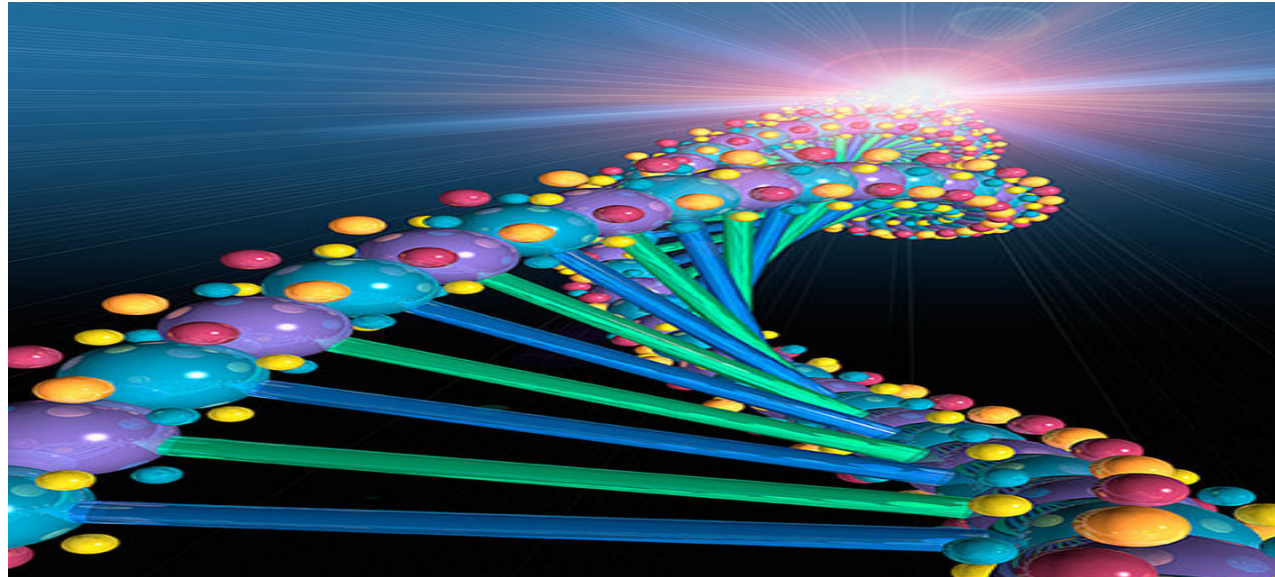


Gene Amplification for cloning



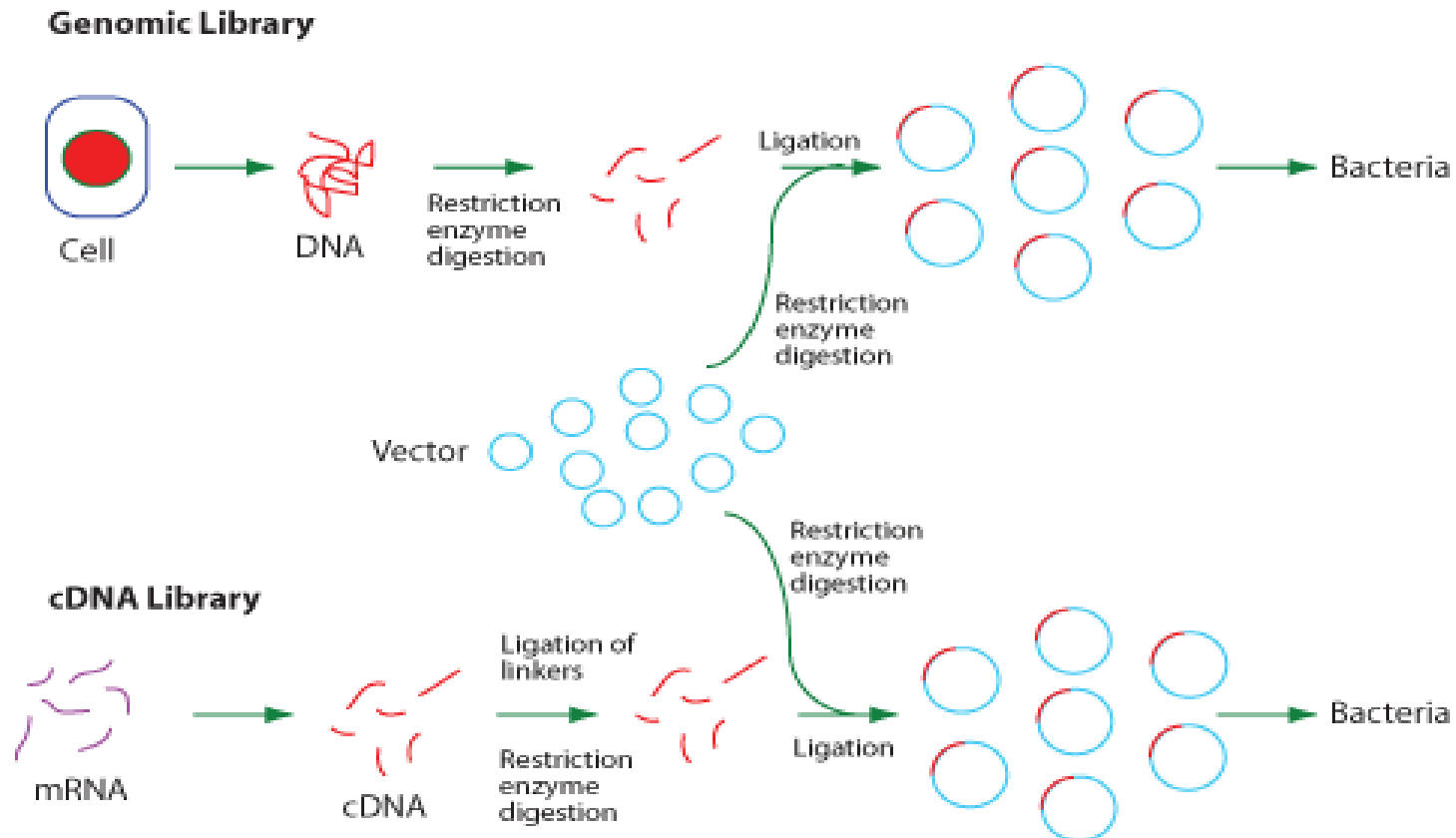
Mohamed N. Seleem

How
to
pick
up



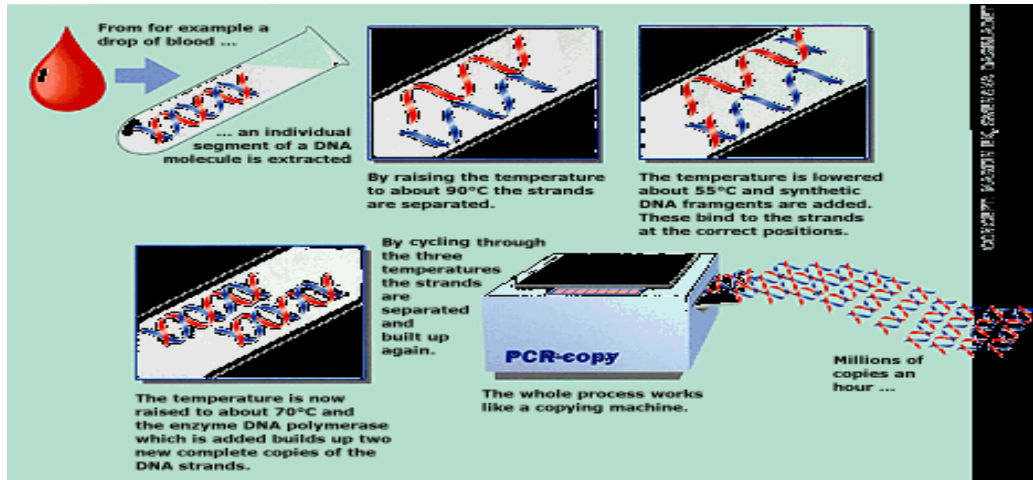
1 out of thousands genes

DNA library and screening



Polymerase Chain Reaction

- Selectively amplifying a particular segment of DNA.
e.g. a specific gene 1 copy can produce billions of copies
- It can be described as a molecular photocopier.



Reagents in a tube

- Sample (DNA with gene of interest)
- Primers (forward and reverse)
