

*Don't see everyone's flaws
Don't listen to everything you're told
Don't speak if it's not kind*



*Always look for the good in people.
Not everything is truth.
Only speak words of kindness*

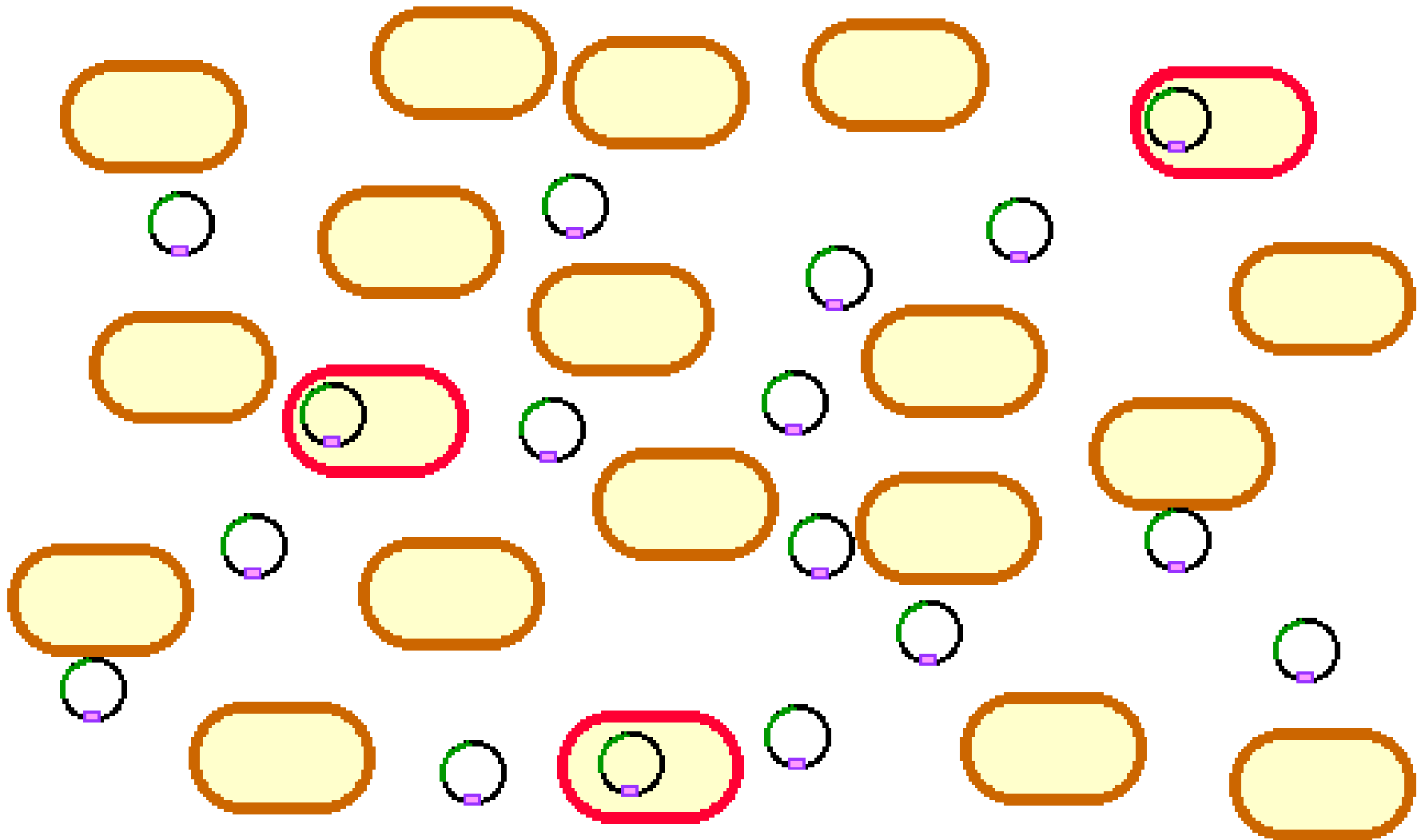
Live Life Positively

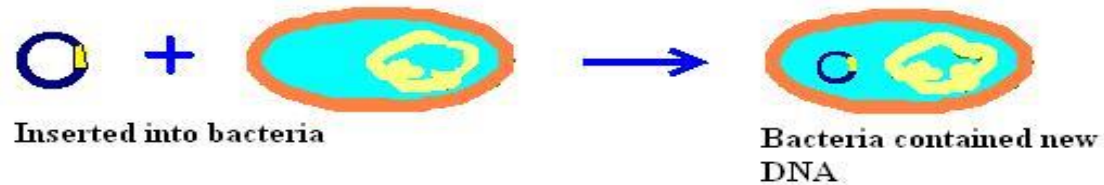
DETECTION OF YOUR CLONING AND PICK UP YOUR RIGHT CLONE

Introduction

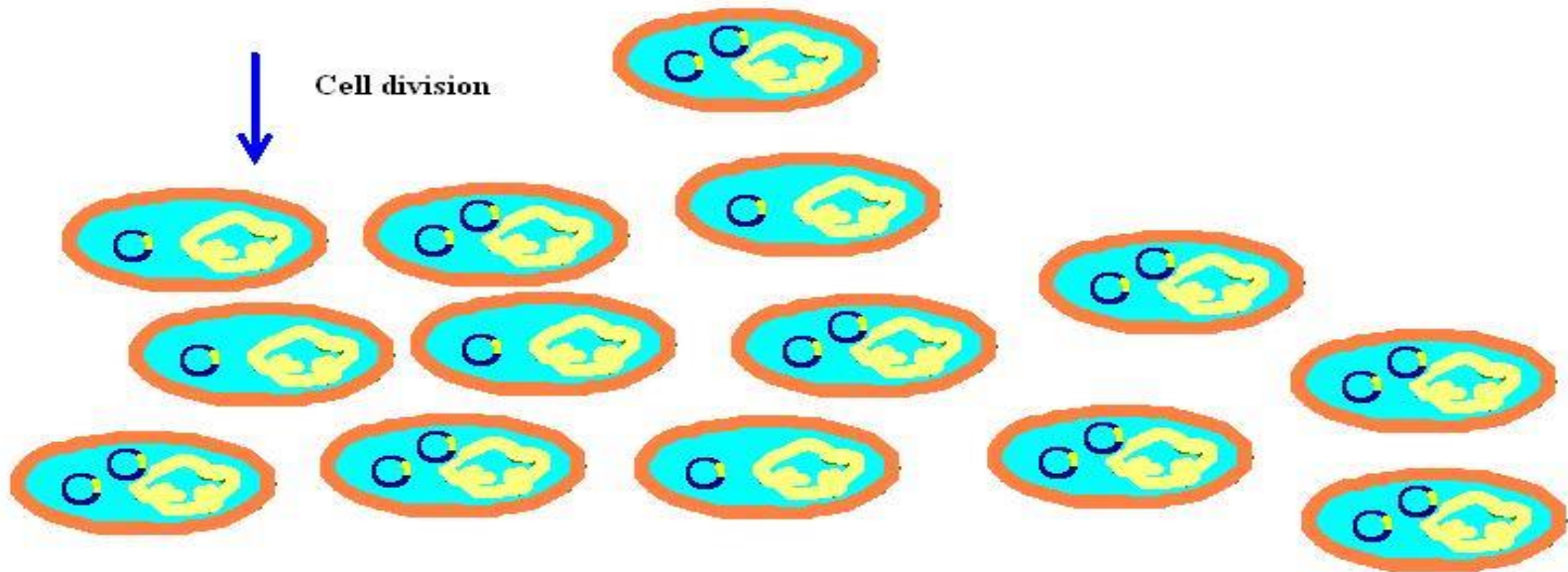
- Once recombinant plasmid is constructed, it is introduced into recipient cells.
- Introduction of recombinant DNA into recipient cells is called transformation: *introduction of foreign DNA changes (transforms) properties of the organism.*
