.1| beta globin [Homo sapiens]
MVHLTPEEKSAVTALWGKVNVDVGGGEALGRLLVVYPWTQRFFESFGDLSTPDVAMGNPKVKAHGKKVLG
AFSDGLAHLNLTGTFATLSELHCDKLHVDPENFRLLGNVLCVLAHHFGKEFTPPVQAAYQKVAVAGVAN
ALAHKYH

>gi|4885393|ref|NP_005321.1| epsilon globin [Homo sapiens]
MVHFTAEEKAAVTSLSKMNVEEAGGEALGRLLVVYPWTQRFFDSFGNLSSPSAILGNPKVKAHGKKVLT
SFGDAIKNMDNLKPAFAKLSELHCDKLHVDPENFKLLGNVMVILATHFGKEFTPEVQAAWQKLVSVAI
ALAHKYH

>gi|6715607|ref|NP_000175.1| G-gamma globin [Homo sapiens]
MGHFTEEDKATITSLWGKVNVEDAGGETLGRLLVVYPWTQRFFDSFGNLSSASAIMGNPKVKAHGKKVLT
SLGDAIKHLDDLKGTFAQLSELHCDKLHVDPENFKLLGNVLTVLAIHFGKEFTPEVQASWQKMVTGVAS
ALSSRYH

>gi|28302131|ref|NP_000550.2| A-gamma globin [Homo sapiens]
MGHFTEEDKATITSLWGKVNVEDAGGETLGRLLVVYPWTQRFFDSFGNLSSASAIMGNPKVKAHGKKVLT
SLGDATKHLDDLKGTFAQLSELHCDKLHVDPENFKLLGNVLTVLAIHFGKEFTPEVQASWQKMVTAVAS
ALSSRYH

>gi|4885397|ref|NP_005323.1| hemoglobin, zeta [Homo sapiens]
MSLTKTERTIIVSMWAKISTQADTIGTETLERLFLSHPQTKTYFPHFDLHPGSAQLRAHGSKVVAAVGDA
VKSIDDIGGALSSELHAYILRVDPVNFLLSHCLLVTLAARFPADFTAEAAAWDKFLSVVSSVLTEK
YR

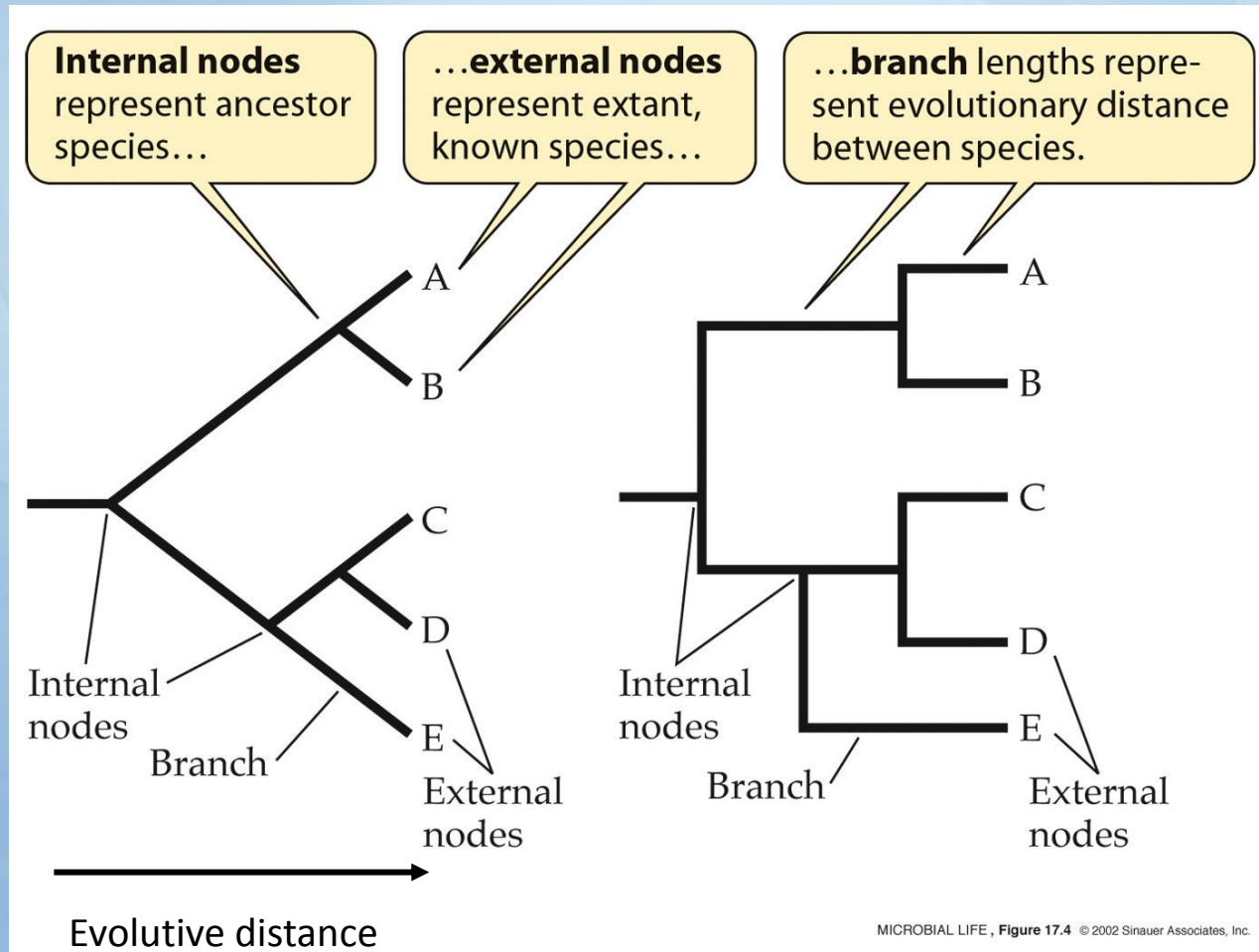
Query type: AA or DNA?