- Enzyme (DNA polymerase, *Taq*)
- dNTPs (A, T, G, C)
- Buffer (50 mM KCl; 10 mM Tris-HCl; 1.5 mM MgCl₂)

Master mix

PCR Cycles

Time and Temperature

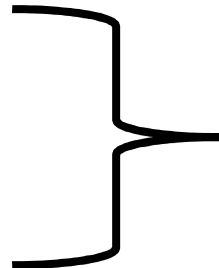
- Initial denature

94 °C for 2 minuets

- denature

- annealing

- Extension



25-40 cycles

94 °C for 30 sec

55 °C for 30 sec

68-72 °C for xxx sec

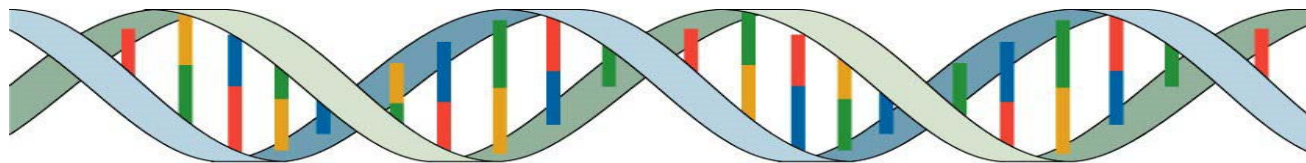
- Final extension

72 °C for 5 minuets

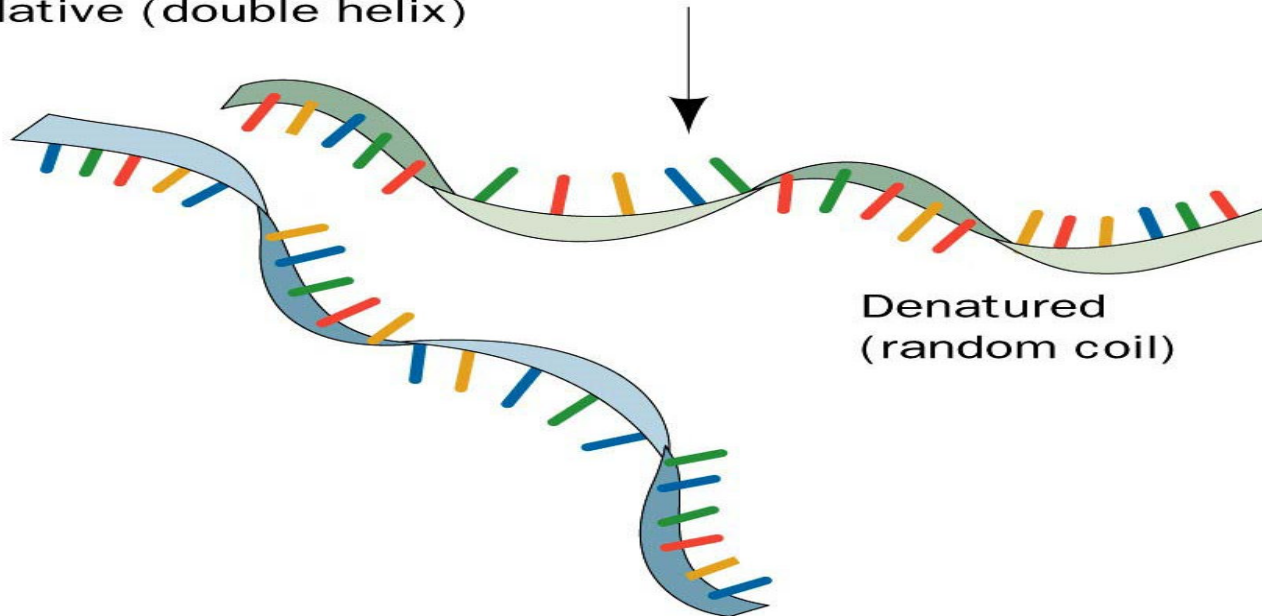
PCR Cycles

- **Denature**

94 °C for 30 sec



Native (double helix)



Denatured
(random coil)

