









Submitted: 6/1/2025

Accepted: 14/3/2025

Published: 30/04/2025

## Diagnostic significance of clinical scoring system and serum lipid profiles as recovery indicators in ketotic dairy cattle using hormonal-steroids-carbohydrates therapeutic protocol

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### ABSTRACT

**Background:** Ketosis is one of the most important metabolic disorders occurring in dairy cows after calving, which is caused by a negative energy balance during the peripartum period or around parturition.

**Aim:** The current study focused on the diagnostic significance of the clinical scoring system, with serum lipid profiles, mainly lecithin cholesterol acyltransferase (LCAT) and apolipoprotein B-100 (ApoB-100), as recovery indicators in ketotic dairy cattle using the hormonal-steroids-carbohydrate therapeutic protocol. The study also discussed the correlation coefficient between serum apo-B100 and serum non-esterified fatty acids (NEFAs) in pretreated and post-treated ketotic cattle.

**Methods:** In this study, 50 ketotic cattle from various dairy farms. The ketotic cows were subjected to a hormonal-steroids-carbohydrate therapeutic regimen extended for a continuous 5 days, including mainly insulin, dexamethasone, and carbohydrate replacement therapy (Glucose solution 50% alternative with dextrose solution 50%) with drenching of propylene glycol. and vitamin B<sub>12</sub> injection. Insulin was administered simultaneously with both of glucose and glucocorticoid (Dexamethasone) and alternatively with a 50% dextrose solution. All animals were undergoing thorough clinical and biochemical investigations including mainly clinical scoring system, serum lipid profiles, and serum enzyme activities (Aspartate aminotransferase; AST) in days 0, 7, 14, 21, and 30.

**Results:** The ketotic dairy cattle needed a maximum 30-day follow-up period post-therapy to restore their physiological status for clinical scoring data (7 days needed) and most serum lipid indices [14 days needed, i.e., apoB-100,  $\beta$ -hydroxy butyric acid, NEFA, glucose, cholesterol, and AST]. A 30-day post-therapy as a follow-up period was urgently required to restore their reference values of serum LCAT. The results of this study indicated that serum LCAT activity and ApoB-100 were the most sensitive indicators of the efficacy of therapy and the restoration of the physiological status. The study revealed a significant negative correlation between serum NEFA and apoB-100 in ketotic cattle. The results of this study indicated that serum LCAT activity and ApoB-100 were the most sensitive indicators of the efficacy of therapy and the restoration of the physiological status. This could be seen by an obvious improvement in serum lipid profiles, mainly LCAT and apoB-100, and clinical findings as well as clinical scoring data. The study identified a significant negative correlation between NEFA and apoB-100 in ketotic cattle.

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**Conclusion:** Exposure of diseased ketotic dairy cattle to this specific therapeutic strategy had a great impact on their recovery from ketosis and improved their health and productivity status. The current study concluded the diagnostic significance of the clinical scoring system (Appetite score, rumen filling score, manure digestion score, and manure condition score), and serum lipid profiles, mainly LCAT and apoB-100, as recovery indicators in ketotic dairy cattle using hormonal-steroids-carbohydrates therapeutic protocol.

**Keywords:** Apolipoprotein B-100, Clinical scoring data, Hormonal-steroids-carbohydrate therapeutic regimen, Ketotic dairy cattle, Lecithin cholesterol acyltransferase.

## Introduction

Dairy cows are vulnerable to a variety of diseases during the postpartum period, particularly in the first 50 days after giving birth. These conditions include abomasal displacement (DA), milk fever, dystocia, metritis, mastitis, and ketosis (Curtis *et al.*, 1985; Khalphallah *et al.*, 2018a, 2018b). Energy-related diseases, such as ketosis, retained placenta, and DA, are occasionally correlated with mastitis, clinical hypocalcemia, and/or dystocia (Khalphallah *et al.*, 2015; Khalphallah *et al.*, 2016a–c; Khalphallah *et al.*, 2018a, b).

The transition period in bovine animals is the period from three weeks prior to calving until three weeks after delivery (Pande *et al.*, 2016). The transition period exhibits distinct metabolic, nutritional, physiological, and nutritional alterations (Quiroz-Rocha *et al.*, 2009; Khalphallah *et al.*, 2015; Khalphallah *et al.*, 2016b; Elmeligy *et al.*, 2021). These adjustments or variations have occurred as a result of adaptations to the animal's endocrine condition, which has promoted late gestation, the initiation of milk production, and delivery events that occurred between (Sundrum, 2015).

Dairy animals are provided benefits for physiological adjustment to elevated energy requirements throughout the transitional phase, particularly after milk yield is delivered. Nonetheless, the success of each animal's adaptation varies widely from one another (Sundrum, 2015). The failure of these physiological adjustments or adaptations later manifests as clinical or subclinical illnesses (Broom, 1993; Khalphallah *et al.*, 2018b).

As a result, research into the physiological adaptations of ruminants has received more attention (Khalphallah *et al.*, 2016c; Fiore *et al.*, 2017; Khalphallah *et al.*, 2018a; Elmeligy *et al.*, 2021; Khalphallah *et al.*, 2023), particularly during this critical period of the production and reproductive cycle of animals.

Curtis *et al.* (1985); Duffield and Bagg (2002) revealed that early lactation cow herds in a state of ketosis had a higher incidence of DA. Conversely, left-displaced abomasum (LDA) was reported to increase the risk or frequency of other post-calving disorders. LDA cows are more likely to develop metritis and complicated ketosis. Therefore, feeding and management strategies that inhibit LDA can reduce the occurrence of some postpartum diseases. Khalphallah *et al.* (2018a) found that the most often observed peripartum period problems (50 DIM) in dairy cows are DA, ketosis, milk fever, retained fetal membranes, dystocia, and mastitis.

Ketosis is one of the most important metabolic disorders noted in dairy cattle after calving and is caused by energy imbalance during calving (Cameron *et al.*, 1998). The manure digestion score (MDS) is considered a very important tool for determining the quality of nutrition recently consumed by the animal, whereas the body condition score (BCS) indicates the nutritional history of the post several weeks to months (Robert, 2013; Khalphallah *et al.*, 2016a).

Manure scores of 1 and 5 are not preferable and usually refer to health problems as well as dietary limitations. Manure score varies with different stages of lactation. This score varies according to lactation stage, as it is 2.5 to 3.5 in close up dry cows, 3 to 4 in far off dry cows, 3 to 3.5 in late lactation cows, 2.5 to 3.5 in early lactation cows, and finally 2 to 2.5 in post-fresh cattle (Hutjens, 1996).

Clinically (CK) or subclinically ketotic (SCK) cattle with DA may be evaluated by observing temperature, rumen filling score (RFS), appetite score, manure condition score (MCS), MDS, and BCS throughout the early postpartum period by applying a health monitoring program during this follow-up period (30 day) is suitable to enable the dairy cows to restore their healthy status following DA correction (Khalphallah *et al.*, 2016a). However, these items have not been completely elucidated in ketotic cattle with reference to their response to a complete therapeutic regimen.

Khalphallah *et al.* (2018a) concluded that serum lecithin cholesterol acyltransferase (LCAT) is a highly sensitive indicator of the transition phase. Furthermore, it had a greater predictive value than the other lipid profile indices, such as glucose, apolipoprotein B-100 (ApoB-100), and non-esterified fatty acids (NEFAs), in transition period disorders or diseases. The changes in serum LCAT activities have a high diagnostic significance because they enable cattle practitioners to predict metabolic disorders and establish specific dietary, management, and medical protocols to prevent their occurrence.