- For coding sequences, AA (protein) data are better
 - Selection operates most strongly at the protein level → the homology is more evident
 - AA – 20 char' alphabet DNA - 4 char' alphabet



lower chance of random homology for AA

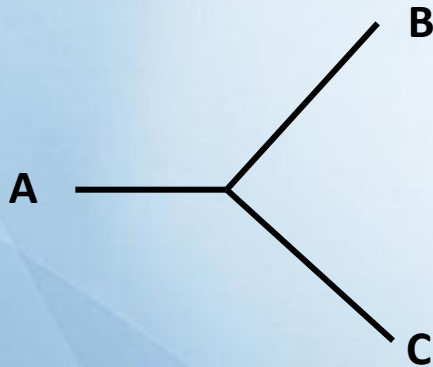
Phylogenetic trees



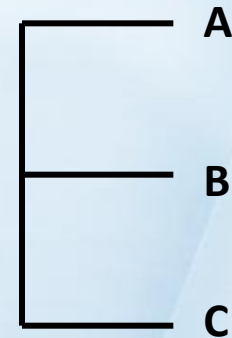
Phylogenetic trees

Rooted and unrooted trees

- **Unrooted trees:** compare one feature of a group of related organisms



Or



Only one shape for a tree with 3 species

Distance matrix: UPGMA



- UPGMA = unweighted pair group method with arithmetic mean

(A) UPGMA method

Table shows sequence of nine-base region of rRNAs of four strains.

Organism (strain)	Site number								
	1	2	3	4	5	6	7	8	9
a	G	C	G	G	A	C	A	A	A
b	G	A	C	G	C	C	A	A	G
c	G	A	A	A	U	C	U	A	A
d	G	A	A	A	G	C	U	A	G

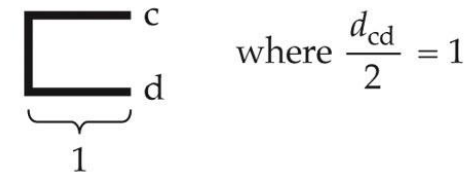
1 Construct matrix showing relatedness between strains.