- Special treatment makes *cells competent* - capable of accepting foreign DNA.
- Usually, these treatments make cell membrane more permeable for a DNA molecule.
- When competent cells are mixed with DNA some cells (actually, very few) become transformed.

Competent cells transformation



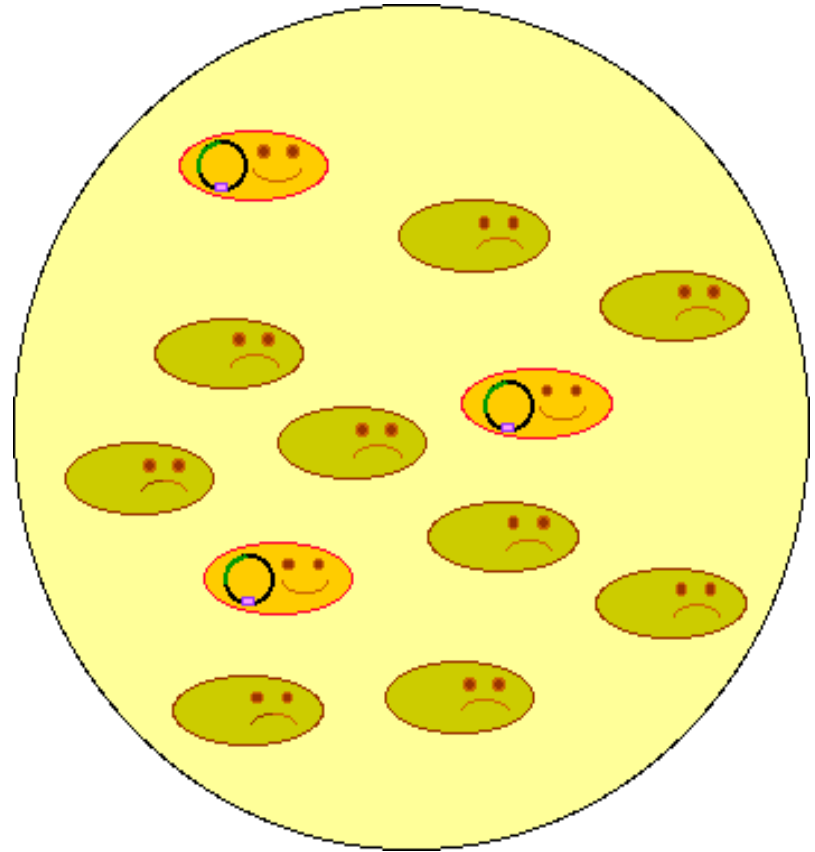


Cell division

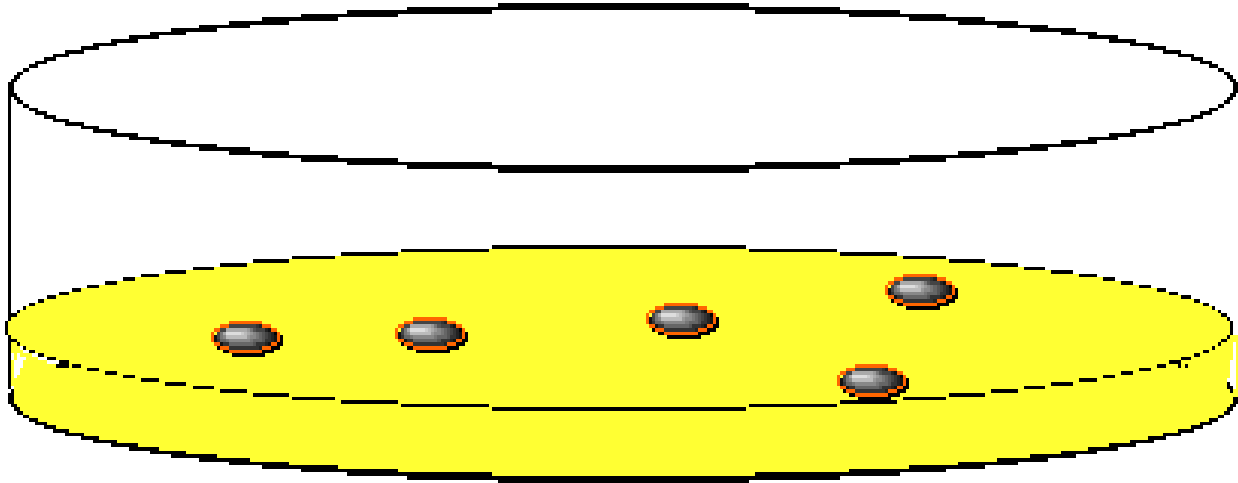


Competent cells culture:

After transformation, cells are plated onto agar medium that contains selective antibiotic: only transformed cells, will survive and form colonies. All the untransformed cells will die.