It is obvious that reduced LCAT values during the peripartum period in dairy cattle precede the occurrence of ketosis or milk fever (Nakagawa and Katoh, 2000). Furthermore, LCAT was recently reported (Satoh *et al.*, 2024) as a predictor of retained placenta in close-up dairy cows. According to Nakagawa and Katoh (1998), cattle that appear healthy but have lower LCAT activity are more susceptible to diseases related to fatty liver than cattle with normal activity.

Many studies have investigated the efficacy of various therapeutic protocols in dealing with ketosis in dairy cows, including both treatment and management (Tufani *et al.*, 2011), in which complex therapeutic programs have been used (Tufani *et al.*, 2011; Madreseh-Ghahfarokhi *et al.*, 2018). On the other hand, other researchers have applied simple therapeutic protocols (Duffield *et al.*, 1998; Radostits *et al.*, 2007; McArt *et al.*, 2011).

Numerous efficient treatments are available for ketosis in cattle, but in some affected animals, the response is only temporary; in rare instances, the disease can persist and result in death or force the animals to be slaughtered (Constable *et al.*, 2017).

The prognosis and response to the applied therapeutic program are associated with ketosis type I or II. Type I ketosis, which is reported at the peak of lactation (3 to 6 weeks post-calving), usually responds rapidly to treatment. However, if the diet is not changed, relapses are frequent. Type II periparturient ketosis usually responds to treatment less immediately (Holtenius and Holtenius, 1996; Herdt, 2000; Anderson and Rings, 2009).

Several components of successful treatment of ketosis are demonstrated as they are based on restoring blood glucose values as soon as possible and replenishing oxaloacetate, an important intermediate in the tricarboxylic acid (TCA) cycle in the liver; thus, there is total oxidation of the fatty acids released from the fat depots. This enhances the availability of dietary glycolytic precursors, particularly propionic acid, and decreases the rate of ketone body production (Andrews *et al.*, 2004). Combined insulin and glucocorticoid therapy may be more efficient than glucocorticoid alone. Insulin is also a strong antiketogenic agent that inhibits NEFA mobilization. Nonetheless, it is essential to provide a glucocorticoid or another medication that can compensate for the hypoglycemia-like effects of insulin (Andrews *et al.*, 2004; Anderson and Rings, 2009).

The current study focused on the diagnostic significance of the clinical scoring system, serum lipid profiles, mainly LCAT and ApoB-100 as recovery indicators in ketotic dairy cattle using the hormonal-steroids-carbohydrates therapeutic protocol. The efficacy of the applied hormonal-steroid-carbohydrates therapeutic protocols and the time elapsed until the dairy cows had restored their physiological status were also assessed. The study also investigated the correlation coefficient between apo-B100 and NEFA and their reflection in recovered cattle.

## Materials and Methods

### Animals

In this study, 50 ketotic cattle from various dairy farms were examined. The dairy farms belonged to the Cairo and Giza governorates in Egypt. The ketotic cows were diagnosed as SCK cows or clinically ketotic

cows (CK) based on clinical findings and estimation of serum  $\beta$ -hydroxy butyric acid (BHBA) levels in day 0 pretherapy, i.e., lactating cows 5 to 50 days in milk. The diseased animals were either SCK (Serum BHBA values 1.2–2.4 mmol/l) or CK (Serum BHBA values  $\geq$  2.5 mmol/l) according to Sakha *et al.* (2006); Sakha *et al.* (2007); Tehrani-Sharif *et al.* (2012); Constable *et al.* (2017). Their body weight varied between 400 and 470 kg. Their age varied between 4 and 6 years. Their rations were mainly the total mixed ration (TMR). The farm's system was a tie stall. Hormonal-steroids-carbohydrates therapeutic protocol was applied to the ketotic cows, which continued for 5 days to induce the recovery of diseased animals. The diseased cows were examined and sampled 5 times as follows; days 0, 7, 14, 21, and 30. Zero day was named 0 day pretreated ketotic cattle (0D-Ket<sup>gr</sup>). 7, 14, 21, and 30 were following treatment as they were named 7 days post-treated ketotic cattle (7Ds-Ket<sup>gr</sup>), 14 days post-treated ketotic cattle (14Ds-Ket<sup>gr</sup>), 21 days post-treated ketotic cattle (21Ds-Ket<sup>gr</sup>), and 30 days post-treated ketotic cattle (30Ds-Ket<sup>gr</sup>), respectively. Ketotic dairy cattle underwent complete clinical examination and laboratory assays to estimate serum lipid profiles, mainly serum LCAT activities, apoB-100, glucose levels, BHBA concentrations, NEFA, cholesterol values, and aspartate aminotransferase (AST) activities.

### Samples

All precautions regarding sample collection and preparation to obtain an accurate assessment of biochemical indices were taken into consideration. Plain vacutainer tubes were used to collect blood serum samples and then stored at  $-20^{\circ}\text{C}$  until analysis, according to Coles (1986). Ketotic cows were sampled before treatment on day 0 and thereafter on days 7, 14, 21, and 30 following treatment.

### Therapeutic strategy

The applied therapeutic regimen (Hormonal-steroids-carbohydrate therapeutic regimen) was administered for a continuous 5 days according to Eddy (2004); Radostits *et al.* (2007); Constable *et al.* (2017); Elmeligy *et al.* (2021). First, insulin [Mixtard<sup>®</sup> 30 HM Vial 100 IU/ml, 1 vial contained 10 ml (1000 IU), Novo Nordisk A/S, Denmark] was administered subcutaneously and simultaneously with both IV 50% glucose solution and glucocorticoid (IM dexamethasone) and, thus, alternatively with IV 50% dextrose. The SC dose of Mixtard<sup>®</sup> 30 HM was 2 ml (200 IU)/cow q 48 h for 5 days/3 doses. Carbohydrate replacement therapy included 500 mL of 50 % IV solution of glucose/cow q 48 h for 5 days/3 doses [Glucose<sup>®</sup>, FIPCO, i.e., Fath for drug and cosmetics industry, Cairo, Egypt] alternatively with 500 ml of 50% IV dextrose solution/cow q48h for 5 days/2 doses (Dextrose 50%<sup>®</sup>, VETWIC, Cairo, Egypt). Successive 5-day treatment with propylene glycol as glucose precursor [Propane-1, 2-diol] and as carbohydrate replacement therapy (Propylene glycol, El Nasr Pharmaceutical Chemical Company,



Cairo-Egypt) via drenching route as a dose of 300 ml/cow/12 hour for 2 days, followed by 150 ml/cow/24hr for another continuous 3 days. Dexamethasone (0.04 mg/kg, i.e., 20 mg/450 kg BWt) as glucocorticoid administration (Dexamethasone®, ADWIA Co., Cairo, Egypt) was used as an IM dose of 0.04 mg/kg BWt q48h for 5 days/3 doses. Steroids (Dexamethasone) were given as a single IM injection over a 1–3 minutes' period after adequate fluid replacement therapy (50 % IV glucose solution) infusion. Vitamin B<sub>12</sub> (KENAZAL® 10 solution, KENANA) was injected via IM route as 5 ml/100 kg BWt q48h for 5 days/3 doses. The feed additives contained amino acids (methionine and lysine) named as Smartamine® M (Kemin company-USA) imported by United Bio-med Co. (Cairo, Egypt) was used as an oral dose of 15 gm d/cow/24hr for successive 60 days.

#### **Clinical examination**

A comprehensive clinical examination was performed on each animal, following Cockcroft's (2015) guidelines. Day 0 before treatment and days 7, 14, 21, and 30 after treatment. The estimation of general health indicators, such as rectal temperature, pulse rate, breathing rate, and rumen motion, was part of the clinical evaluation. Every cow's BCS was determined using a 5-point rating system (Ferguson *et al.* 1994). According to Hutjens (1996); Hulsen (2007); Burfeind *et al.* (2010); Götze *et al.* (2019), the clinical scoring system and manure scoring of dairy cattle includes estimation of RFS, appetite score, MDS, and MCS.