PCR Cycles

- **Primer annealing**

55 °C for 30 sec

ACCATCGGACTGCATCAGTACCATCGG———**ACTGCATCAGTACCATCGGACTGCATCAGA**

TGGTAGCCTGA

oligonucleotide=Primers

oligonucleotide=Primers

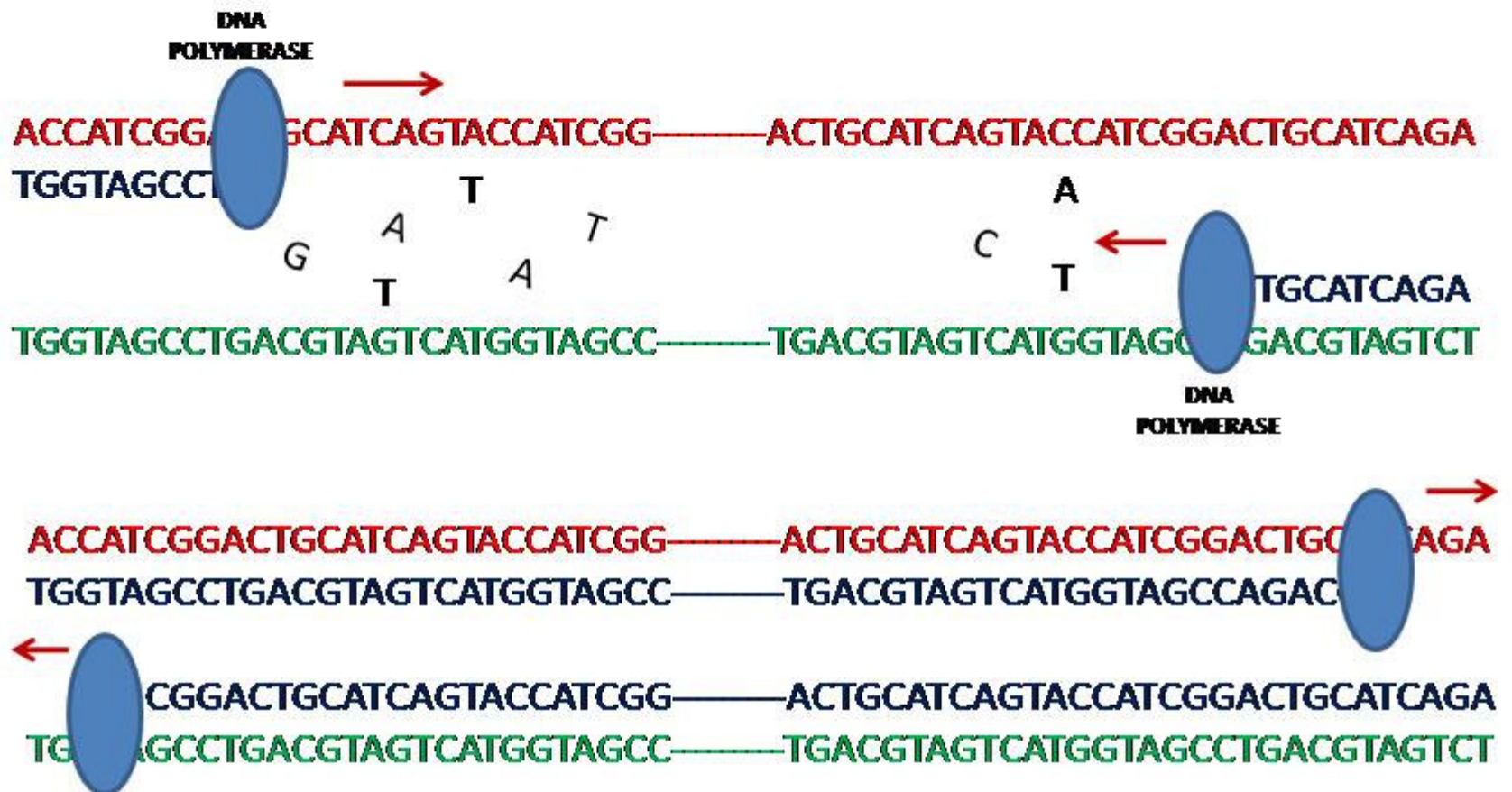
CTGCATCAGA

TGGTAGCCTGACGTAGTCATGGTAGCC———**TGACGTAGTCATGGTAGCCTGACGTAGTCT**

PCR Cycles

•Extension

72 °C for **XXX** sec



2nd cycle

•Denature

94 °C for 30 sec

ACCATCGGACTGCATCAGTACCATCGG———ACTGCATCAGTACCATCGGACTGCATCAGA

TGGTAGCCTGACGTAGTCATGGTAGCC———TGACGTAGTCATGGTAGCCAGACGTAGTCT

ACCATCGGACTGCATCAGTACCATCGG———ACTGCATCAGTACCATCGGACTGCATCAGA

TGGTAGCCTGACGTAGTCATGGTAGCC———TGACGTAGTCATGGTAGCCTGACGTAGTCT

2nd cycle

•Primer annealing

55 °C for 30 sec

ACCATCGGACTGCATCAGTACCATCGG———ACTGCATCAGTACCATCGGACTGCATCAGA