First matrix

	a	b	c
b	$d_{ab} = 4$	—	—
c	$d_{ac} = 5$	$d_{bc} = 5$	—
d	$d_{ad} = 6$	$d_{bd} = 4$	$d_{cd} = 2$

2 Diagram relatedness between strains.

Beginning tree



Distance matrix: UPGMA

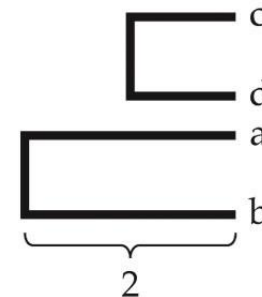
c & d are considered as one unit

Second matrix

	(cd)	a
a	$d_{(cd)a} = 11/2$	—
b	$d_{(cd)b} = 9/2$	$d_{ab} = 4$

3 Construct matrix to assess distance between (cd) and (a and b).

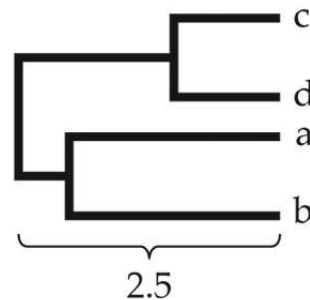
Second tree



4 Determine and diagram their relatedness.

where $\frac{d_{ab}}{2} = 2$

Final tree

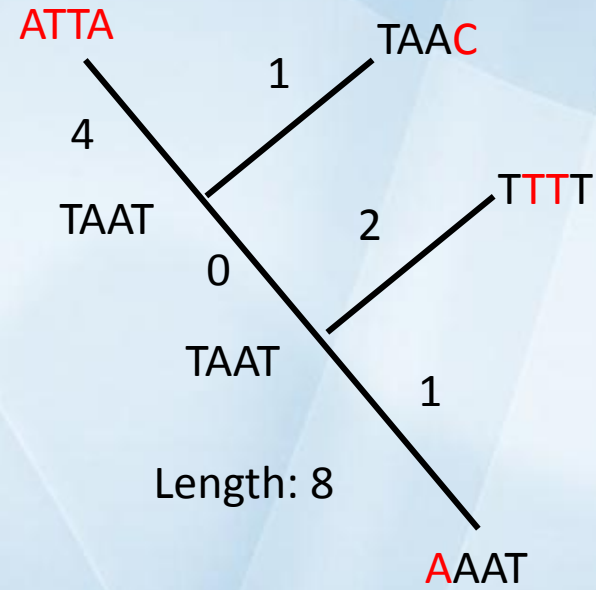
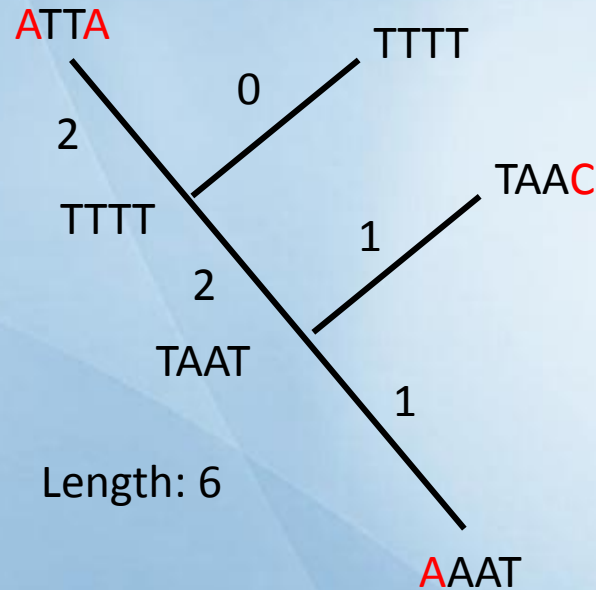


5 Determine relatedness between cd and a and b.

where $d_{(cd)(ab)} = \frac{(11/2 + 9/2)/2}{2} = 2.5$

Other method: maximum parsimony

Sequence 1: ATTA
Sequence 2: TAAC
Sequence 3: AAAT
Sequence 4: TTTT

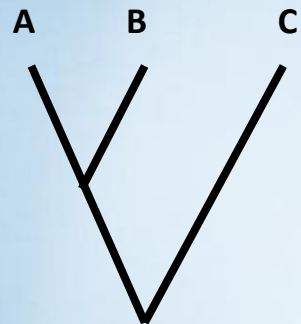


Most likely tree

The alignment problem

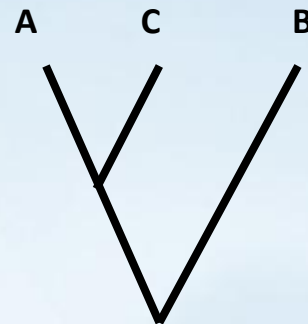


- ◆ What happens when a sequence alignment is wrong?