#### **Lipid profile indicators**

Using commercial kits (LCAT ELISA kit; Bovine LCAT ELISA Kit; MyBioSource, Inc., San Diego, USA), serum LCAT activity was measured by sandwich enzyme-linked immunosorbent assay (ELISA- Sandwich Protocol). The sandwich enzyme-linked immunosorbent assay (Sandwich ELISA, Double Antibody) was used to estimate the amount of ApoB-100 in serum. Commercial kits, such as the Bovine ApoB100 (Apolipoprotein B100) ELISA Kit (MyBioSource, Inc., San Diego, USA, were used for this assay.

Serum BHBA levels were estimated using the kinetic enzymatic method using a Spectro Ultraviolet-Vis RS spectrophotometer (Labomed, Inc., Los Angeles, CA, USA) with commercial kits (Ranbut, Ranbut, Randox Ltd, UK) provided by Randox Ltd.

The Spectro Ultraviolet-Vis RS spectrophotometer (Labomed, Inc., Los Angeles, CA, USA) was used to measure serum levels of NEFA, glucose, total cholesterol (TC), free cholesterol (FC), and AST (AST, Gamma Trade Company, Egypt) using commercial kits. Reagent kits for serum NEFA were obtained from Randox Ltd (NEFA, Randox Ltd, UK), whereas those for glucose, TC, FC, and AST were supplied by Gamma Trade Company (Cairo, Egypt). Cholesteryl ester (CE) was detected by subtracting the FC concentration from that of TC.

#### **Statistical analysis**

To evaluate all of the collected data, computer software (SPSS version 16.0, Chicago, USA) was employed. The normal distribution of all parameters was tested using Kolmogorov-Smirnov normality test. All parameters were distributed normally. The values of the data were presented as mean  $\pm$  SD. A general linear model repeated measures analysis of variance was used to examine the data from the biochemical and clinical studies, with a level of significance of  $p < 0.05$ . On certain sample days (days 0, 7, 14, 21, and 30), the significance of discrepancies between the means was evaluated. Using Pearson Correlation at  $p < 0.05$  or  $p < 0.01$ , the correlation coefficient between apoB-100 and NEFA in ketotic cattle was determined in the current study.

#### **Ethical approval**

The current study was approved by the ethical committee of the Faculty of Veterinary Medicine, Assiut University, Egypt, licensed number 06/2025/0295, and was conducted in accordance with the Egyptian bylaws and OIE animal welfare standards for animal care and use in research and education. The investigated cows were taken from the farm by obtaining informed consent from the owner.

### **Results**

#### **Therapeutic protocols**

The hormonal-steroids-carbohydrate therapeutic regimen used during the present work included mainly SC insulin (2 ml/cow/48 hour for 5 days/3 doses) and IM dexamethasone as glucocorticoids (0.04 mg/kg BWt/48 hour for 5 days/3 doses), as a single IM injection over a 1–3 minutes' period after adequate fluid replacement therapy [Carbohydrate replacement therapy, i.e., 50 % IV glucose solution] infusion. Insulin was administered subcutaneously and simultaneously with both IV 50% glucose solution and glucocorticoids (IM dexamethasone) or alternatively with IV 50% dextrose. Carbohydrate replacement therapy included IV infusion of glucose solution (50% 500 ml/cow every 48 hour for 5 days/3 doses) alternative with dextrose solution (50% 500 ml/cow every 48 hour for 5 days/2 doses). Furthermore, successive 5-day treatment with propylene glycol as glucose precursor [Propane-1, 2-diol] and as carbohydrate replacement therapy was applied via drenching route as a dose of 300 ml/cow/12 hour for 2 days followed by 150 ml/cow/24 hour for another continuous 3 days. Vitamin B<sub>12</sub> was also used via IM route as 5 ml/100 kg BWt q48h for 5 days/3 doses. The feed additives contained amino acids (methionine and lysine) named as Smartamine was used as an oral dose of 15 gm d/cow/24 hour for successive 60 days. A hormonal-steroid-carbohydrate therapeutic regimen induced high recovery rates from ketosis and very low relapse rates for ketosis. Moreover, the hormonal-steroids-carbohydrate therapeutic regimen had high efficacy as well as significantly improved the health

**Table 1.** Mean values of Body weight, temperature, pulse, respiration, and rumen movement in pretreated and post-treated ketotic cattle ( $n = 50$ ).

	Temperature (C°) (38.5–39.5) <sup>1</sup> (38–39) <sup>2</sup>	Pulse (Beat/min) (60–80) <sup>1,2</sup>	Respiration (/min) (10–30) <sup>1</sup> (15–30) <sup>2</sup>	Rumen (cycle/2 mins) (2–4) <sup>1</sup>	BCS (2.5–4) <sup>3</sup>
0D-Ket <sup>gr</sup>	38.32 ± 0.27 <sup>a</sup>	72.58 ± 3.37 <sup>a</sup>	20.14 ± 2.08 <sup>a</sup>	1.12 ± 0.29 <sup>c</sup>	3.18 ± 0.16 <sup>a</sup>
7Ds-Ket <sup>gr</sup>	38.57 ± 0.41 <sup>a</sup>	73.63 ± 2.91 <sup>a</sup>	19.85 ± 2.37 <sup>a</sup>	2.24 ± 0.36 <sup>b</sup>	3.06 ± 0.22 <sup>a</sup>
14Ds-Ket <sup>gr</sup>	38.69 ± 0.27 <sup>a</sup>	72.06 ± 3.16 <sup>a</sup>	18.33 ± 2.11 <sup>a</sup>	2.77 ± 0.44 <sup>ab</sup>	3.14 ± 0.24 <sup>a</sup>
21Ds-Ket <sup>gr</sup>	38.24 ± 0.35 <sup>a</sup>	68.19 ± 3.26 <sup>a</sup>	21.47 ± 2.39 <sup>a</sup>	3.1 ± 0.28 <sup>a</sup>	3.05 ± 0.21 <sup>a</sup>
30Ds-Ket <sup>gr</sup>	38.73 ± 0.21 <sup>a</sup>	70.06 ± 2.82 <sup>a</sup>	20.28 ± 2.77 <sup>a</sup>	3.1 ± 0.28 <sup>a</sup>	3.11 ± 0.17 <sup>a</sup>

0D-Ket<sup>gr</sup> = 0 day pretreated ketotic cows; 7Ds-Ket<sup>gr</sup> = 7 days post-treated ketotic cows; 14Ds-Ket<sup>gr</sup> = 14 days post-treated ketotic cows; 21Ds-Ket<sup>gr</sup> = 21 days post-treated ketotic cows; 30Ds-Ket<sup>gr</sup> = 30 days post-treated ketotic cows; BCS = body condition score. <sup>abc</sup>Means within the same column with different superscript letters between days 0, 7, and 14 were significantly different ( $p < 0.05$ ) in ketotic cows. Reference value according to Radostits *et al.* (2007)<sup>1</sup>; Jackson and Cockcroft (2002)<sup>2</sup>; Ferguson *et al.* (1994)<sup>3</sup>.

**Table 2.** Mean values of the appetite score, RFS, MDS, and MCS in pretreated and post-treated ketotic cattle ( $n = 50$ ).