TGGTAGCCTGA

CTGCATCAGA

TGGTAGCCTGACGTAGTCATGGTAGCC———TGACGTAGTCATGGTAGCCAGACGTAGTCT

ACCATCGGACTGCATCAGTACCATCGG———ACTGCATCAGTACCATCGGACTGCATCAGA
TGGTAGCCTGA

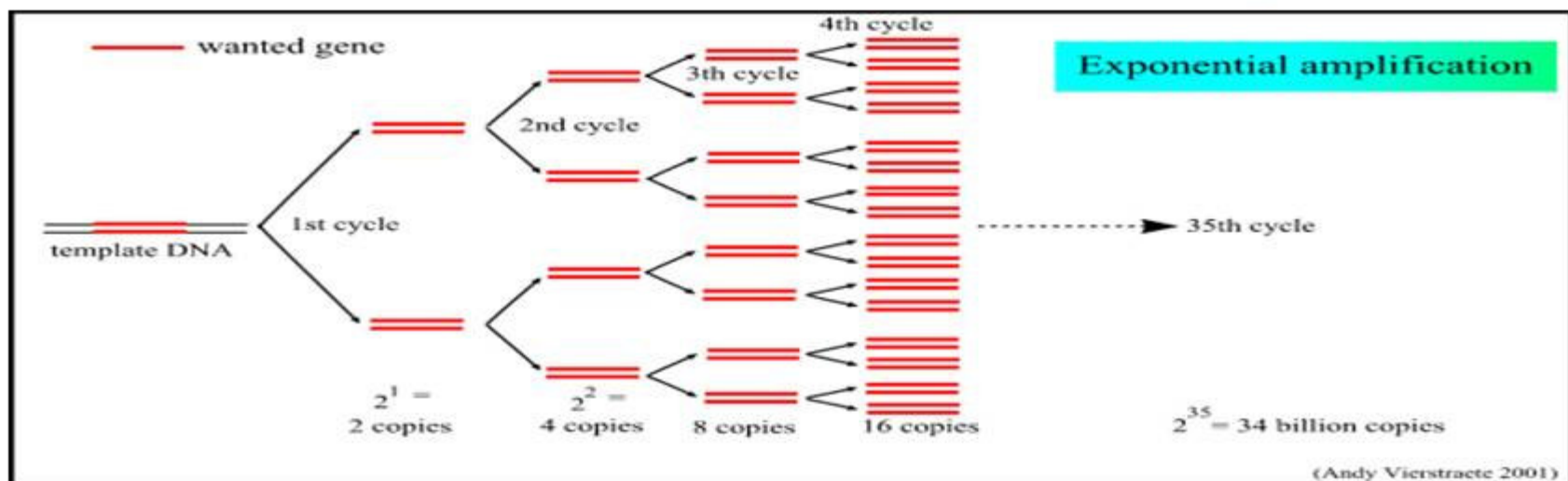
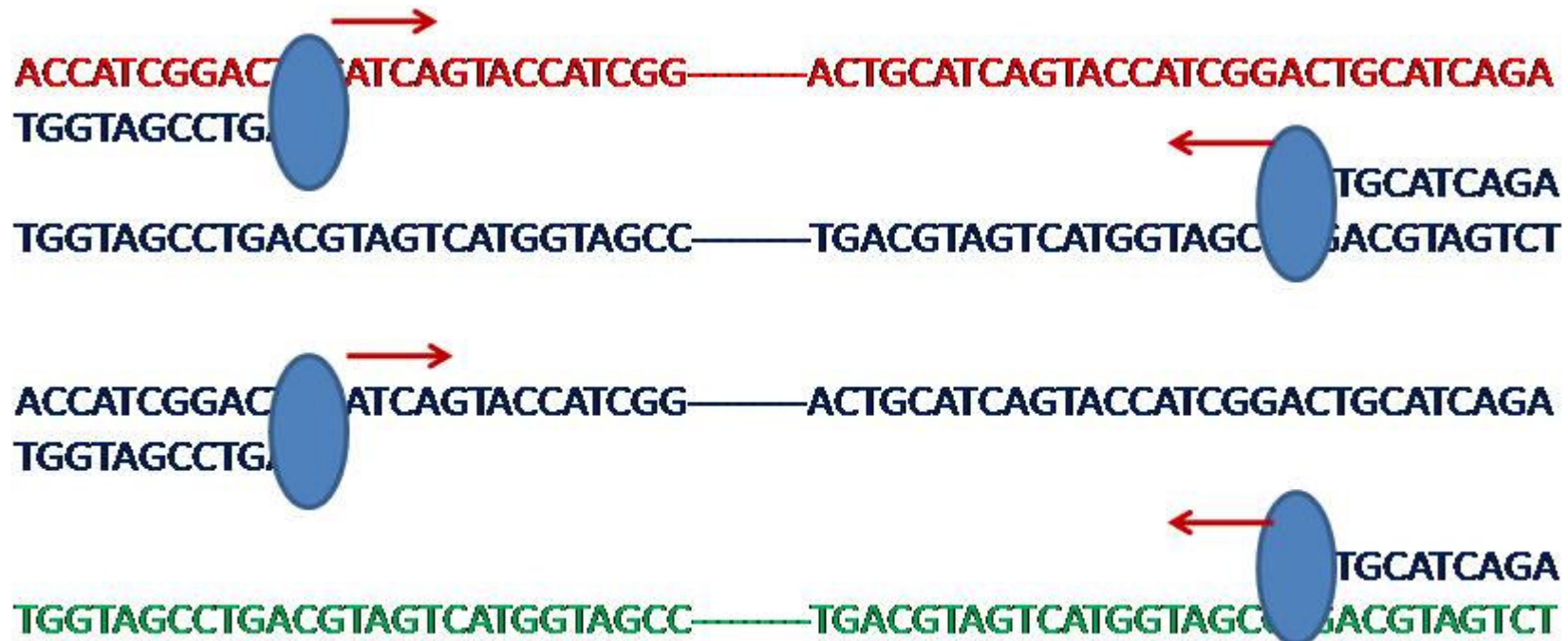
CTGCATCAGA

TGGTAGCCTGACGTAGTCATGGTAGCC———TGACGTAGTCATGGTAGCCTGACGTAGTCT

•Extension

2nd cycle

72 °C for **XXX** sec



Set up PCR reaction



Reagents in a tube

1- 5 μ l (200 ng)

1 μ l F and 1 μ l R 20 pmol

25 μ l – 50 μ l

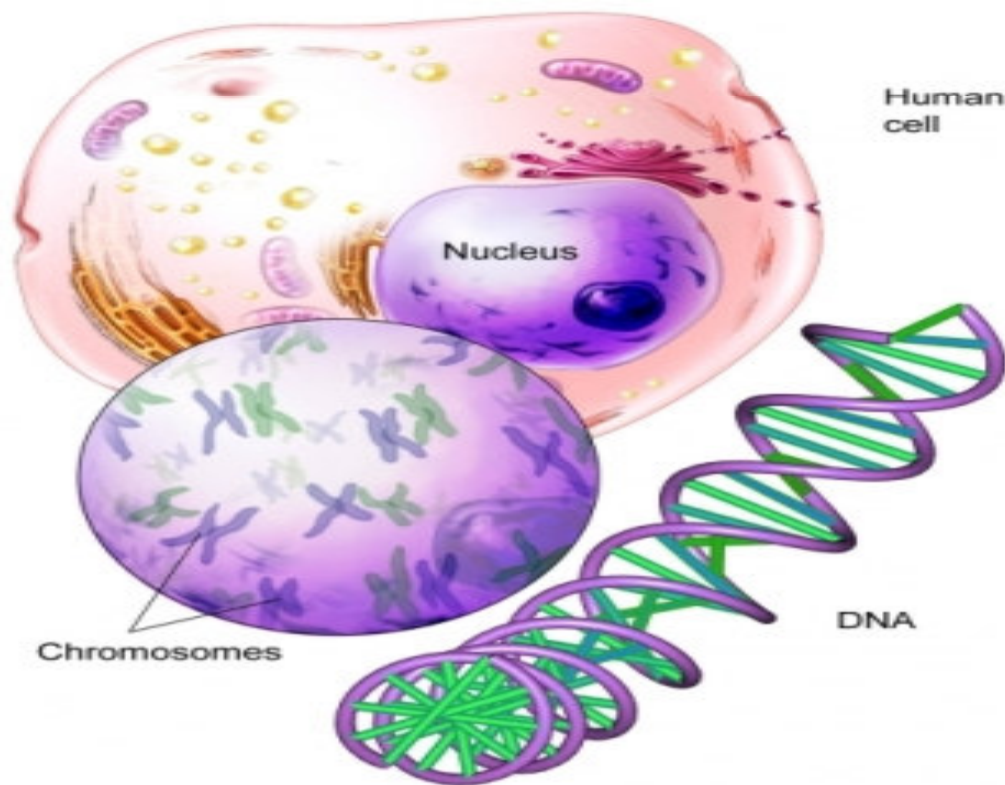
- Sample (DNA)
 - Primers (forward and reverse)
 - Enzyme (DNA polymerase, *Taq*)
 - dNTPs (A, T, G, C)
 - Buffer (50 mM KCl; 10 mM Tris-HCl; 1.5 mM MgCl₂)
- If needed
- Water DNAase free water