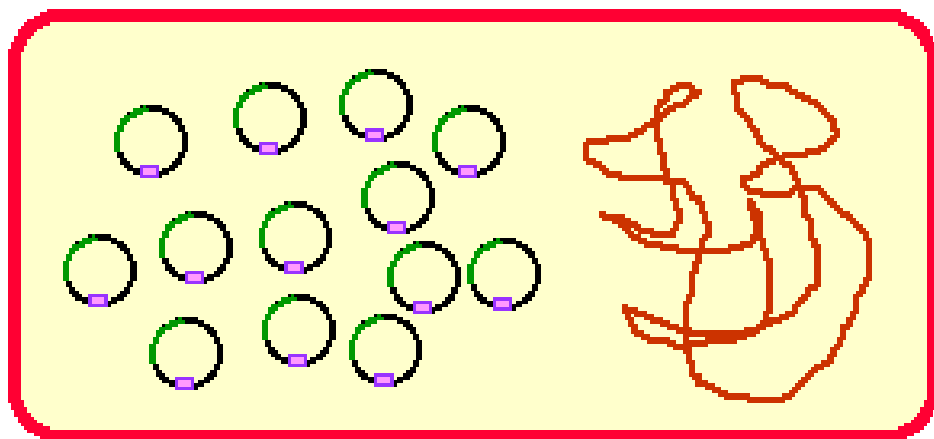
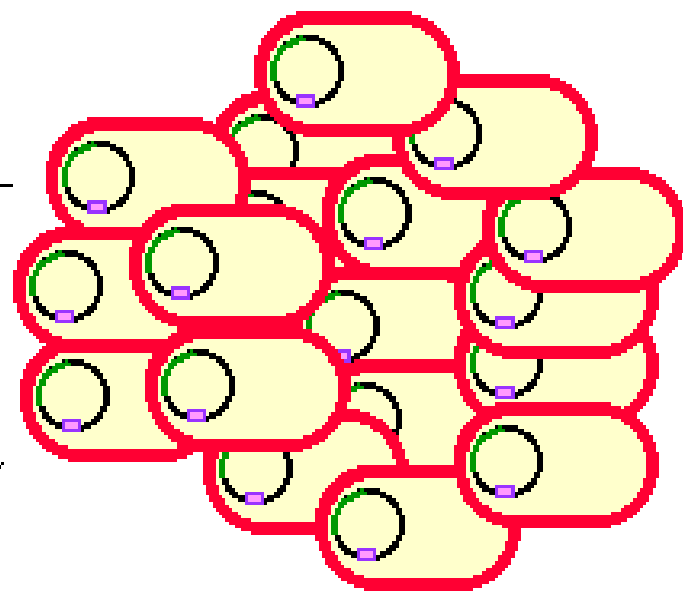
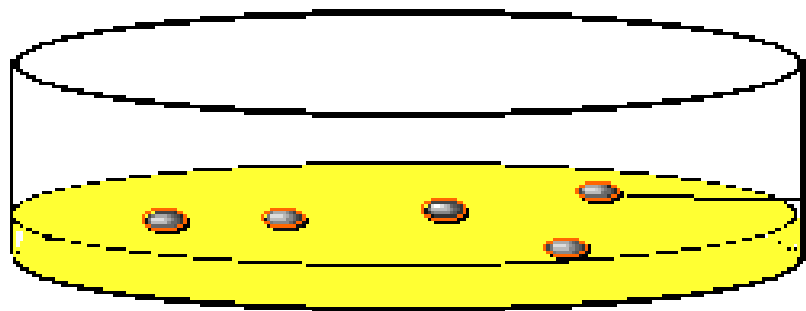


In each colony formed on the agar plate, all cells are descendants of one transformed cell.



agar medium with antibiotic

All cells in the clone are genetically identical and contain the same recombinant vector



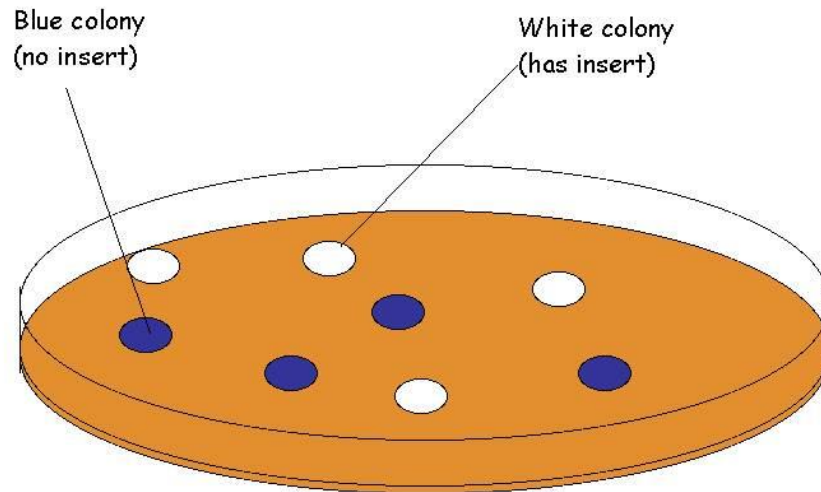
How to Find and Pick Up the Right Clone?