	Appetite score	RFS (1 = flat, 5 = distended or $\leq 2/5$ ) <sup>1</sup>	MDS (2.5–3) <sup>2</sup>	MCS (2.5–3) <sup>2</sup>
0D-Ket <sup>gr</sup>	1.21 ± 0.47 <sup>c</sup>	1.91 ± 0.47 <sup>c</sup>	1.58 ± 0.54 <sup>b a</sup>	2.68 ± 0.25 <sup>a</sup>
7Ds-Ket <sup>gr</sup>	2.16 ± 0.27 <sup>b</sup>	2.64 ± 0.68 <sup>a</sup>	2.64 ± 0.27 <sup>a</sup>	2.76 ± 0.36 <sup>a</sup>
14Ds-Ket <sup>gr</sup>	2.88 ± 0.39 <sup>a</sup>	2.72 ± 0.47 <sup>a</sup>	2.82 ± 0.44 <sup>a</sup>	2.71 ± 0.32 <sup>a</sup>
21Ds-Ket <sup>gr</sup>	2.91 ± 0.11 <sup>a</sup>	2.83 ± 0.39 <sup>a</sup>	2.88 ± 0.36 <sup>a</sup>	2.76 ± 0.36 <sup>a</sup>
30Ds-Ket <sup>gr</sup>	3 <sup>a</sup>	2.81 ± 0.42 <sup>a</sup>	2.77 ± 0.29 <sup>a</sup>	2.71 ± 0.28 <sup>a</sup>

0D-Ket<sup>gr</sup> = 0 day pretreated ketotic cows; 7Ds-Ket<sup>gr</sup> = 7 days post-treated ketotic cows; 14Ds-Ket<sup>gr</sup> = 14 days post-treated ketotic cows; 21Ds-Ket<sup>gr</sup> = 21 days post-treated ketotic cows; 30Ds-Ket<sup>gr</sup> = 30 days post-treated ketotic cows; RFS = Rumen filling score; MDS = Manure digestion score; MCS = Manure condition score. <sup>abc</sup>Means within the same column with different superscript letters between days 0, 7, and 14 were significantly different ( $p < 0.05$ ) in ketotic cows. Reference values were obtained from Bramley *et al.* (2013)<sup>1</sup>; Hutjens (1996)<sup>2</sup>.

and productivity status of diseased ketotic dairy cattle by facilitating their recovery from ketosis and the dairy cattle needed a maximum 30 days follow up period in order to restore their physiological status either for clinical scoring data or for lipid profiles indices mainly serum LCAT. A significant ( $p < 0.05$ ) improvement in clinical findings and clinical scoring data (Tables 1, 2) included rumen movement, appetite score, RFS, and MDS values post-therapy as they needed only 7 days follow up following treatment to restore their reference intervals. Blood lipid profiles (Table 3) were clearly changed and improved post-therapy in days 7–30 whereas remarkable ( $p < 0.05$ ) elevations in serum LCAT activities, apoB-100 values, and cholesterol levels, and significant ( $p < 0.05$ ) declines in serum levels of serum BHBA, NEFA, and AST activities, were reported. Dairy cows needed 14 days' post-therapy to restore reference intervals of serum apoB-100, BHBA, NEFA, and AST while 30 days' post-therapy as a follow up period was needed to restore their reference values of serum LCAT. Glucose and cholesterol levels were within their reference ranges.

#### **Risk factors, clinical findings, and clinical scoring data**

Ketotic cattle are often thought to be at risk for a variety of postpartum diseases, including DA, mastitis, and/or retained placenta. Of the 50 (100%) cows, 20 (40%)

retained placenta, 15 (30%) had DA, five (10%) had mastitis, and 12 (24%) had no problems at all.

There were no significant variations in the values of temperature, pulse rate, or respiratory rate of the sick ketotic cows between days 0 (pre-treatment), 7, 14, 21, and 30. Each value was within the physiological reference range. Significant ( $p < 0.05$ ) elevations in ruminal movement values were reported in 7Ds-Ket<sup>gr</sup>, 14Ds-Ket<sup>gr</sup>, 21Ds-Ket<sup>gr</sup>, and 30Ds-Ket<sup>gr</sup> compared with their values in 0D-Ket<sup>gr</sup>, whereas they had become within their reference intervals by the 7<sup>th</sup> day post-therapy. BCS in ketotic cows showed no clear changes between days 0, 7, 14, 21, and 30; thus, BCS values were still within their reference intervals (Table 1).

There was an improvement in clinical scoring data, including appetite score, RFS, and MCS as a response to treatment as they were remarkably ( $p < 0.05$ ) raised in 7D-Ket<sup>gr</sup>, 14Ds-Ket<sup>gr</sup>, 21Ds-Ket<sup>gr</sup>, and 30Ds-Ket<sup>gr</sup> compared with their values in 0D-Ket<sup>gr</sup>, whereas they restored their reference intervals by the 7<sup>th</sup> day post-therapy. The MDS values showed no significant variations between days 0 and 30 in ketotic cattle and were still within their reference intervals (Table 2). These changes were correlated with the clear improvement in appetite and rumen movement observed in days 7–30 following therapy.

### Serum lipid profiles

Blood lipid profiles in Ketotic cattle post-therapy showed clear improvement including elevations of blood levels of LCAT, apoB-100, glucose, and TC as well as reductions of blood concentrations of BHBA, NEFA, and AST were noticed in days 7–30 compared with their pretreatment values in day 0 (Table 3).

The serum activities of LCAT were highly changed after therapy in ketotic dairy cattle as they were significantly ( $p < 0.05$ ) raised in days 7–30 following treatment compared with those on day 0 (Pretreatment). Nevertheless, they remained below the reference values even after treatment (Days 7–21). Thirty days post-therapy as a follow-up period was needed by recovered dairy cows to restore their reference values of serum LCAT (Table 3).

Ketotic cattle had remarkably ( $p < 0.05$ ) lower concentrations of serum apoB-100 in 0D-Ket<sup>gr</sup> than their post-therapeutic values in days 7–30 and their reference intervals. Clear improvement in serum apoB-100 levels was observed following therapy, whereas the reference values were reached on day 14 posttherapy (Table 3).

The current study reported no significant variations in serum glucose levels in ketotic cows either pretherapy (Day 0) or posttherapy (Days 7, 14, 21, and 30). Moreover, the blood glucose levels of the investigated dairy cows were within the reference intervals (Table 3).

The serum concentrations of BHBA were significantly ( $p < 0.05$ ) higher in ketotic cows before therapy (0D-Ket<sup>gr</sup>) than in 7Ds-Ket<sup>gr</sup>, 14Ds-Ket<sup>gr</sup>, 21Ds-Ket<sup>gr</sup>, and 30Ds-Ket<sup>gr</sup>. Later on, a significant ( $p < 0.05$ ) drop in serum BHBA values was reported as a response

to therapy in days 7–30 whereas they restored their reference values by the 14<sup>th</sup> day. Serum BHBA levels reached their lowest levels in post-therapeutic ketotic cows on day 21 (21Ds-Ket<sup>gr</sup>) (Table 3).

Post-therapeutic ketotic cattle in days 7–30 had significantly ( $p < 0.05$ ) lower values of serum NEFA levels than those on day 0 (0D-Ket<sup>gr</sup>). The serum NEFA concentrations in the investigated dairy cattle reached their reference ranges on day 14 following therapy. The serum NEFA levels reached their lowest levels in post-therapeutic ketotic cows on day 21 after treatment (Table 3).

The present study revealed significantly ( $p < 0.05$ ) higher serum levels of cholesterol, i.e., TC, CE, and FC, following therapy in ketotic cows on days 7, 14, 21, and 30 compared with their values in the 0D-Ket<sup>gr</sup>. However, serum cholesterol concentrations were within reference ranges throughout the present study (Table 3).

The reported results showed severe reductions in serum enzyme activities of AST in post-therapeutic ketotic cattle, whereas their values in 7Ds-Ket<sup>gr</sup>, 14Ds-Ket<sup>gr</sup>, 21Ds-Ket<sup>gr</sup>, and 30Ds-Ket<sup>gr</sup> were remarkably ( $p < 0.05$ ) lower than those in pretherapeutic ketotic cattle (0D-Ket<sup>gr</sup>). Dairy cows needed a 14-day follow-up period (Post-therapy) to restore their reference intervals of serum AST (Table 3).