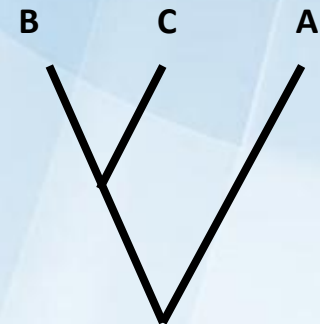


A: AGT
B: AT
C: ATC

A: AGT
B: A -T
C: ATC



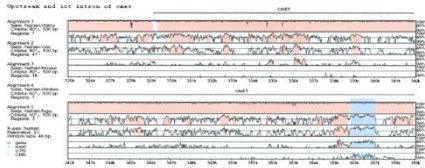
A: AGT -
B: A -T -
C: A -TC



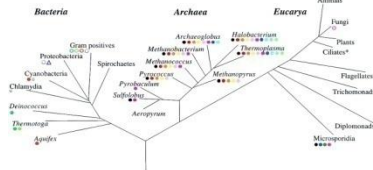
A: AGT
B: AT -
C: ATC

Central role of multiple alignments

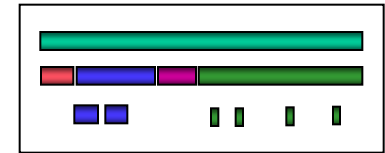
Comparative genomics



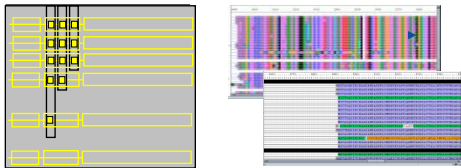
Phylogenetic studies



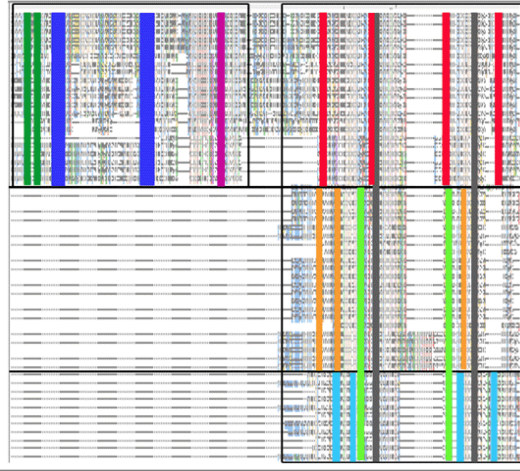
Hierarchical function annotation: homologs, domains, motifs