Thermocycler

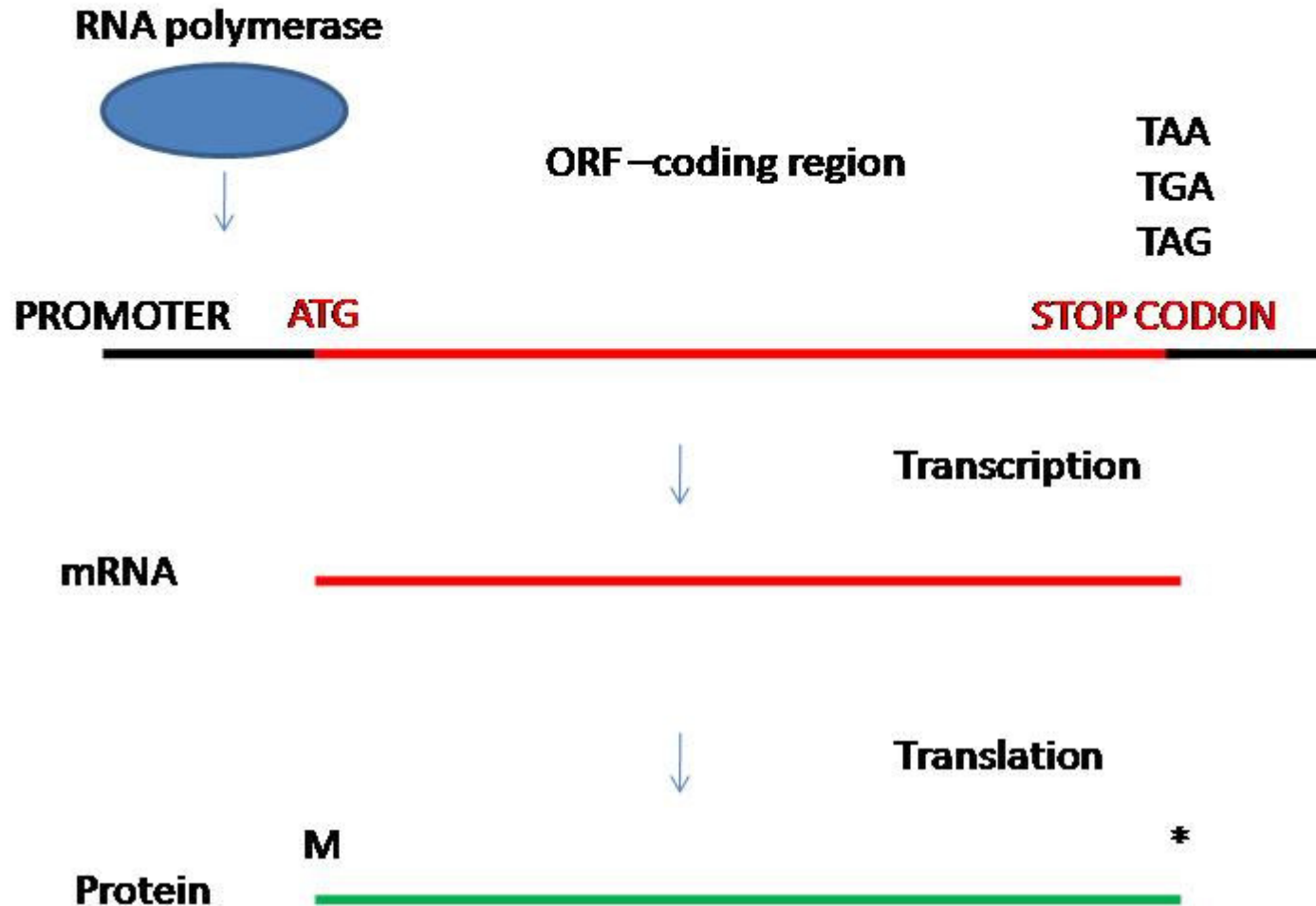


PCR for cloning

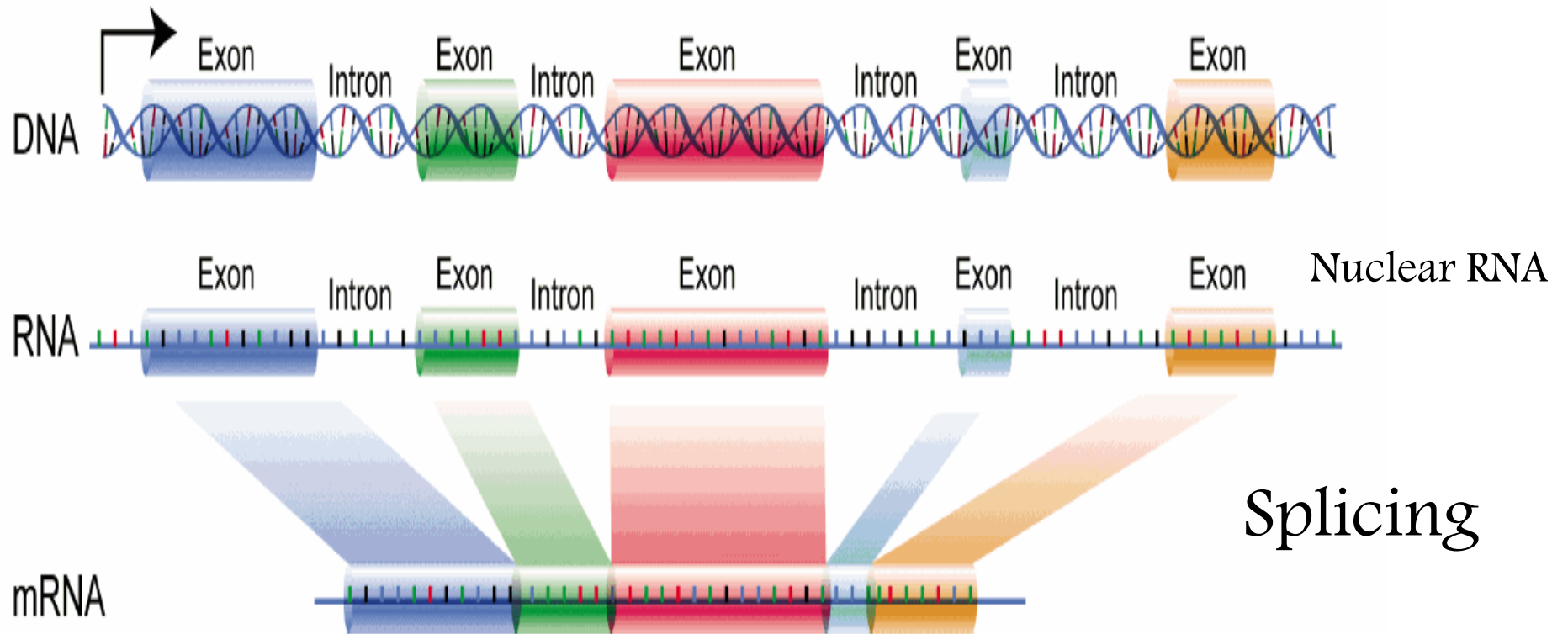
What is different ?



