Genome sequencing



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Genome sequencing



DNA Cloning Overview



Overview: Dideoxy (Sanger) Method



Sanger Method: Generating Read

- Start at primer (restriction site)
- 2. Grow DNA chain
- 3. Include ddNTPs
- 4. Stops reaction at all possible points
- Separate products by length, using gel electrophoresis



Automatic DNA sequencing



Cloning vectors for genome sequencing



Sequencing vector M13



DNA Cloning and Sequencing with BACs



Sizes of inserts in sequencing vectors

| <u>Vector</u> | <u>Size (approx.)</u> | |
|----------------|-----------------------|--|
| P1 | 100 Kb | |
| YAC | 300 -1500 Kb | |
| BAC | 70 - 300 Kb) | |
| Cosmid | ~ 40 Kb | |
| Plasmid | 2 -10 Kb | |
| M13 or Phagmid | ~ 1 Kb | |

Genome Sequencing



Genome Sequencing Strategies

1. Clone-by-clone or hierarchical sequencing

2. Shotgun sequencing

Clone-by-clone or hierarchical sequencing strategy

 After constructing a complete physical map, clone by clone sequencing can be started in any specific region.





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Clone-by-clone or hierarchical sequencing strategy

Advantages:

- **1.** Ability to fill gap and re-sequence the regions.
- 2. Ability to distribute the clones to other labs.
- 3. Ability to check the produced sequence by restriction enzymes

Disadvantages:

- 1. Expensive and time-consuming for construction of the physical map.
- 2. Experienced personnel are required.

VENTER'S SHOTGUN Whole genome shotgun sequencing

- It is a much faster approach, and enabled researchers to speed up the timetable for sequencing enormously.
- The shotgun method was developed by J.
 Craig Venter et al., 1996.

Shotgun Sequencing

genomic segment



cut many times at random (*Shotgun*)



Get one or two reads from each segment

Shotgun Sequencing

- Used to sequence whole genomes
- Steps:
 - DNA is broken up randomly into smaller fragments
 - Dideoxy method produces reads
 - Look for overlap of reads



Fragment Assembly



Overlap reads and extend to reconstruct the original genomic region





Shotgun Sequencing

- Very efficient process for small-scale (~10 kb) sequencing (preferred method)
- First applied to whole genome sequencing in 1995 (*H. influenzae*)
- Now standard for all prokaryotic genome sequencing projects
- Successfully applied to **D**. melanogaster
- Moderately successful for *H. sapiens and other genotypes* ...

Comparison of two sequencing methods



Sequencing Successes

T7 bacteriophage

completed in 1983

Escherichia coli







Sacchoromyces cerevisae completed in 1996 12,069,252 bp, 5800 genes

39,937 bp, 59 coded proteins

Sequencing Successes



Drosophila melanogaster completed in 2000 116,117,226 bp, 13,601 genes



Homo sapiens completed in 2003 3,201,762,515 bp, 31,780 genes



Oryza Sativa (Rice) 430 million base 60,000 genes

Genomes to Date

- 8 vertebrates (human, mouse, rat, fugu, zebrafish)
- 2 plants (arabadopsis, rice)
- 2 insects (fruit fly, mosquito)
- 2 nematodes (C. elegans, C. briggsae)
- 1 sea squirt
- 4 parasites (plasmodium, guillardia)
- 4 fungi (S. cerevisae, S. pombe)
- 200 bacteria and archebacteria
- 1900+ viruses

Sequenced Genomes



http://www.genomenewsnetwork.org/



RNA Alternative Splicing and Gene Function in Eukaryoutes





calcitonin gene



Sequencing strategies

• Whole genome



Rapid re-sequencing of human Ad1: Time trial

- Have sequence of Ad 1.
- In theory, have a minimally tiled set of PCR primers to cover entire 36,001 base genome.
- In theory, have a minimally tiled set of sequencing primers as well.
- Want draft sequence in a minimal time, including primer delivery from a vendor.
- In practice design two parallel sets of minimally tiled PCR primers and amplify two sets.
- In practice, assume 750 base reads--> 48 primers, one direction.
- Compare with consensus: Determine accuracy, timing and evaluate operation.

| | | | | | 1 |
|--------|--------|--------|--------|-------|---|
| | | | | 7,315 | |
| | | | 14,500 | 7,300 | |
| | | 21,600 | 14,400 | | |
| | 28,700 | 21,500 | | | |
| 35,885 | 28,600 | | | | |