The most common methods include:

- 1. Phenotypic screening.**
- 2. Screening with antibodies.**
- 3. DNA hybridization.**

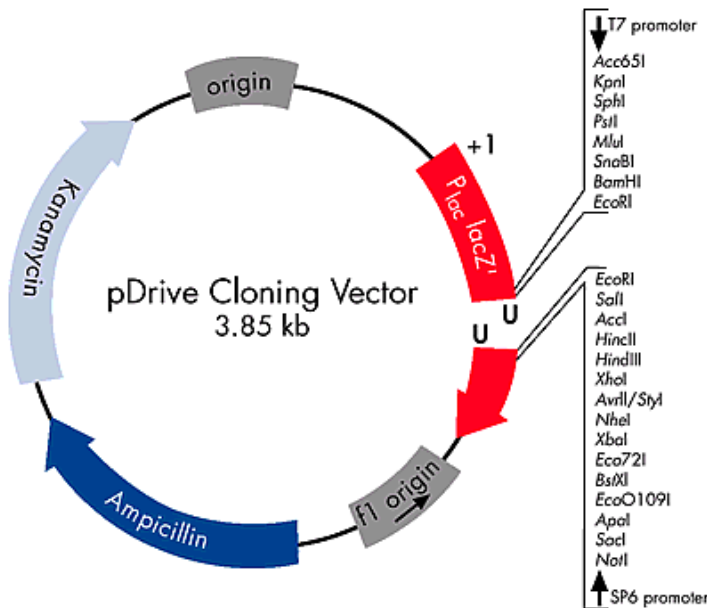
Phenotypic screening

Phenotypic screening is used when cloned gene is expressed and changes properties of the cell in an "obvious way". E.g. **Blue-White Screening**



Blue-White Screening

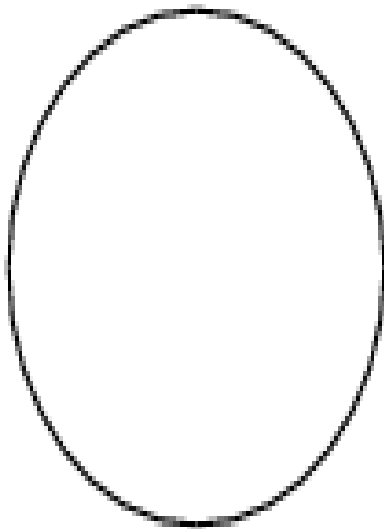
pDrive expresses LacZ α -peptide \rightarrow Provide β -galactosidase activity. X-gal is a colourless analog of lactose cleaved by β -galactosidase to form a bright blue insoluble pigment.



- No insert \rightarrow intact lacZ \rightarrow Blue
- Insert \rightarrow disrupted lacZ \rightarrow White

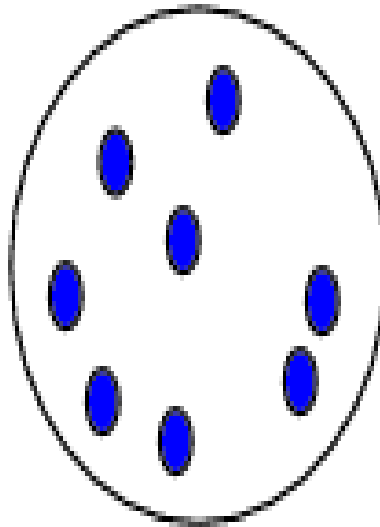
BLUE – WHITE SCREENING

A)



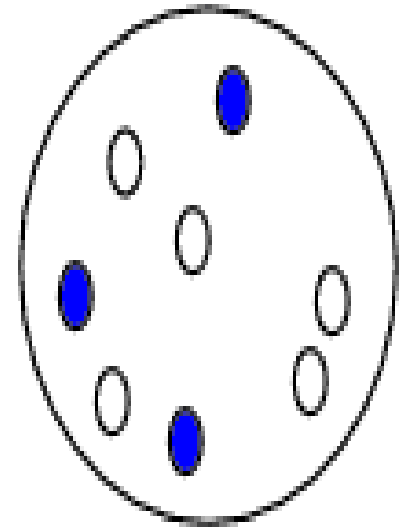
Agar plate
+ ampicillin
+bacteria
+no plasmid

B)



Agar plate
+ampicillin
+bacteria
+plasmid

C)



Agar plate +
ampicillin +
bacteria +
recombinant plasmid

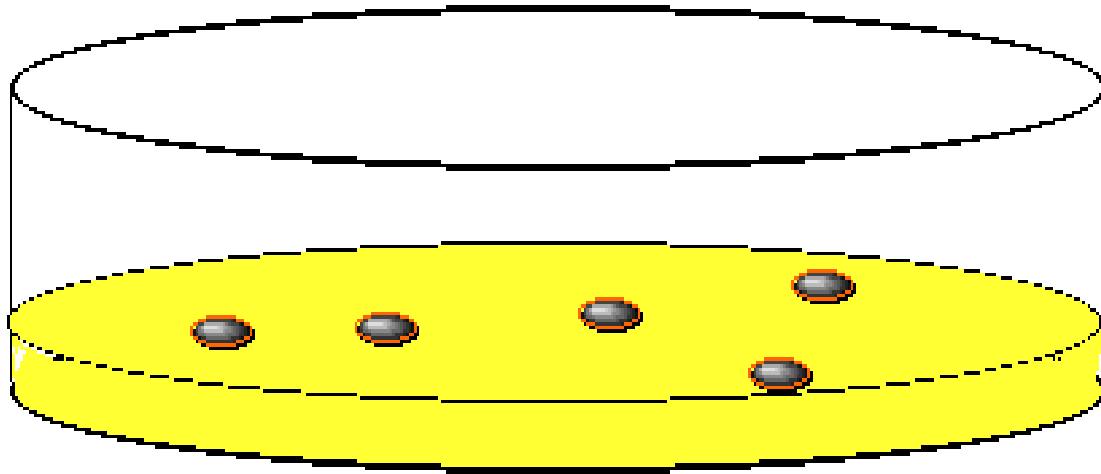
BLUE – WHITE SCREENING



Screening with antibodies

Screening with antibodies is used when cloned gene is expressed and antibodies recognizing the encoded protein are available