The study reported a significant negative correlation between serum NEFA and apoB-100 levels in ketotic cattle either pre- or post-therapy where the remarkable elevation of NEFA levels on day 0 was associated with clear declines in serum apoB-100 levels. In contrast, the significant reduction in serum NEFA levels after treatment (Days 7–30) was correlated with a significant

**Table 3.** Mean values of serum lipid profiles in pre-treated and post-treated ketotic cattle (n=50)

	LCAT (U) (929–1059) <sup>1</sup> (898.39 ± 120.89) <sup>2</sup>	ApoB-100 (g/l) (0.2–0.32) <sup>3</sup>	Glucose (mmol/l) (4.50–6.71) <sup>4</sup> (1.9–3.8) <sup>5,6</sup>	BHBA (mmol/l) (0.35–0.47) <sup>5</sup> (0.41 ± 0.03) <sup>7</sup>	NEFA (mmol/l) (0.21– 0.62) <sup>8</sup>	TC (mmol/l) (1–5.6) <sup>5</sup> (2.07–3.11) <sup>7</sup>	CE	FC	AST (U/l) (78–132) <sup>5</sup> (105 ± 27) <sup>7</sup>
0D-Ket <sup>gr</sup>	247.69 ± 33.93 <sup>c</sup>	0.09 ± 0.02 <sup>d</sup>	5.55 ± 0.78 <sup>a</sup>	2.33 ± 0.98 <sup>a</sup>	1.72 ± 0.55 <sup>a</sup>	5.26 ± 1.76 <sup>b</sup>	4.18 ± 1.43 <sup>b</sup>	1.09 ± 0.36 <sup>c</sup>	340.44 ± 22.07 <sup>a</sup>
7Ds-Ket <sup>gr</sup>	338.33 ± 15.74 <sup>d</sup>	0.15 ± 0.07 <sup>c</sup>	5.50 ± 0.88 <sup>a</sup>	0.73 ± 0.14 <sup>b</sup>	0.84 ± 0.13 <sup>b</sup>	6.23 ± 1.60 <sup>b</sup>	4.93 ± 1.42 <sup>b</sup>	1.31 ± 0.23 <sup>b</sup>	228.11 ± 19.42 <sup>b</sup>
14Ds-Ket <sup>gr</sup>	647.47 ± 25.52 <sup>c</sup>	0.22 ± 0.03 <sup>b</sup>	5.69 ± 0.77 <sup>a</sup>	0.36 ± 0.16 <sup>c</sup>	0.41 ± 0.08 <sup>c</sup>	11.20 ± 2.70 <sup>a</sup>	9.01 ± 2.21 <sup>a</sup>	2.15 ± 0.62 <sup>a</sup>	108.78 ± 26.47 <sup>c</sup>
21Ds-Ket <sup>gr</sup>	812.67 ± 17.08 <sup>b</sup>	0.37 ± 0.06 <sup>a</sup>	5.22 ± 0.48 <sup>a</sup>	0.29 ± 0.12 <sup>c</sup>	0.22 ± 0.08 <sup>d</sup>	12.45 ± 2.44 <sup>a</sup>	9.81 ± 2.04 <sup>a</sup>	2.75 ± 0.71 <sup>a</sup>	66.79 ± 11.64 <sup>d</sup>
30Ds-Ket <sup>gr</sup>	948.28 ± 22.19 <sup>a</sup>	0.32 ± 0.05 <sup>a</sup>	5.44 ± 0.53 <sup>a</sup>	0.31 ± 0.13 <sup>c</sup>	0.21 ± 0.07 <sup>d</sup>	12.29 ± 2.12 <sup>a</sup>	9.73 ± 1.65 <sup>a</sup>	2.61 ± 0.82 <sup>a</sup>	72.43 ± 14.73 <sup>d</sup>

0D-Ket<sup>gr</sup> = 0 day pretreated ketotic cows; 7Ds-Ket<sup>gr</sup> = 7 days post-treated ketotic cows; 14Ds-Ket<sup>gr</sup> = 14 days post-treated ketotic cows; 21Ds-Ket<sup>gr</sup> = 21 days post-treated ketotic cows; 30Ds-Ket<sup>gr</sup> = 30 days post-treated ketotic cows; LCAT = Lecithin cholesterol acyltransferase; ApoB-100 = Apolipoprotein B-100; BHBA =  $\beta$ -hydroxy butyric acid; NEFA = Nonesterified fatty acid; TC = Total cholesterol; CE = cholesterol ester; FC = Free cholesterol; AST = Aspartate Aminotransferase. <sup>a–c</sup>Means within the same column with different superscript letters between days 0, 7, and 14 were significantly different ( $p < 0.05$ ) in ketotic cows. Reference value according to Nakagawa and Katoh (1998)<sup>1</sup>; Khalphallah et al. (2018a)<sup>2</sup>; Itoh et al. (1997)<sup>3</sup>; Zadnik (2003)<sup>4</sup>; Radostits et al. (2000)<sup>5</sup>; Jackson and Cockcroft (2002)<sup>6</sup>; Kaneko et al. (2008)<sup>7</sup>; Oikawa and Katoh (2002)<sup>8</sup>.

**Table 4.** Pearson's correlation coefficient between serum apoB-100 and NEFA levels in ketotic cows.

Variables		
Apo-B100 (g/l)	NEFA (mmol/l)	Correlation (r)= ?0.641** p value = 0.003

ApoB-100 = Apolipoprotein B-100; NEFA = Nonesterified fatty acid. \*Significant (two-tailed)  $p < 0.05$ . \*\*Significant (two-tailed)  $p < 0.01$ .

increase in serum ApoB-100 levels ( $r = ?0.642$ ,  $p < 0.05$ ; Sig. 2-tailed = 0.0033) (Table 4).

## Discussion

### Therapeutic protocols

Most of the time, cows with type I ketosis reacted favorably to various ketosis interventions. All they required was an extra boost to get back on track in their struggle to meet energy needs (Holtenius and Holtenius, 1996; Oetzel, 2007; Elmeligy *et al.*, 2021). Tufani *et al.* (2011) revealed that the importance of hormonal therapy, i.e., dexamethasone or nandrolone, in association with parenteral administration of glucose and B-complex ( $B_1$ ,  $B_2$ , and  $B_3$ ) induced a very low rate of recurrence and a great recovery rate. McArt *et al.* (2011) added that oral propylene glycol therapy proved a highly successful way to prevent and treat ketosis in dairy cows in addition to clearly increasing milk production. Regarding the present study, the hormonal-steroids-carbohydrate therapeutic regimen used during the present work included mainly SC insulin and glucocorticoids as dexamethasone, as a single IM injection over a 1–3 minutes' period after adequate fluid replacement therapy [Carbohydrate replacement therapy, i.e., 50 % IV glucose solution] infusion. Insulin was administered subcutaneously and simultaneously with both IV 50% glucose solution and glucocorticoids (IM dexamethasone) or alternatively with IV 50% dextrose. Carbohydrate replacement therapy included a 50% IV alternative with a dextrose solution of 50% IV. Furthermore, successive 5-day treatment with propylene glycol as glucose precursor [Propane-1, 2-diol] was used as carbohydrate replacement therapy. IM injection of Vitamin  $B_{12}$  as well as the use of the feed additives (Smartamine) were also used. A hormonal-steroid-carbohydrate therapeutic regimen induced high recovery rates from ketosis and very low relapse rates for ketosis. Moreover, exposure of diseased ketotic dairy cattle for a specific therapeutic strategy had a great impact on their recovery from ketosis and improving their healthy and productivity status whereas this therapeutic regimen approved high efficacy and the dairy cattle needed maximum of 30 days follow up period in an order to restore their physiological status either for clinical scoring data or for lipid profiles indices mainly serum LCAT. 7 days were needed through which dairy cows could restore