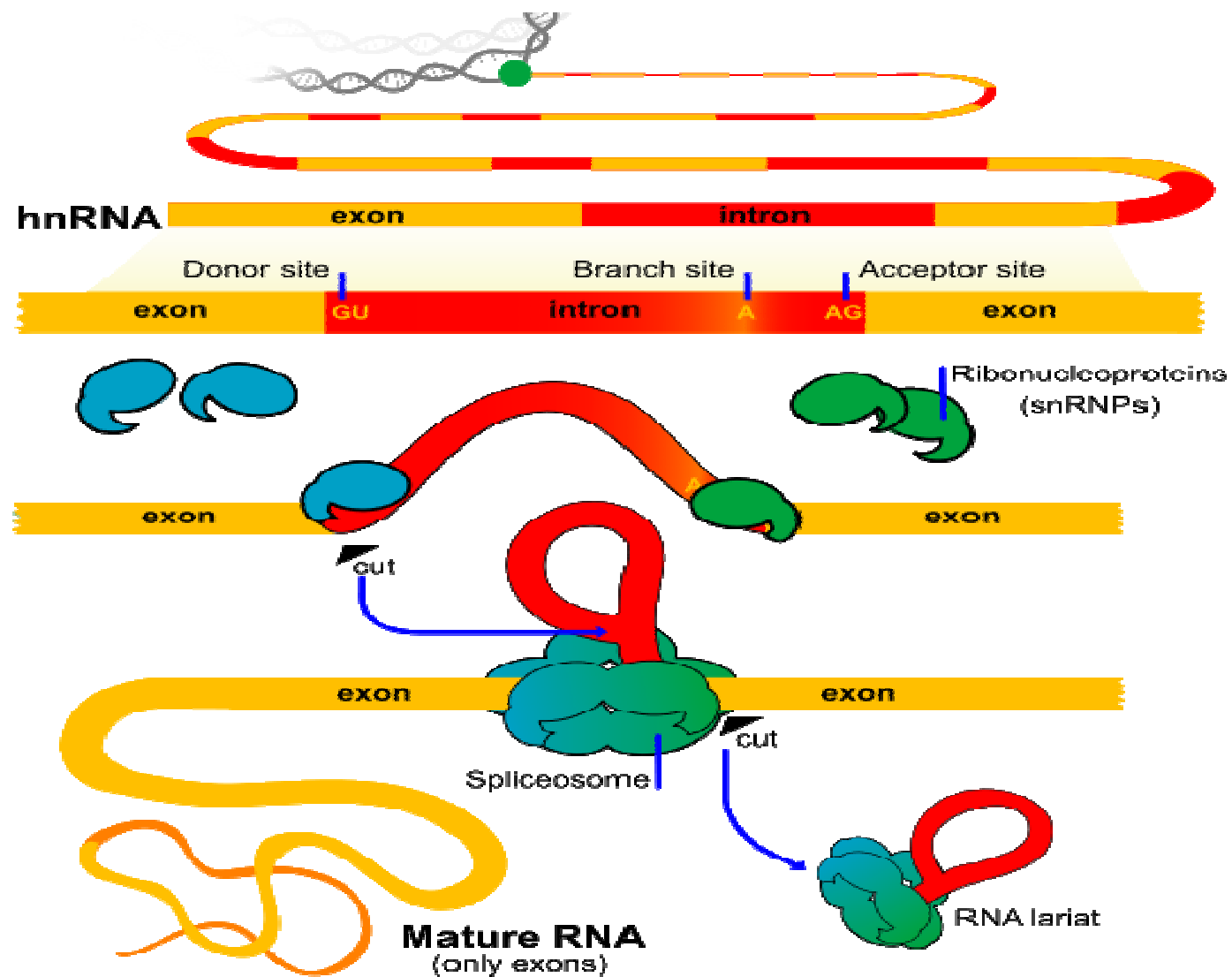
Gene expression



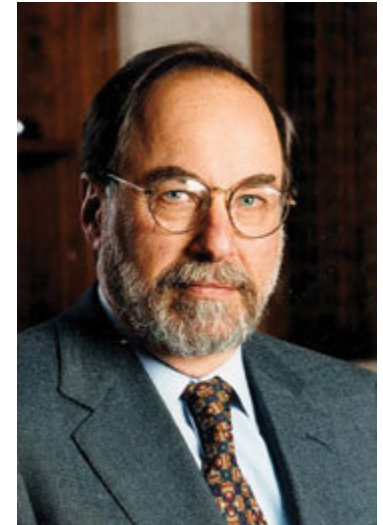
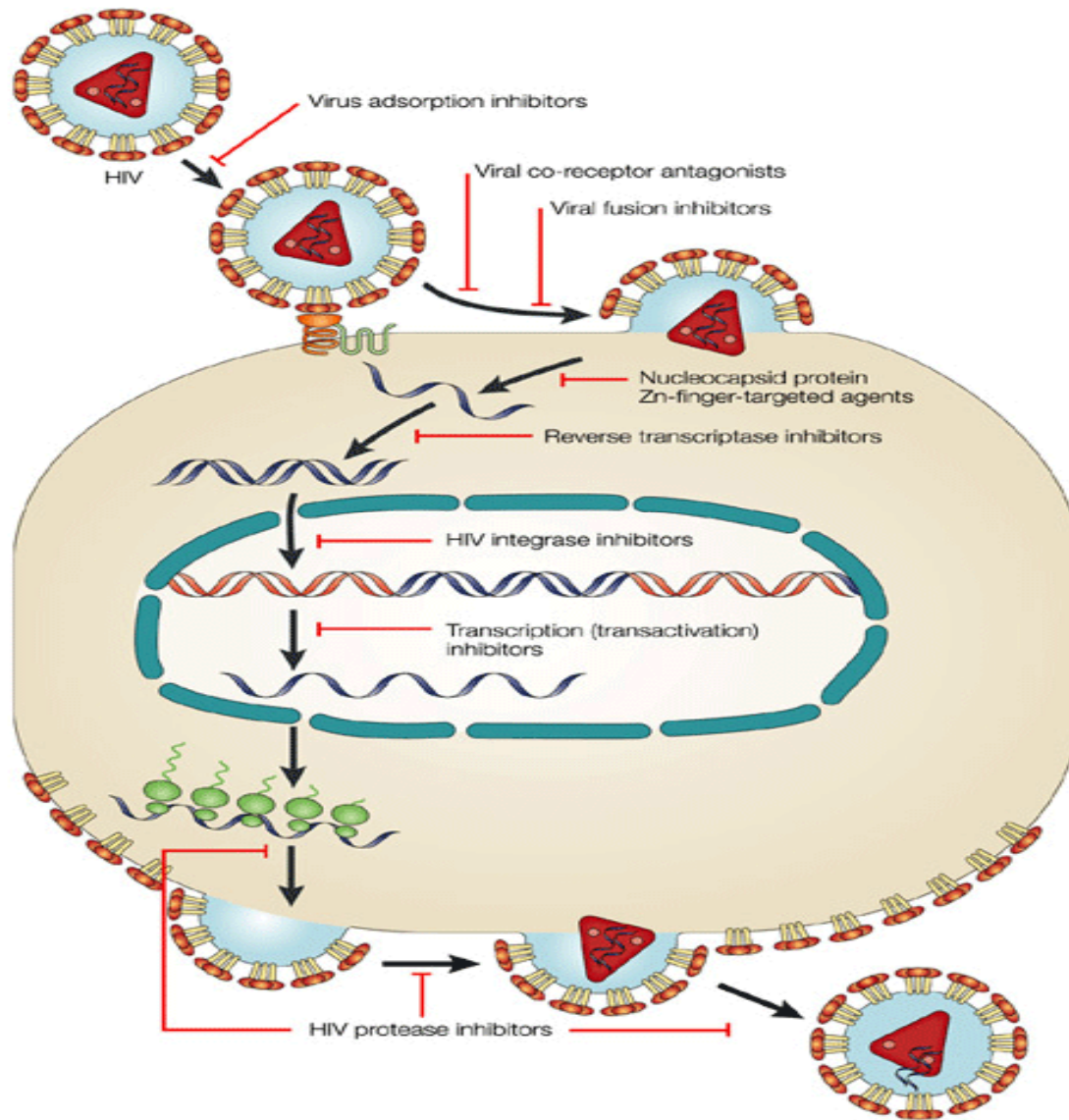
Eukaryotic gene expression