→ **(Antibiotic Resistance).**



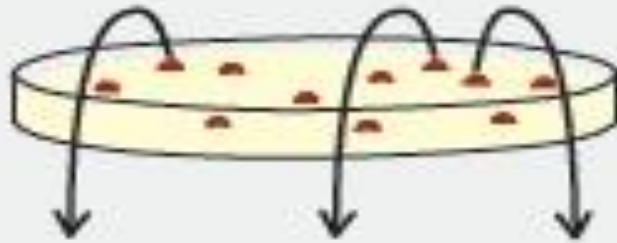
agar medium with antibiotic

Confirmation

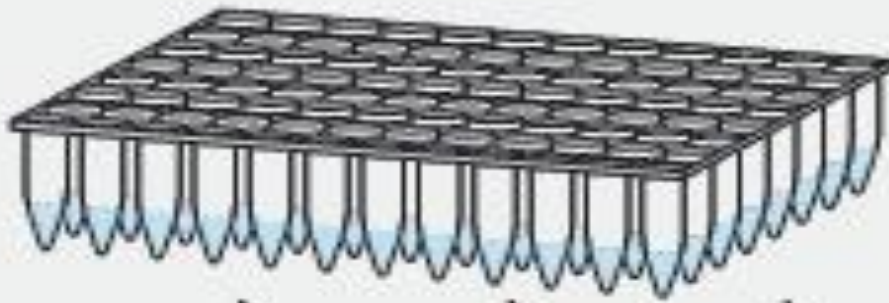
By using PCR:

The same primer
to detect the
insert

Specific primer
for vector
detection

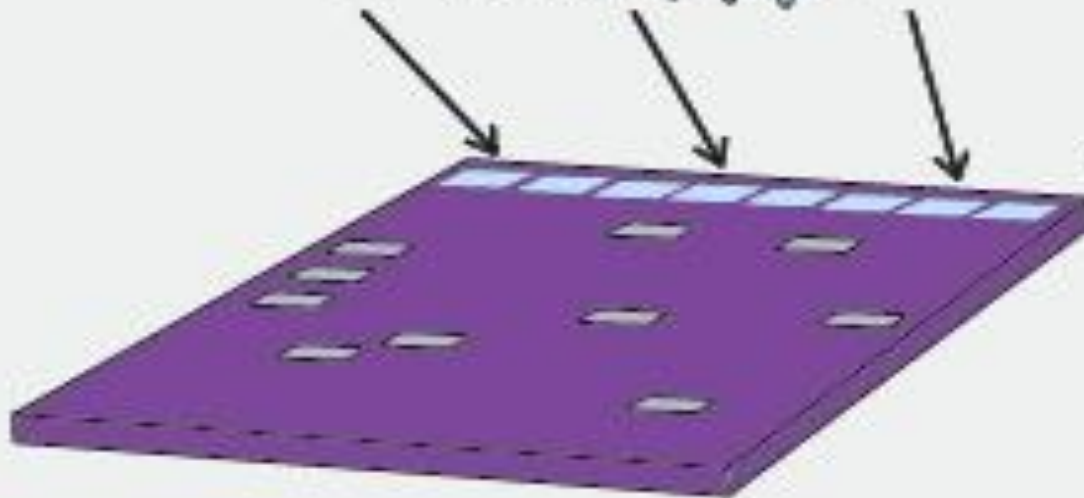


1. Pick colony into a microcentrifuge tube or microtiter well.



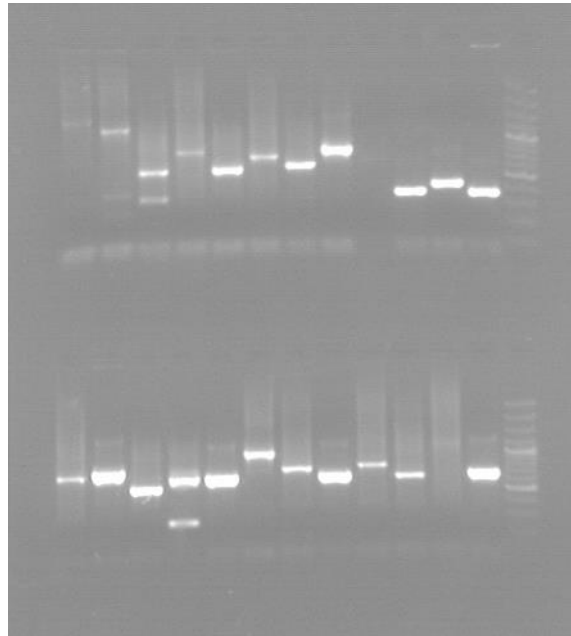
2. Add PCRLyse™ Solution, vortex, heat 5 min at 99°C.