their clinical score data. Dairy cows needed 14 days' post-therapy to restore reference intervals of serum apoB-100, BHBA, NEFA, and AST while 30 days' post-therapy as a follow up period was needed to restore their reference values of serum LCAT. Furthermore, our current research has shown the great effectiveness of treatment in ketotic cows receiving 200 IU SC of insulin in conjunction with 500 mL of IV glucose 50% and 20 mg/450 kg of dexamethasone once daily every other day. This regimen should be followed for consecutive five days, alternating with 50% dextrose. According to Sakai *et al.* (1993), ketotic cows given only intravenous glucose daily recovered more slowly than those given intravenous glucose and subcutaneous insulin injections for four days. Additionally, the responses of cows with primary ketosis to treatment with corticosteroids and IV glucose were shown to be superior than therapy with corticosteroids or IV glucose alone, with fewer relapses. These findings were corroborated by studies conducted by Radostits *et al.* (2007); Constable *et al.* (2017). According to other research, insulin promoted hepatic gluconeogenesis, inhibited lipolysis, and improved cellular absorption of glucose. insulin, when given in conjunction with glucose or corticosteroids, might be effective in treating early-onset ketosis that was not responded to glucose or corticosteroids (Djokovic *et al.*, 2007; Radostits *et al.*, 2007; Elmeligy *et al.*, 2021), and thus IV dextrose infusion must always be seen as a combined insulin and glucose therapy (Djokovic *et al.*, 2007; Constable *et al.*, 2017). On the other hand, the importance of the injection of exogenous insulin in ketosis therapy that was reported in the current study was supported by Hayirli *et al.* (2002); Hayirli (2006). According to Hayirli (2006), restoring nutritional status could alleviate the lowered responsiveness and sensitivity of extrahepatic tissues to insulin that resulted from a negative energy balance. This would increase the effectiveness of exogenous insulin in treating hepatic lipidosis and ketosis, preserving health and enhancing performance. Radostits *et al.* (2007); Grünberg *et al.* (2011) stated that IV infusion of glucose solution 50% (500 ml) led to elevated insulin secretion and reduced glucagon release as well as transient hyperglycemia with lowered plasma concentration of NEFA. Compared to therapy with corticosteroids or glucose alone, the response of cows with primary ketosis to corticosteroid treatment with IV glucose infusion was superior, with fewer relapses (Shpigel, 1996). Because it reduced the synthesis of ketone bodies through the use of Acetyl-CoA generated through fatty acid oxidation, and because it increased the availability of glucose precursors in the liver, glucocorticoid treatment was also beneficial. Usually, a single dose was administered when the injection could be repeated. The repeated glucocorticoid therapy had one disadvantage including reduced appetite and milk yield. Ketotic cattle treated with glucocorticoids were



less vulnerable to relapses than were those treated with IV glucose therapy alone, although relapses were still reporting (Andrews *et al.*, 2004).

**Risk factors, clinical findings, and clinical scoring data:**

Ketosis of both CK and SCK is a prevalent metabolic disease during the periparturient stage because it is associated with several concomitant diseases and financial losses (Dohoo *et al.*, 1984; McArt *et al.*, 2012; Khalphallah *et al.*, 2018a; Elmeligy *et al.*, 2021). Iwersen *et al.* (2009) reported that SCK was frequently hidden in the absence of clinical findings and that the actual prevalence in dairy herds was undervalued. Ketosis negatively affected the reproductive, productive, and healthy performance of cows. According to the current study, ketotic cattle were typically regarded as risk factors for a variety of postpartum illnesses, including mastitis, retained placenta, and/or DA. Previous reports have mentioned that subclinical or clinical ketosis is one of the outcomes of negative energy balance (Andersson, 1984; Andersson, 1988). The dairy sector suffered economic losses due to ketosis because it reduced milk output, reproductive efficiency, increased involuntary culling, and increased treatment expenses (Miettinen, 1990; Miettinen *et al.*, 1991; Detilleux *et al.*, 1994; Elmeligy *et al.*, 2021). On the other hand, dairy herds with ketosis, particularly early lactation cattle, were at higher risk of DA and retained fetal membranes (Curtis *et al.*, 1985; Duffield and Bagg, 2002; Melendez *et al.*, 2003) and were more prospective to have reduced fertility and longer periods between calving and pregnancy (Opsome *et al.*, 2000; Gillund *et al.*, 2001). The clinical findings in diseased ketotic cattle throughout the current study, including temperature, pulse rate, and respiratory rate, revealed no significant variations between days 0 (pre-treatment), 7, 14, 21, and 30. All clinical parameters were within the physiological reference range reported by Jackson and Cockcroft (2002); Radostits *et al.* (2007). Significant elevations in ruminal movement values were described in post-treated cows compared with their values in day zero, whereas they had become within their reference intervals reported by Radostits *et al.* (2007) in the 7<sup>th</sup> day post-therapy. These findings agreed with Tufani *et al.* (2011), who noted a typical, sweetish smell of urine, milk, and breath in the majority of the cases. Other previous reports mentioned abrupt decreases in milk production and an unwillingness to consume concentrate feeds and grains as clinical indications of ketosis. The breath and milk had a pleasant acetone odor. Nervous signs in cows have been also reported (Moore and Ishler 1997; Constable *et al.*, 2017).

BCS in ketotic cows during the current study revealed no clear variations between days 0, 7, 14, 21, and 30; thus, BCS values were still within the reference values mentioned by Ferguson *et al.* (1994). These results were relatively consistent with Khalphallah *et al.* (2018a), who reported no remarkable changes in BCS values between diseased and healthy cattle either in close-

up or postfresh periods. Edwards and Tozer (2004) revealed an association between ketosis in dairy cows and a gradual decline in their BCS over several days or weeks. Previous studies have stated that most dairy cows with higher BCS prepartum faced a negative energy balance in early lactation due to their lower postpartum feed intake, which left them in a negative energy balance for a longer period than cattle with normal or low BCS (Hayirli *et al.*, 2002). Dyk (1995) mentioned cows with excess BCS at parturition were more likely to experience ketosis. It is possible that the higher reported incidence of LDA in cows with higher BCS at parturition was due to the higher prevalence of ketosis in these cows. Moreover, greater amounts of dry matter intake (DMI) reduction had been reported during the transition period of dairy cows compared with heifers (Hayirli *et al.*, 2002), suggesting greater energy disturbances, which might explain their higher susceptibility to postures metabolic diseases (Curtis *et al.*, 1985).

Burfeind *et al.* (2010) revealed that RFS is related to feed intake, suggesting that it should be estimated consistently at the same time of day to detect changes in dry matter intake. Moreover, Götze *et al.* (2019) reported that the RF score is a sensitive tool for monitoring feed intake during the first week after giving birth. It was only appropriate for the identification of animals at risk who had significantly decreased their feed intake over a few days due to poor prepartum response. The ketotic cows in the current study showed a clear improvement in their clinical scoring data, including appetite score, RFS, and MCS as a response to treatment as they were remarkably raised following therapy in 7D-Ket<sup>gr</sup>, 14Ds-Ket<sup>gr</sup>, 21Ds-Ket<sup>gr</sup>, and 30Ds-Ket<sup>gr</sup> compared with their values in 0D-Ket<sup>gr</sup> whereas they restored their reference intervals mentioned by Hutjens (1996); Bramley *et al.* (2013) by the 7<sup>th</sup> day post-therapy. MDS values showed no significant variations between days 0 and 30 in ketotic cattle and were still within the reference intervals mentioned by Hutjens (1996). These changes were correlated with the clear improvement in appetite and rumen movement observed in days 7–30 following therapy. These results were supported by Khalphallah *et al.* (2016a); Götze *et al.* (2019). Khalphallah *et al.* (2016a) reported a clear improvement in RFS in dairy cows following surgical operation in recovered DA cows either with SCK or with CK, particularly on day 30. MDS was greatly improved (Declined undigested feed particles) in the recovered DA cows with either SCK or CK, particularly on day 30. The ideal manure score was three with normal pat form, as described by Allen and Beede (1996); Hutjens (1996); Robert (2013). Khalphallah *et al.* (2016a) reported that MCS ( $3.40 \pm 0.7$ ) was clearly increased (Optimal consistency) in all ketotic cattle with DA on postoperative day 30, which was indicated by good consistency of the feces, reflecting restoration of normal digestion in the cow. Khalphallah



*et al.* (2016a) reported that the improvement in MDS (Reduced undigested feed particles) in cows with DA with ketosis required a longer period, which extended to day 30 postoperatively.