RNA Splicing

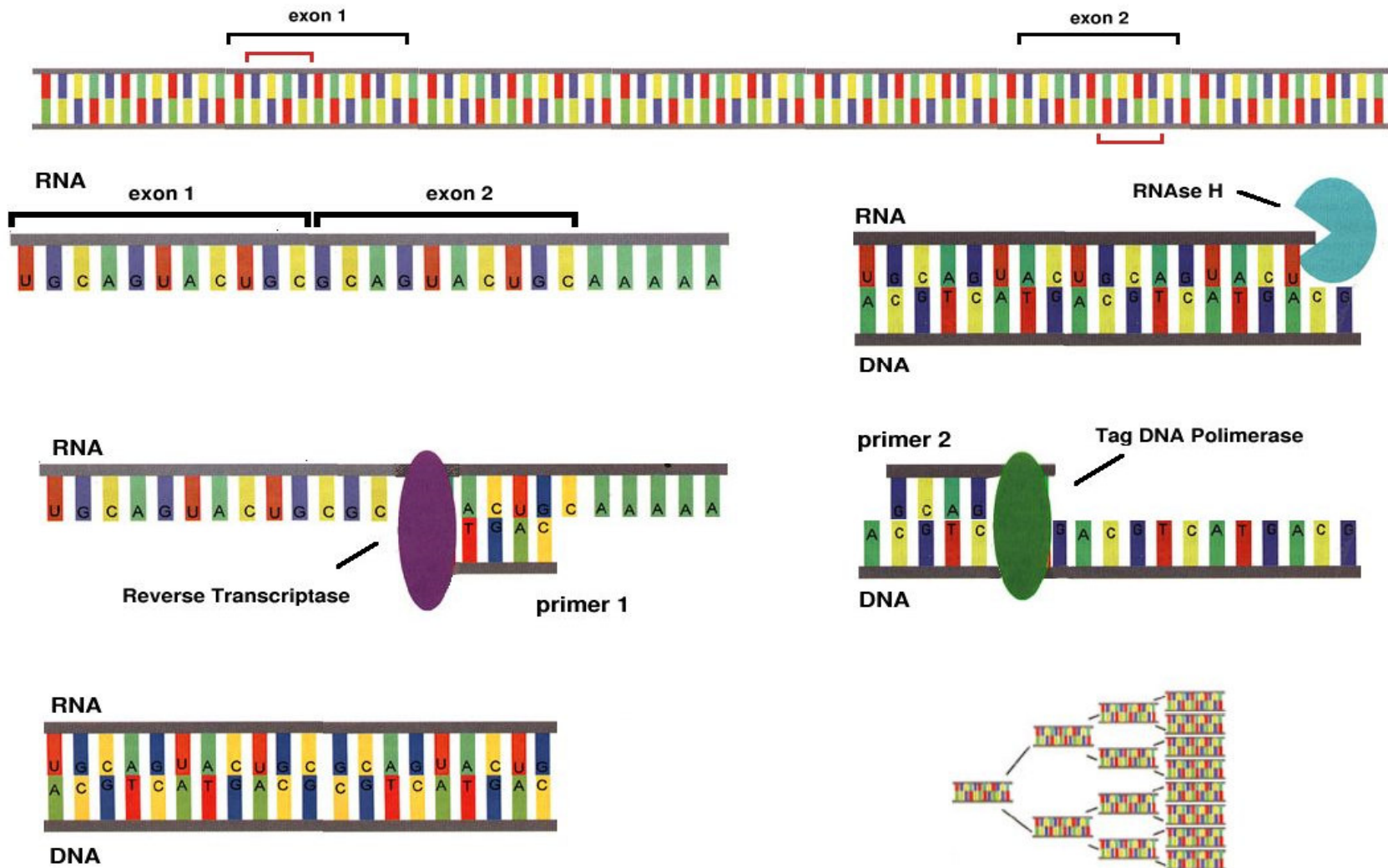


HIV- Reverse transcriptase

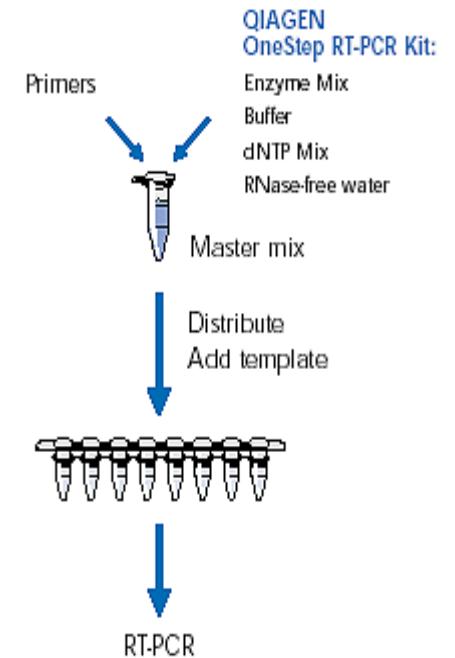


David Baltimore
1975 Nobel Prize
in Medicine

RT-PCR= reverse transcriptase PCR



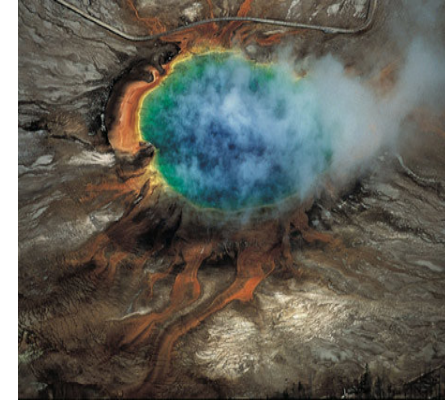
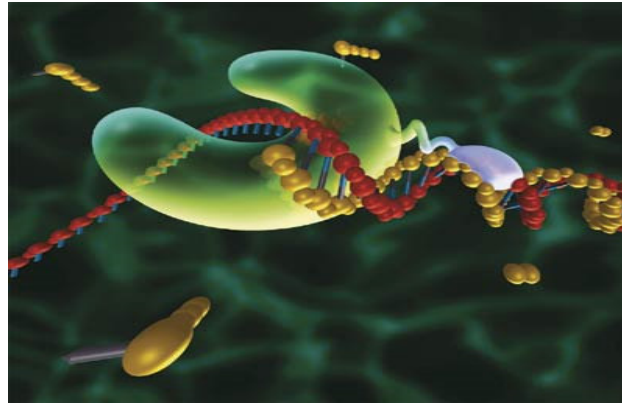
- One step RT-PCR
- Two steps RT and PCR