3. Perform PCR using an aliquot of the lysed cells.

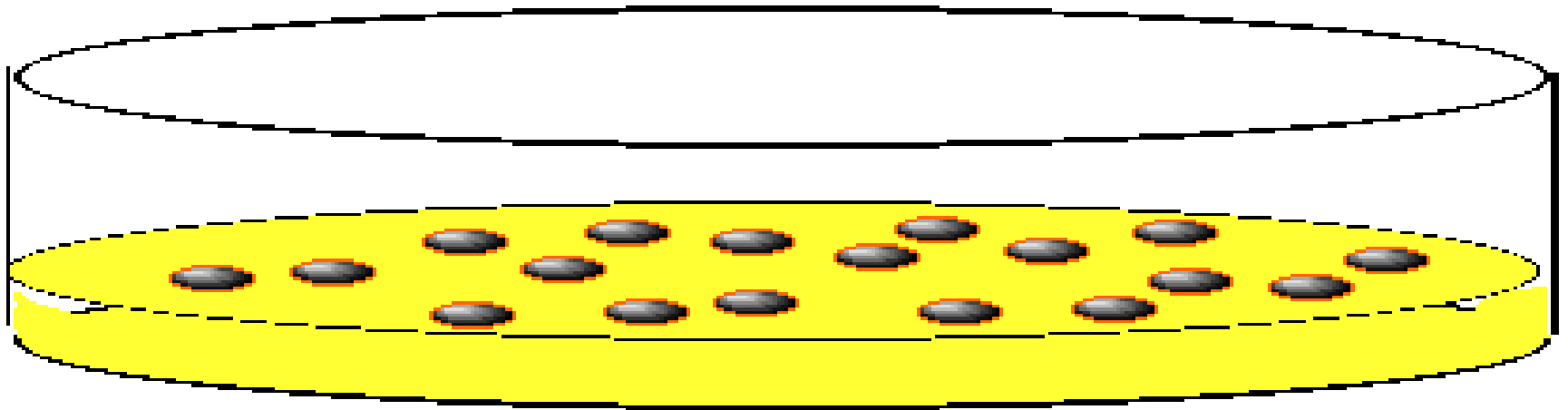


4. Analyze by gel electrophoresis.

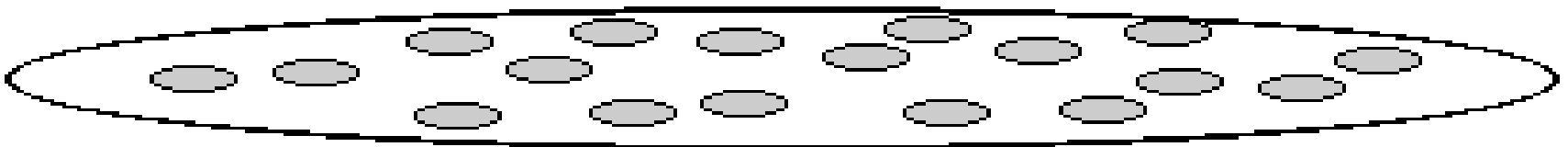
PCR for detection of the vector which carry the insert:



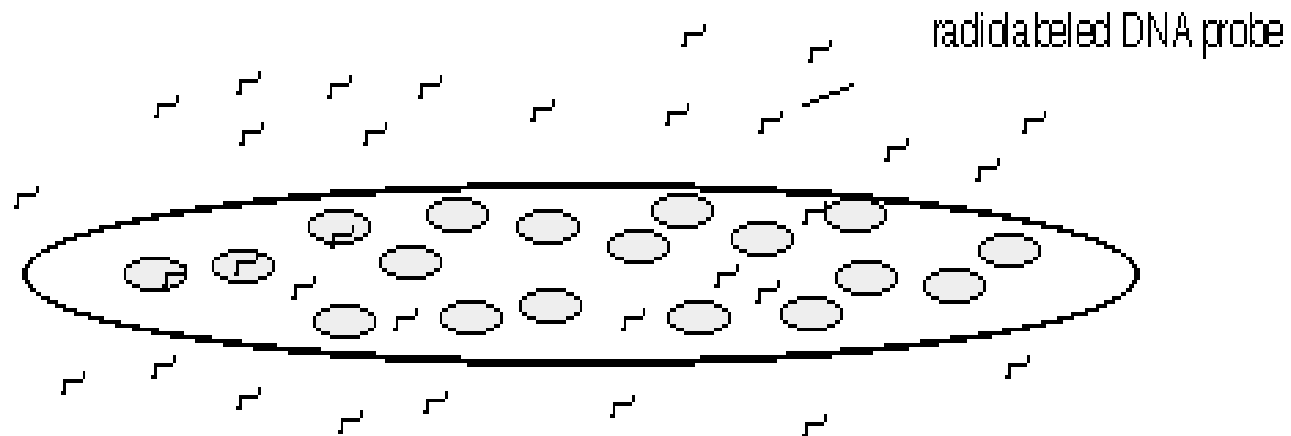
DNA hybridization



TRANSFER COLONIES TO
A NYLON FILTER

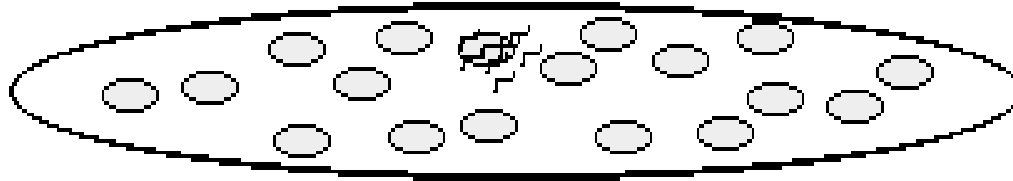


DNA hybridization

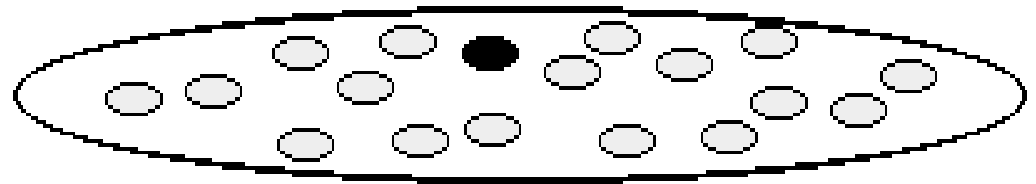


1. Synthesize a DNA fragment complementary to any strand of "our" gene.
2. Radiolabel the probe.
3. Lyse cells on the filter and denature DNA.
4. Hybridize the probe with the filter.
5. Wash out excess of the probe.