#### **Serum lipid profiles**

Blood lipid profiles in Ketotic cattle post-therapy showed clear improvement, including elevations of blood levels of LCAT, apoB-100, glucose, and TC, as well as reductions of blood concentrations of BHBA, NEFA, and AST were noticed in days 7–30 comparing with their values pretherapy in day 0. These significant changes in blood lipid profiles, LCAT, and apoB-100 revealed the improvement of animal health as biosynthesis, metabolism, and transport of these parameters were conducted within the liver and by enzymes synthesized in the liver. This interpretation was supported by Drackley (1999), who added that diseases of transition period like ketosis, retained fetal membranes, and DA were mainly correlated with fatty liver. Furthermore, Jonas (1998) added that LCAT was mainly synthesized by the liver and, in plasma, catalyzed the transesterification of FC with lecithin, i.e., phosphatidylcholine, and resulted in CE. Therefore, LCAT activity, together with the CE concentration, was similarly reduced in cows with ketosis and LDA (Nakagawa and Katoh, 1998). Ketotic cows showed the same changes in serum LCAT during the present study; accordingly, serum activities of LCAT were highly changed after therapy in ketotic dairy cattle as they were significantly increased in days 7–30 following treatment compared with their zero-day pretreatment values. However, they were still lower than the reference values reported by Nakagawa and Katoh (1998); Khalphallah *et al.* (2018a) even after therapy (days 7–21). A 30-day post-therapy as a follow-up period was needed by recovered dairy cows to restore their reference values of serum. Khalphallah *et al.* (2016b) reported a significant increase in serum LCAT activity in ketotic cows with DA after surgery on day 30 compared with that on day 0 before surgery. Other reports mentioned a reduction in serum LCAT activity in diseased cattle, whether in the close-up or post-fresh period, compared with their values in control cattle (Khalphallah *et al.*, 2018a). The decrease in LCAT levels was reported before the diagnosis of ketosis, clinical hypocalcemia, or milk fever (Nakagawa and Katoh 2000). Some studies referred to the use of serum LCAT activities as a diagnostic marker for fatty liver-related disorders, including DA and ketosis, as they were more diagnostic than blood apoB-100 levels because the reduction in serum LCAT activities usually preceded clinical signs, as well as their activities were not altered during the peripartum period, at least in some healthy cattle (Nakagawa and Katoh, 1998).

ApoB-100 concentrations were decreased in dairy cows with hepatic lipidosis-related diseases, such as ketosis, retained fetal membranes, and LDA (Oikawa *et al.*, 1997). The similarly reduced rates of apoB-100

suggested that this drop in apoB-100 concentrations in milk fever, ketosis, downer cow syndrome, retained placenta, and LDA were predominantly caused by fatty liver, confirming the theory that fatty liver was the origin of these diseases (Morrow, 1976; Morrow *et al.*, 1979; Gerloff *et al.*, 1986; Herdt, 1988). These results supported the current results in which ketotic cattle had remarkably lower concentrations of serum apoB-100 in 0D-Ket<sup>sr</sup> than their post-therapeutic values in days 7–30 and their reference intervals. Clear improvement in serum apoB-100 levels was observed following therapy, whereas their reference values mentioned by Itoh *et al.* (1997) were reached on day 14 post-therapy. The previous literature revealed that serum levels of apoB-100 showed significant changes in ketotic cows with DA as their blood concentrations were significantly decreased in the diseased status on day 0 compared with their recovered status after surgery on day 30. Their levels on day 30 were within the reference ranges (Khalphallah *et al.*, 2016b).

The current results reported no significant variations in serum glucose concentrations in ketotic dairy cattle either pretherapy (Day 0) or post-therapy (Days 7, 14, 21, and 30). Moreover, blood glucose levels in the investigated dairy cows were within the reference intervals reported by Radostits *et al.* (2000); Jackson and Cockcroft (2002); Zadnik (2003). The blood glucose, TC, triglycerides, and albumin concentrations were greatly lower during prepartum, i.e., 1–15 days, in puerperal ketotic cattle than in healthy cows (Đoković *et al.*, 2012). Furthermore, a strong association between reduced blood glucose and an elevated incidence of ketosis has been reported in post-fresh dairy cows (Radostits *et al.*, 2007). Cows with SCK had lower blood glucose concentrations than those without ketosis (Sakha *et al.*, 2007; Tehrani-Sharif *et al.*, 2012); however, in dairy cattle with SCK ketonemia, remarkable hypoglycemia was reported (Gröhn *et al.*, 1983). Moreover, Moyes *et al.* (2009) reported an association between higher levels of prepartum glucose in cattle and an increased incidence of postpartum diseases. Khalphallah *et al.* (2018a) mentioned that blood glucose levels in close-up or post-fresh healthy cows, as well as cattle with postpartum diseases (Ketosis, DA, and retained placenta), were still within the physiological reference values; however, blood concentrations of glucose were significantly lower in the diseased cows with postpartum diseases than in healthy cattle in the close-up period. These significant changes were absent in the post-fresh period between healthy cows and those with postpartum diseases. In contrast, Khalphallah *et al.* (2016b) reported that ketotic cows with DA showed no significant changes in blood glucose levels, and their values were within the reference ranges before or after treatment.

Serum BHBA levels were negatively correlated with serum glucose levels in SCK cattle (Sakha *et al.*, 2007). Plasma glucose values and plasma BHBA

concentrations were used together to categorize cattle as having a low energy state (Whitaker *et al.*, 1983). According to the present study, serum BHBA concentrations were significantly higher in ketotic cows before therapy (0D-Ket<sup>tr</sup>) than their values in days 7–30 (Post-therapy). Later on, a significant drop in serum BHBA values was reported as a response to therapy in days 7–30 whereas they restored their reference values were restored, as reported by Radostits *et al.* (2000); Kaneko *et al.* (2008) in the 14<sup>th</sup> day (21Ds-Ket<sup>tr</sup>). Serum BHBA levels reached their lowest levels in post-therapeutic ketotic cows on day 21 (21Ds-Ket<sup>tr</sup>). In contrast, Khalphallah *et al.* (2016b) stated that although there was a significant reduction in serum BHBA in ketotic cows treated with DA on days 7 and 30 compared with their values on day 0, their levels were still higher than their reported reference values, which were mentioned by Radostits *et al.* (2000); Kaneko *et al.* (2008). In contrast, the DA cattle showed no remarkable changes in serum BHBA either pre-treatment or post-treatment and were still within the reference range (Khalphallah *et al.*, 2016b). During 1–15 days prepartum in puerperal ketotic cows, significant increases in hepatic fat content and blood BHBA concentrations were reported (Đoković *et al.*, 2012). Furthermore, Plasma NEFA was a more accurate indicator of energy balance than serum BHBA concentrations as serum BHBA was usually influenced by glucose and energy balance. Elevated levels of serum BHBA are related to decreased milk yield, elevated clinical ketosis, and LDA, and decreased fertility (Herd *et al.*, 2001).