DNA Polymerase

Thermus aquaticus
Taq enzyme
Heat stable
1988

Error 1:10,000

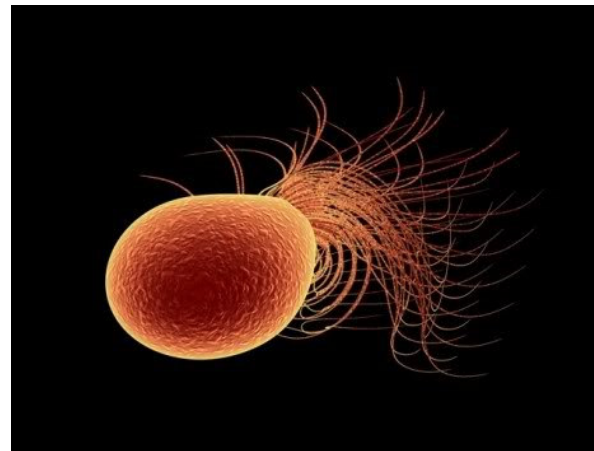


Hot spring

Pyrococcus furiosus
Pfu enzyme
More stable
1991

Error 1: Million

3' to 5' exonuclease proofreading activity



Things to avoid in Primer Design

A primer may be self-complementary and fold into a **hairpin**:

5-GTTGACTTGATATTATCAAG-3

5'-GTTGACTTGATA

|||||T

3'-GAACTAT

A primer may form a **dimer** with itself or with the other primer.

5'-ACCGGTAGCCACGAATTCGT-3'

5'-ACCGGTAGCCACGAATTCGT-3'

|||||||

3'-TGCTTAAGCACCGATGGCCA-5'

PCR Cycles and volume

More much
better than
little

Increase cycle to 40 cycles
Increase volume to 100 μ l

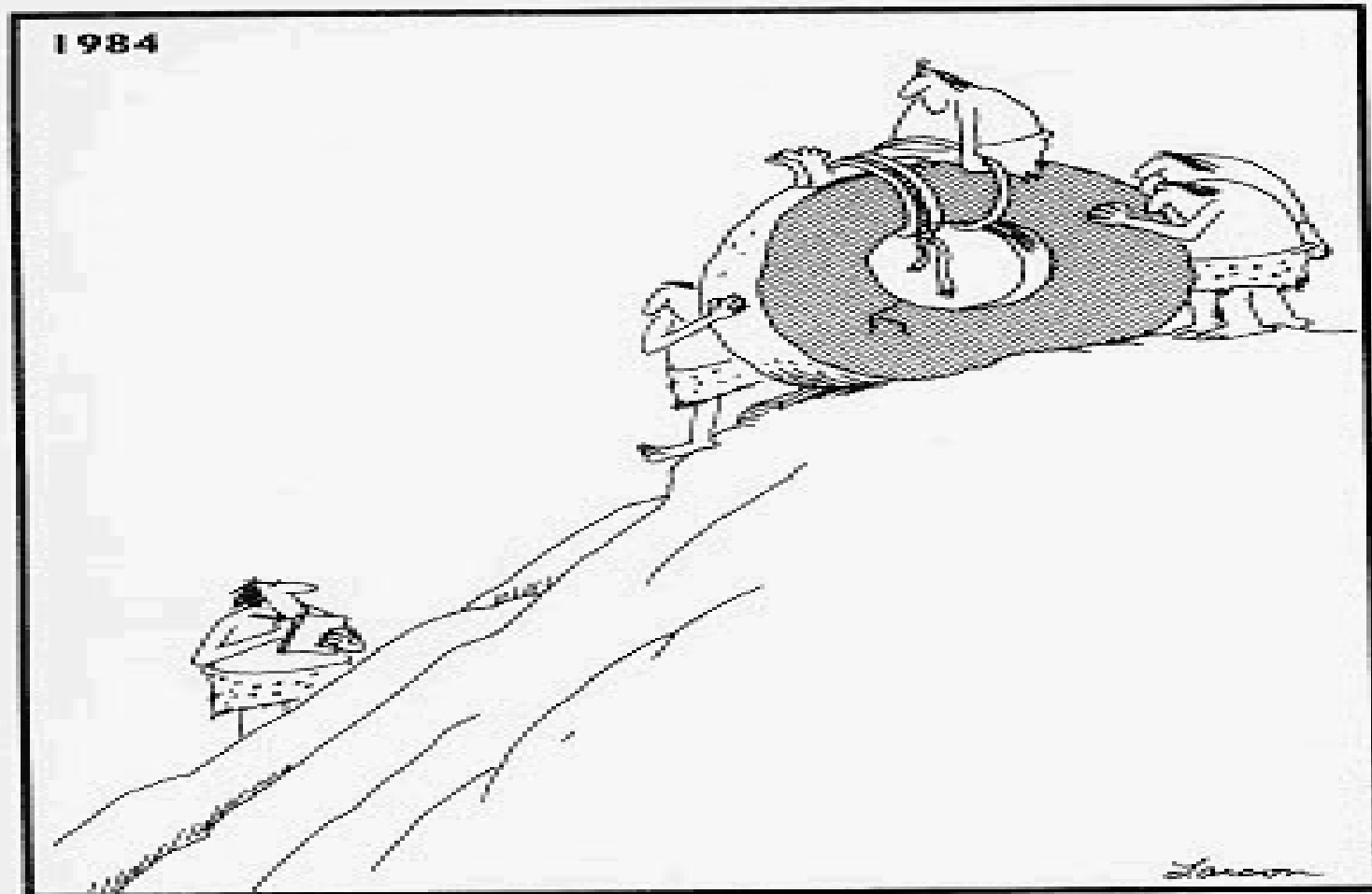
Overnight Thermocycler



Last step incubation

- 10 °C for gene cloning
- Room temperature for diagnosis

Questions!!!



Early experiments in transportation