DNA hybridization



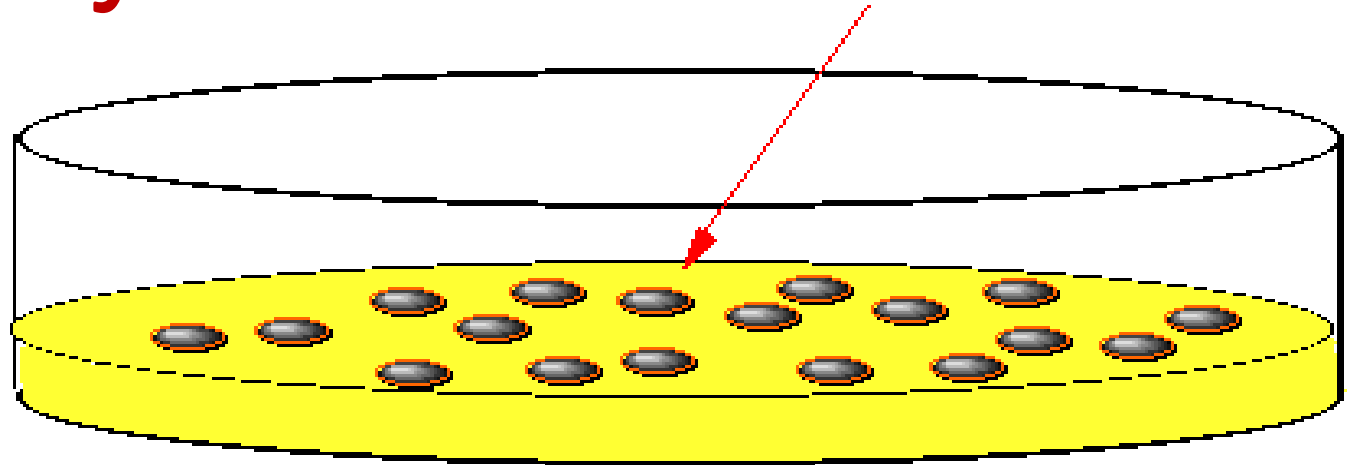
Only the colony that contains plasmids with “our gene” will hybridize with the probe and become radiolabeled.



Expose the filter to the X-ray film: a black spot will appear on the place of a colony with “our gene”.