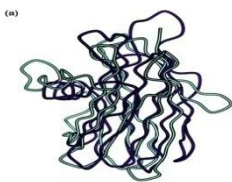
Gene identification, validation



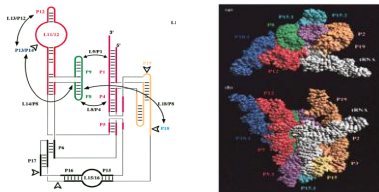
Multiple alignment



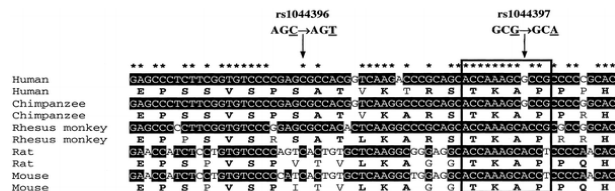
Structure comparison, modelling



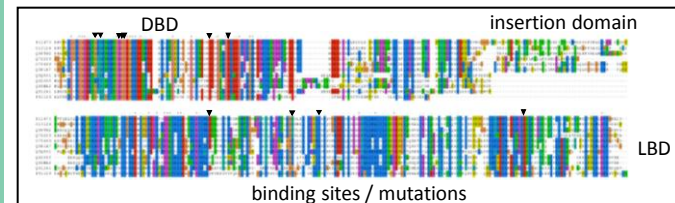
RNA sequence, structure, function



Human genetics, SNPs



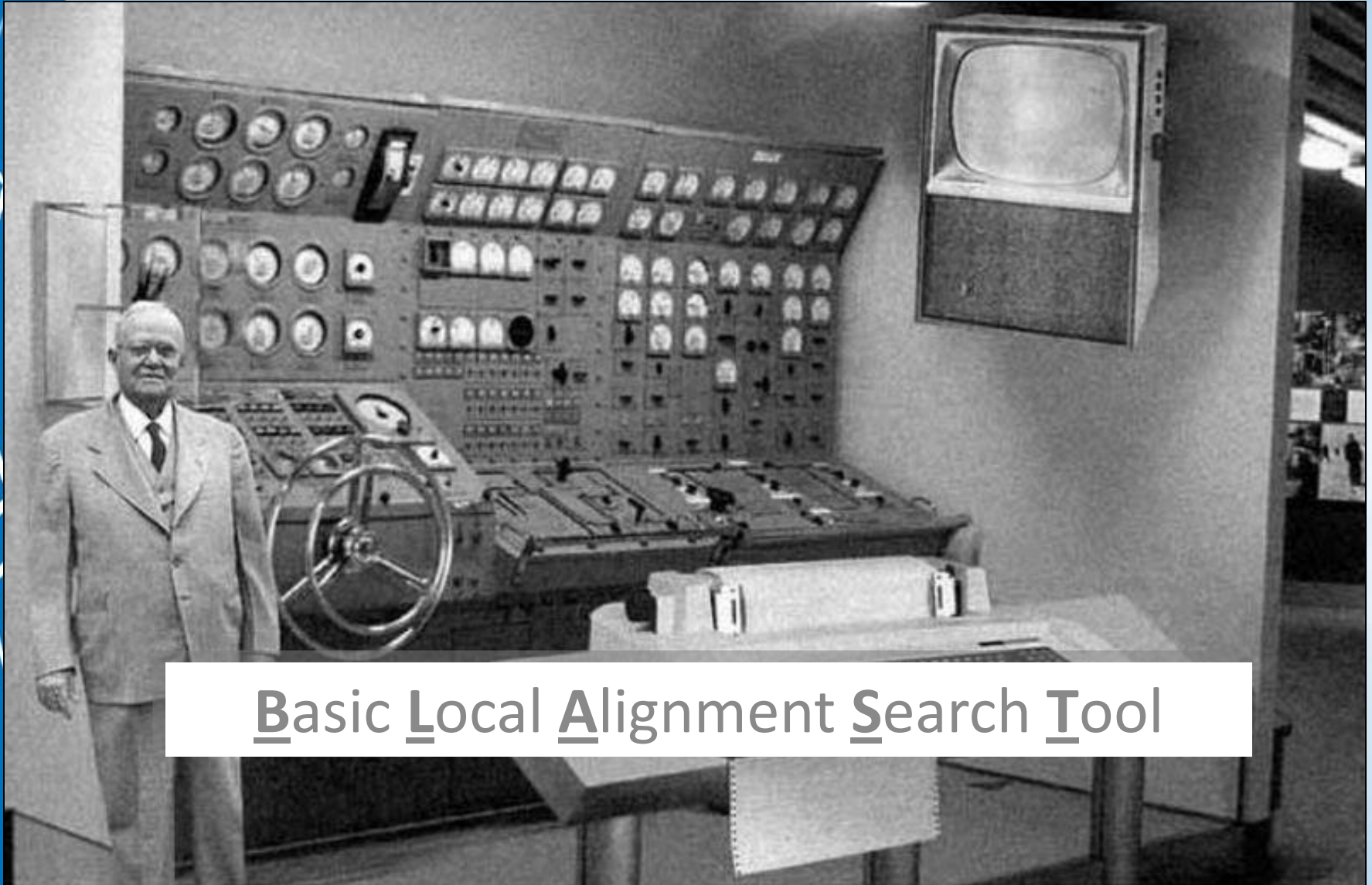
Therapeutics, drug design



Most important sequence databases

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- **Genbank** – maintained by USA National Center for Biology Information (NCBI)
 - All biological sequences
 - www.ncbi.nlm.nih.gov/Genbank/GenbankOverview.html
 - Genomes
 - www.ncbi.nlm.nih.gov:80/entrez/query.fcgi?db=Genome
 - **Swiss-Prot** – maintained by EMBL- European Bioinformatics Institute (EBI)
 - Protein sequences
 - www.ebi.ac.uk/swissprot/

Sequence Similarity Searching



Basic Local Alignment Search Tool