Holtenius and Holtenius (1996); Herdt (2000) stated that the diagnostic test of type I ketosis was an estimation of blood levels of postures BHBA, while the diagnostic test of type II ketosis was an estimation of blood levels of prepartum or refresh NEFA and their prevention strategy was based on improvement of nutrition and management just before calving for type II ketosis and in the early lactation for type I ketosis. Serum NEFA normally increases in the last few days before the expected delivery or calving in pregnant cattle (Oetzel, 2003). Plasma NEFA concentrations increased significantly after calving, indicating that the cow felt a negative energy balance. This negative energy balance was also demonstrated by the decrease in BCS and body weight after calving (3 weeks after calving). A severe decrease in body weight and BCS after calving may indicate dramatically increased plasma NEFA concentrations (Tharwat *et al.*, 2012). Throughout the present study, the post-therapeutic ketotic cattle in days 7–30 had significantly lower serum NEFA values than those on day 0 (0D-Ket<sup>tr</sup>). The serum NEFA concentrations in the investigated dairy cattle reached the reference ranges reported by Oikawa and Katoh (2002) on day 14 following therapy. Serum NEFA levels were lowest in post-therapeutic ketotic cows on day 21 after treatment. On the other

hand, the reported results showed severe reductions in the serum enzyme activities of AST in post-therapeutic ketotic cattle, whereas their values in 7Ds-Ket<sup>tr</sup>, 14Ds-Ket<sup>tr</sup>, 21Ds-Ket<sup>tr</sup>, and 30Ds-Ket<sup>tr</sup> were remarkably lower than those in pretherapeutic ketotic cattle (Day 0). Dairy cows needed a 14-day follow-up period (Post-therapy) to restore their reference intervals of serum AST, as mentioned by Radostits *et al.* (2000); Kaneko *et al.* (2008). These findings were supported by Đoković *et al.* (2012), who reported that puerperal ketotic cows showed significant elevations in both blood NEFA concentrations and AST activities during 1–15 days before the expected calving. Khalphallah *et al.* (2016b) added that DA cattle with CK or SCK had higher serum NEFA and AST levels on day 0 before surgery than on days 7 and 30 post-surgery. Khalphallah *et al.* (2018a) reported no significant differences in serum NEFA levels between healthy cattle and those with postpartum disease during the close-up period. Serum. The serum NEFA levels were still within their reference intervals. Furthermore, Cameron *et al.* (1998) stated that because milking cows were supposed to have a negative energy balance, blood NEFA concentrations were high following calving, although this fluctuated and was difficult to measure. Consequently, compared with NEFA concentration, postpartum serum BHBA was a more sensitive and specific test.

Fatty liver in dairy cows usually preceded ketosis, whereas fatty liver was reported in almost all natural states of bovine ketosis (Shaw, 1956; Oikawa *et al.*, 1997; Tharwat *et al.*, 2012). As reported in fatty liver cases, overfeeding was epidemiologically suggested as a significant contributor to ketosis (Markusfeld, 1985). Moreover, higher levels of blood NEFA (Kronfeld, 1965) as well as lower CE and PL levels were reported in ketotic cows (Nakagawa and Katoh, 1998). The present study revealed significantly higher serum levels of cholesterol, i.e., TC, CE, and FC, following therapy in ketotic cows on days 7, 14, 21, and 30 compared with their values in the 0D-Ket<sup>tr</sup>. However, serum cholesterol concentrations were within the reference ranges mentioned by Radostits *et al.* (2000); Kaneko *et al.* (2008) throughout the present study. Alterations in serum cholesterol concentrations are usually associated with both fat intake (Duske *et al.*, 2009) and feed intake (Janovick Guretzky *et al.*, 2006). Khalphallah *et al.* (2016b) added that TC concentrations were clearly altered in ketotic cattle with DA before and after surgical correction of DA, where they were significantly elevated after surgery, particularly on day 30 compared with their values on day 0. Lower blood CE and PL concentrations have been reported in cows with ketosis (Nakagawa and Katoh, 1998). Decreased cholesterol concentrations have been demonstrated in hepatic lipidosis (Bobe *et al.*, 2004; Van den Top *et al.*, 2005) and metabolic diseases (Itoh *et al.*, 1998), and have been associated with feed intake (Janovick Guretzky *et al.*, 2006).

The study reported a significant negative correlation between NEFA and apoB-100 in ketotic cattle either pre- or post-therapy, whereas remarkable elevations of NEFA levels on day 0 were consistent with significant declines in serum apoB-100. In contrast, the significant reduction in serum NEFA levels reported after treatment (Days 7–30) was correlated with a higher increase in serum ApoB-100 levels. These findings were supported by Khalphallah *et al.*, (2016b), who reported the same relationship between serum NEFA concentration and serum apoB-100 values in DA with clinical ketosis either before or after surgery. This might be due to an elevation in serum NEFA levels associated with the development of fatty liver as well as the reduced ability of the liver to secrete apoB-100 as a very low-density lipoprotein to the circulation, which eventually causes a reduction in serum apoB-100. On the other hand, previous studies have mentioned that VLDL formation depends on apolipoprotein and cholesterol. Accordingly, apolipoprotein supply was limited in cows; therefore, the ruminant liver had a limited or weak capacity to secrete VLDL (Grummer, 1993), and overproduction of TGs then accumulated in the hepatocyte cytosol, producing fatty liver (Cavestany *et al.*, 2005; Ingvarsten *et al.*, 2003).

### Conclusion

The current study concluded the diagnostic significance of the clinical scoring system [Appetite score, RFS, MDS, and MCS], and serum lipid profiles, mainly LCAT and ApoB-100, as recovery indicators in ketotic dairy cattle using hormonal-steroids-carbohydrates therapeutic protocol. Exposure of diseased ketotic dairy cattle to this specific therapeutic strategy had a great impact on their recovery from ketosis and improved their health and productivity status. Therefore, the ketotic dairy cattle needed a maximum 30 days follow-up period post-therapy to restore their physiological status for clinical scoring data [7 days needed] and most serum lipid indices [14 days needed, i.e., serum ApoB-100, BHBA, NEFA, glucose, cholesterol, and AST]. A 30-day post-therapy as a follow-up period was needed to restore their reference values of serum LCAT. The results of this study indicated that serum LCAT activity and ApoB-100 were the most sensitive indicators of the efficacy of therapy and the restoration of the physiological status. This could be seen by an obvious improvement in serum lipid profiles, mainly LCAT and apoB-100, and clinical findings as well as clinical coring data. The study identified a significant negative correlation between NEFA and apoB-100 in ketotic cattle.

### Acknowledgment

Not applicable.

### Funding

The study did not receive any external funding.

### Authors' contributions

A.K., A.H.A., E.E., R.H.M., and K.A.K. prepared the study conception and design. A.K., E.E., MS, W.M.E., A.A., K.A.K., and S.A.M. conducted the field study and cattle examination. A.K., E.A.A., MS, R.H.M., S.A.M., E.S.A., and E.E. collected laboratory samples and conducted laboratory analyses. K.A.K., A.H.A., MS, R.H.M., A.I.A., and A.K. manipulated and statistically analyzed the data. A.A., A.H.A., E.A.A., E.S.A., and A.I.A. performed the analysis, data curation, and interpretation of the data. A.I.A., W.M.E., A.K., A.H.A., and E.E. drafted the manuscript. E.E., E.S.A., A.H.A., E.A.A., and A.K. performed final writing, critical review, and revision. All authors have read and approved the final manuscript.

### Conflict of interest

The authors declare that they have no conflicts of interest.

### Data availability

The datasets used and/or analyzed during the current study are available from the corresponding author upon reasonable request. Data was also available after publishing in this journal.

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