DNA hybridization

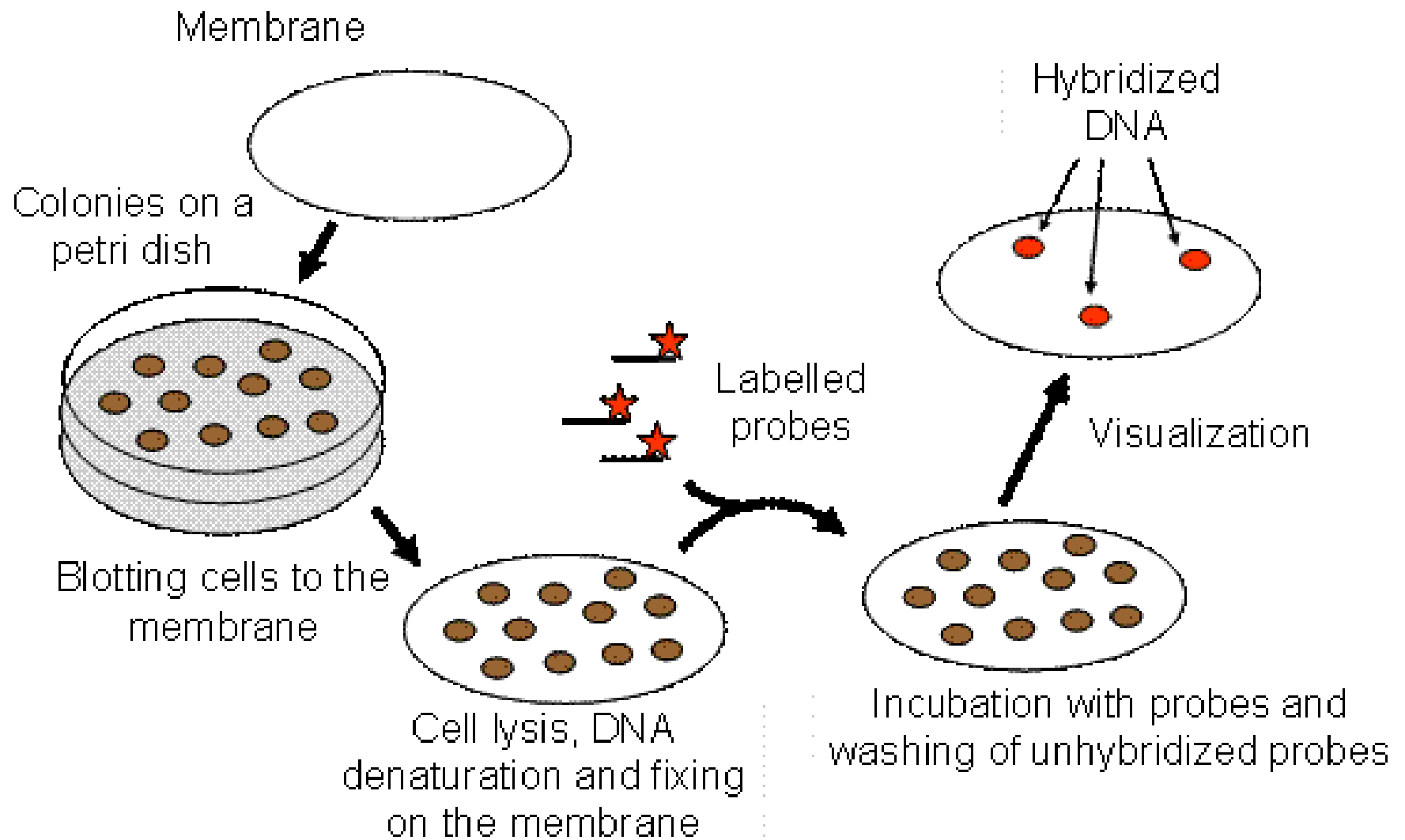
Master plate

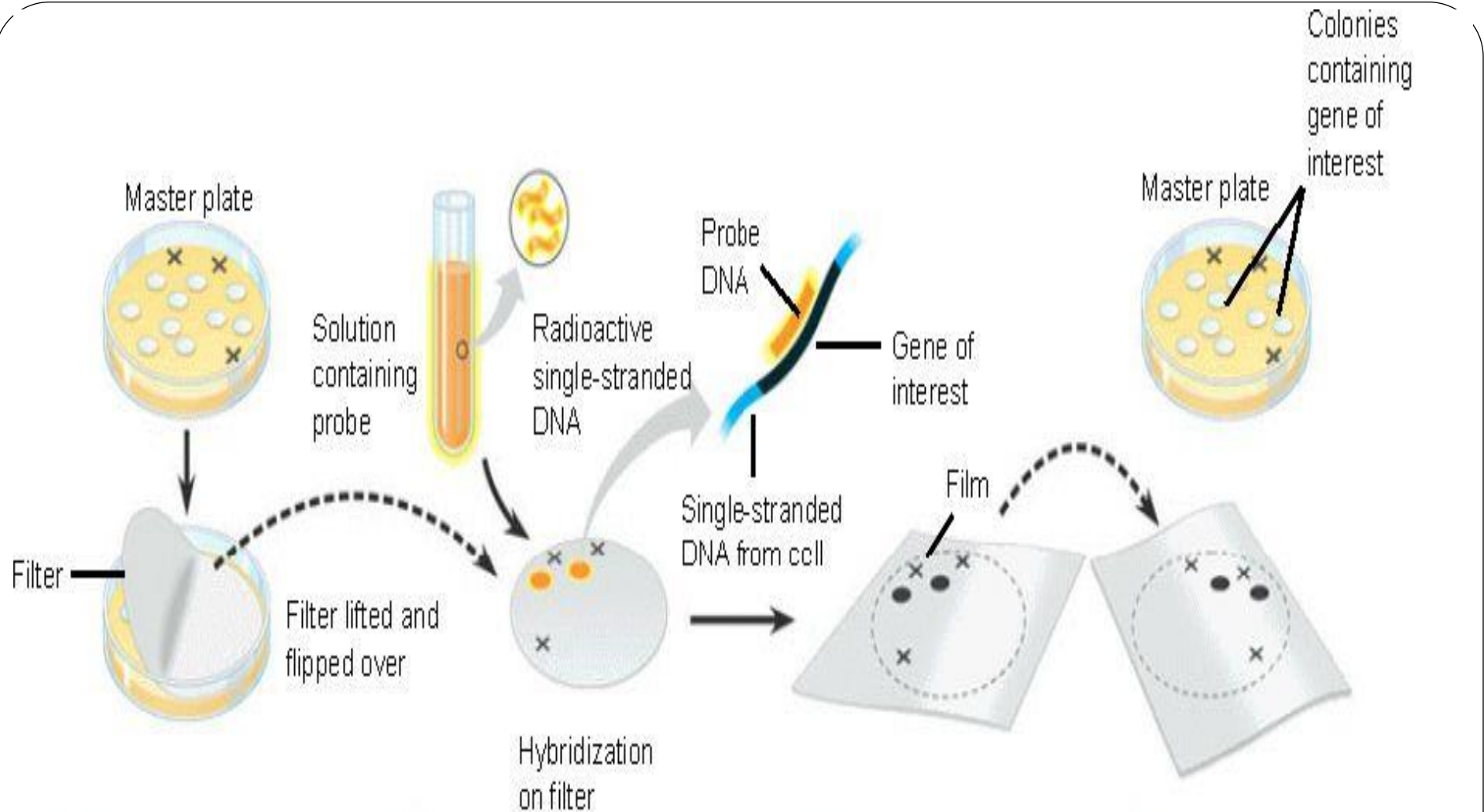


X-ray film



Find the colony on the master plate that contains the cloned gene.





- 1 A special filter paper is pressed against the master plate, transferring cells to the bottom side of the filter.
- 2 The filter is treated to break open the cells and denature their DNA; the resulting single-stranded DNA molecules are treated so that they stick to the filter.
- 3 The filter is laid under photographic film, allowing any radioactive areas to expose the film (autoradiography).
- 4 After the developed film is flipped over, the reference marks on the film and master plate are aligned to locate colonies carrying the gene of interest.

Bacterial colonies containing cloned segments of foreign DNA

Radioactive DNA

Solution containing probe

1 Transfer cells to filter

Filter

2 Treat cells on filter to denature DNA

3 Add probe to filter

Probe DNA

Gene of interest

Single-stranded DNA from cell

Hybridization on filter

4 Autoradiography

Colonies containing gene of interest

Developed film

5 Compare autoradiograph with master plate

Master plate

DNA on membrane (a band "up close")

Low specific activity probe bound to DNA on membrane

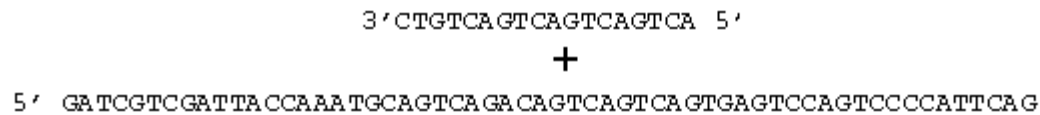
weak signal on Southern

High specific activity probe bound to DNA on membrane

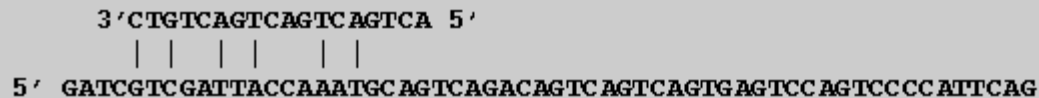
strong signal

DNA hybridization

If two single stranded DNA molecules have complementary nucleotide sequences they can **hybridize**: form a stable double stranded complex



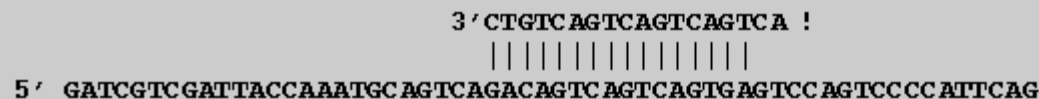
Incubate DNA fragments together




unstable complex



unstable complex



stable complex



Always laugh when you
can. It is cheaper than
medicine.

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Thanks a lot

with my Best Regards and My Best wishes

Amira A. AL-Hosary

E-mail: Amiraelhosary@yahoo.com

Mob. (002) 01004477501